

1 **Do infections with parasites and exposure to pollution affect susceptibility to**
2 **predation in johnny darters (*Etheostoma nigrum* Rafinesque, 1820)?**

3
4 **Est-ce que les infections parasitaires et l'exposition à la pollution affectent la**
5 **susceptibilité à la prédation chez raseux-de-terre noir (*Etheostoma nigrum***
6 **Rafinesque, 1820)?**

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22 **Abstract:** Johnny darters (*Etheostoma nigrum* Rafinesque, 1820) were collected from
23 five localities in the St. Lawrence River in southwestern Quebec to test the effects of
24 natural parasite infections and exposure *in situ* to pollution on their anti-predator
25 behaviour. Three measures of antipredator behaviour were made: capture time, capture
26 order and flight initiation distance. Capture time, the time taken to catch individual fish,
27 was used as a proxy for ability to evade predation, capture order was the order in which
28 fish kept in a single tank were taken from the tank, and flight initiation distance was the
29 distance at which the fish moved when approached by a model predator. Only capture
30 time showed a significant correlation with parasitism or pollution status. A non-
31 parametric permutational multivariate ANOVA showed that capture time was
32 significantly correlated with capture location and the abundance of the brain-encysting
33 trematode *Ornithodiplostomum* sp. Infection with *Ornithodiplostomum* sp. may have led
34 to an increase in activity, which would be maladaptive for this cryptic, benthic fish under
35 natural predation conditions. Pollution may have an indirect effect on predator
36 susceptibility in johnny darters, by reducing the abundance of a behaviour-modifying
37 parasite.

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39 **Résumé:** Des raseux-de-terre noir (*Etheostoma nigrum* Rafinesque, 1820) ont été
40 récoltés à cinq stations le long du fleuve Saint-Laurent (sud-ouest du Québec), dans le but
41 d'examiner l'effet conjoint de l'infection parasitaire naturelle et de l'exposition à des
42 niveaux réalistes de pollution sur le comportement anti-prédateur des poissons. Trois
43 mesures de comportement anti-prédateur ont été réalisées : 1) le temps de capture, soit le
44 temps nécessaire à la capture d'un poisson donné à l'aide d'un filet, 2) l'ordre de capture,

45 soit l'ordre dans lequel les poissons d'un bassin étaient capturés et 3) la distance
46 d'amorce de la fuite, soit la distance à partir de laquelle un poisson se déplaçait lorsque
47 approché par un prédateur factice. Seul le temps de capture a montré une corrélation
48 significative avec le parasitisme ou le niveau de pollution du milieu d'origine. Cette
49 mesure a donc été utilisée comme témoin de la capacité d'un poisson d'échapper à un
50 prédateur. Une analyse de variance non-paramétrique multidimensionnelle avec tests par
51 permutations a montré que le temps de capture était significativement corrélé à la station
52 d'échantillonnage et à l'abondance d'*Ornithodiplostomum* sp., un trématode enkysté dans
53 le cerveau. L'infection par *Ornithodiplostomum* sp. pourrait conduire à une hyperactivité,
54 un comportement potentiellement mésadapté dans des conditions de prédation naturelle
55 pour ce poisson benthique au mœurs cryptiques. Par ailleurs, la pollution pourrait avoir
56 des effets négatifs indirects sur la susceptibilité aux prédateurs chez le raseux-de-terre
57 noir, en réduisant l'infection par un parasite capable de modifier le comportement de son
58 hôte.

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68 **Introduction**

69 Parasites and pollution are common stressors in aquatic ecosystems, and both may
70 affect fish behaviour. The effects of parasites on fish behaviour may be adaptive or
71 simply reflect pathology caused by the parasite (Poulin 1995; Barber et al. 2000; Moore
72 2002; Barber and Rushbrook 2008). Larval stages of trophically transmitted parasites
73 commonly manipulate behaviour in fish intermediate hosts. They may increase their
74 transmission success by modifying their host's behaviour to increase its susceptibility to
75 predation by the downstream host in the life cycle. Such changes include increased
76 flashing and surfacing, reduced schooling, and altered habitat use. Pathological changes
77 include lethargy, increased or decreased foraging activity, and altered social interactions
78 (reviewed in Barber et al. 2000; Moore 2002; Barber and Rushbrook 2008).

79 Chronic exposure to sublethal levels of pollutants can also cause changes in fish
80 anti-predator behaviour. Studies of direct effects of exposure to metals, organic chemicals
81 and pesticides show that exposed fish may be more susceptible to predation (reviewed by
82 Atchison et al. 1987; Clotfelter and Levering 2004; Scott and Sloman 2004), because of
83 impaired physiological performance, sensory perception or information processing
84 (Sloman 2007). Pollution can also indirectly affect fish behaviour by eliminating,
85 decreasing or increasing the abundance of behaviour-modifying parasites (Lafferty 1997;
86 Sures 2004; Marcogliese 2005).

87 Parasitism and pollution stress may have combined effects on fish health. Juvenile
88 Chinook salmon (*Oncorhynchus tshawytscha*) experimentally infected with
89 metacercariae of the digenetic trematode *Nanophyetus salmincola* and exposed to PCBs
90 had lower immune function and were more susceptible to infection by the bacterium

91 *Listonella anguillarum* than fish exposed to only individual stressors (Jacobson et al.
92 2003). Yellow perch (*Perca flavescens*) exposed to environmental levels of industrial and
93 agricultural pollution and naturally infected with larvae of the nematode *Raphidascaris*
94 *acus* had higher oxidative stress levels than fish exposed to only one of these stressors
95 (Marcogliese et al. 2005). Spottail shiners (*Notropis hudsonius*) exposed to
96 environmental levels of urban and industrial effluents and naturally infected with the
97 trematode *Plagioporus sinitsini* had more pigmented macrophages in their spleens (a
98 general indicator of stress), and lower condition indices than fish exposed to either
99 stressor alone (Thilakaratne et al. 2007).

100 Behaviour is an important indicator of stress in fish, linking the physiological
101 effects of parasites and pollution with ecological processes (Scott and Sloman 2004;
102 Barber and Rushbrook 2008). Changes in antipredator behaviour are of particular
103 ecological relevance because they have direct consequences for future host fitness.
104 Although parasitism and pollution both have the potential to affect fish behaviour, no
105 studies published to date have considered the combined effects of these two stressors. In
106 this study, we test the combined effects of parasitism and pollution on the antipredator
107 behaviour of johnny darters (*Etheostoma nigrum* Rafinesque, 1820), using fish from
108 contaminated and reference localities in the St. Lawrence River. Johnny darters are small,
109 cryptically coloured benthic fish commonly found in the St. Lawrence River in
110 southwestern Quebec, Canada. They inhabit both relatively pristine and polluted areas of
111 the river, and are host to a diverse community of parasites. The parasite communities of
112 johnny darters from the St. Lawrence River show differences that are correlated with
113 pollution status of sampling localities, as well as the type of pollution (Krause et al.

114 2010). Here we specifically examine whether pollution and parasitism have a combined
115 effect on fish behaviour, and whether changes in fish parasite community assemblages
116 related to pollution have additional effects on fish behaviour. This study uses field-
117 collected specimens to examine the combined effects of natural parasite communities and
118 mixtures of contaminants, both of which are more relevant to understanding natural fish
119 populations than simplified laboratory experiments that focus on single species and
120 chemicals (Jobling 1995; Marcogliese 2005; Bordes and Morand 2009). While the nature
121 of the study location precludes perfect replication of particular pollution mixtures or
122 parasite community assemblages, we expected to see differences between polluted and
123 reference localities, based on other studies using different indicators of pollution and
124 parasite stress in fish collected from the same localities (e.g. Marcogliese et al. 2005;
125 Thilakaratne et al. 2007; Marcogliese et al. 2010).

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127 **Materials and Methods**

128 **Study localities**

129 Fish were collected in June 2008 from five localities in the St. Lawrence River in
130 southwestern Quebec, Canada. These included two reference localities, Îles de la Paix
131 (IPA; 45°20.022' N; 73°51.362' W) and Île Dorval (IDO; 45°26.016' N; 73°44.234' W),
132 and three polluted localities, Beauharnois (BEA; 45°19.051' N; 73°52.020' W), Îlet Vert
133 (IVT; 45°42.230' N; 73°27.143' W) and Île Beaugard (IBE; 45°44.965' N; 73°24.910'
134 W) (Fig. 1). Localities were characterized in previous studies, based on concentrations of
135 metals, polychlorinated biphenyls (PCBs), and other contaminants in the sediments
136 (Loiselle et al. 1997; Marcogliese et al. 2005, 2006; Dautremepuits et al. 2009). These

137 measures are considered an accurate representation of pollution status because sediment
138 contamination is relatively stable over time in this system (Dautremepuits et al. 2009).
139 They are also biologically significant, because johnny darters are benthic organisms that
140 spend their lives in close contact with the sediment and feed on benthic invertebrates
141 (Strange 1991). The reference localities, IPA and IDO, are located upstream of the Island
142 of Montreal in Lake St. Louis. No contaminants surpassing the Canadian Environmental
143 Quality Guidelines Probable Effects Level (PEL) (<http://ceqg-rcqe.ccme.ca/> for aquatic
144 life) were detected at either locality (Marcogliese et al. 2006). One polluted locality,
145 BEA, is also located in Lake St. Louis, at the mouth of the St. Louis River. It is primarily
146 affected by industrial and agricultural activity upstream in the St. Louis River. BEA has
147 high levels of PCBs, organochlorines, and several metals, particularly mercury, which
148 surpass the PEL (Loiselle et al. 1997; Marcogliese et al. 2005; Dautremepuits et al.
149 2009). The other polluted localities, IVT and IBE, are located downstream of Montreal in
150 the plume of the Montreal sewage treatment plant outfall. They both have high levels of
151 organochlorines, PCBs and some metals. PCB levels at IBE and chromium levels at IVT
152 surpass the PEL (Marcogliese et al. 2006; M. Pelletier, personal communication, 2009).

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154 **Study organisms**

155 One hundred and seventy-eight johnny darters were collected using a beach seine
156 (22.6 × 1.15m; 3mm mesh) and transported live to the laboratory. Fish from each locality
157 ($n = 35-36$ per locality) were kept in separate tanks (90cm × 45cm × 35cm). Tanks were
158 lined with aquarium gravel, filled with 60L of dechlorinated tap water, and were
159 continuously aerated. Tanks were covered on three walls with opaque black plastic to

160 prevent fish from seeing those in neighbouring tanks. Fish were kept at 20°C, in a 14:10
161 light:dark regime and were fed with Nutrafin™ fish flakes *ad lib*. The sex ratio of the fish
162 was approximately 1:1, and all fish were presumed to be from the 1+ age class, as
163 demonstrated by the length frequency distributions from each locality (Bagenal and
164 Tesch 1978).

165

166 **Behaviour experiments**

167 Fish were acclimated in the laboratory for at least six days prior to testing (Smith
168 1979). Behaviour experiments for fish from each locality were conducted over two to
169 three consecutive days, within 14 days of collection. Behavioural metrics were chosen
170 based on results from preliminary experiments on johnny darters from two localities in
171 the St. Lawrence River, IPA and IVT, in September 2007. Two experiments were
172 conducted. The first experiment measured capture time of each fish ($n = 178$), defined as
173 the time taken to catch individual fish, and was considered a proxy for susceptibility to
174 predation. Capture time was tested in the same tanks used for acclimation, to minimize
175 unnecessary handling of the fish. The experiment consisted of catching fish one by one
176 from the large tank using a hand-held dip net (43cm long, 13cm × 16cm opening). During
177 the experiment, the dip net was placed in the middle of the water column in the centre of
178 the tank and shaken vigorously to alert the fish to the net “predator.” The net was then
179 moved in a regular manner counterclockwise along the walls of the tank, at an
180 approximately constant speed of 20cm/s around the tank until a fish was caught. This
181 method of capturing the fish, including the capture speed of 20cm/s, was optimized
182 during a pilot study. This procedure was repeated until all fish were caught. The order in

183 which fish were removed from each tank was recorded as “capture order” and examined
184 as an additional behaviour measure.

185 After fish were caught in the capture time experiment, they were transferred into
186 test tanks for the second behaviour experiment, a measurement of flight initiation
187 distance. Fish were paired in narrow test tanks (90cm × 30cm × 35cm, 50L), which were
188 covered on three walls with black, opaque plastic to hide the experimenter from view of
189 the fish. They were left to acclimate in the tanks for two hours before beginning the trial,
190 during which time two carbon water filters were run in the test tanks to remove any
191 chemical cues left by fish previously tested in the tanks. The water filters were turned off
192 during the trial. Flight initiation distance was measured by moving a model of a predatory
193 fish towards the two fish at an approximate speed of 10 cm/s, starting from the end of the
194 tank farthest from the fish. The speed of approach was identified during preliminary trials
195 as the optimal speed for the experiment. The predator model used was a semi-realistic
196 plastic model of a fish, approximately five times larger than the johnny darters. Flight
197 initiation distance was measured for the “focal” fish ($n = 89$), the fish closest to the
198 approaching predator; the second “dither” fish was placed in the tank to reduce the stress
199 level of the focal fish (Brown et al. 2006). The experiment was filmed and flight
200 initiation distance, defined as the distance from the predator model at which the fish
201 initiated movement, was measured from the video recording.

202 Following the behaviour experiments, fish were killed with an overdose of clove
203 oil solution (50 mg/L) and frozen for later necropsy. All animal collection and
204 experimental procedures were in accordance with guidelines of the Canadian Council on
205 Animal Care in effect at the time of the study.

206 **Examination for parasites**

207 Frozen mass (mg) and standard length (mm) were measured for each fish and
208 followed by a complete necropsy. Parasites from tissues and organs, including fins, skin,
209 gills, eyes, brain, body cavity, gastrointestinal tract, liver, heart, spleen, gonads and
210 muscle were collected following standard parasite examination protocols (Marcogliese
211 2002). During the necropsy, all parasites were enumerated and identified to genus, with
212 the exception of acanthocephalans, non-gyrodactylid monogeneans, and a few rare
213 trematodes, which could only be identified to higher taxonomic levels. Representative
214 samples of parasites recovered from each locality were preserved in 70% ethanol for later
215 identification. Trematodes, cestodes, acanthocephalans and some monogeneans were
216 stained with acetocarmine, cleared with xylene and mounted in Permount or Canada
217 balsam. Other monogeneans were mounted unstained in Hoyer's medium. The remaining
218 monogeneans and all nematodes and copepods were cleared in glycerine alcohol and
219 examined in temporary mounts. Identifications were made using keys in Beverly-Burton
220 (1984), Kabata (1988), Caira (1989), Moravec (1994), Gibson (1996), Scholz (1997) and
221 Hoffman (1999).

222

223 **Statistical analysis**

224 Mean total parasite number, infracommunity species richness, standard length,
225 capture time and flight initiation distance of fish were tested among localities and
226 between polluted and reference localities. Comparisons among localities and between
227 treatments were made using ranked data by one-way ANOVAs (Scheirer and Hare 1976)
228 followed by Tukey-Kramer HSD tests, except for standard length, which was tested using

229 untransformed data. Because capture order of individual fish was dependent on capture
230 order of other fish from within the same tank (i.e. locality), it could not be compared
231 among localities or between fish pooled by pollution status. Separate regression
232 comparisons for each locality were made between capture order of individual fish and
233 their total parasite number and infracommunity species richness (15 comparisons total).
234 All univariate tests were conducted using JMP® 7.0.1 (© 2007 SAS Institute Inc.).

235 Multivariate analyses were conducted using the PERMANOVA+ add-on for
236 PRIMER (© 2006 Plymouth Routines In Multivariate Ecological Research, Plymouth,
237 UK). A stepwise regression of capture time with abundances of all parasite species was
238 performed with a distance-based linear model (DISTLM). This test allows for a stepwise
239 test of continuous variables that are not normally distributed. Species that significantly
240 correlated with capture time were included as covariates in a permutational multivariate
241 ANOVA (PERMANOVA) of capture time. PERMANOVA is a nonparametric test
242 analogous to a multivariate ANOVA. It gives the test statistic Pseudo- F , which is
243 analogous to the F statistic in measuring the among-group to within-group variation. The
244 initial model also included mean total parasite number, mean infracommunity species
245 richness, mean standard length, locality, and interactions between variables. The final
246 model included only terms that significantly explained capture time.

247 Mean abundances of parasite species included in the PERMANOVA were
248 individually compared among all localities using ANOVAs on ranked data, followed by a
249 Tukey HSD tests. Tests between polluted and reference localities were performed with
250 nonparametric Wilcoxon tests.

251

252 **Terminology**

253 Parasite terminology adheres to definitions of Bush et al. (1997). Prevalence is the
254 percentage of hosts infected with a given parasite species in a sample. Abundance is the
255 number of parasites of a given species infecting a given host, whether the host is infected
256 or not. Mean abundance is the number of parasites of a given species averaged over the
257 whole host sample, and includes hosts with and without infections. Intensity is the
258 number of parasites of a given species infecting a host, and mean intensity is the number
259 of parasites of that species averaged across infected hosts in a sample. An
260 infracommunity refers to all the individuals of all the parasite species within an
261 individual host. Locality refers to the geographic area from which the host was collected,
262 and site refers to the specific host tissue or organ from which the parasite was collected.

263

264 **Results**

265 Twenty-four species of parasites were identified in the 178 darters examined. The
266 prevalence and mean intensity of each parasite species at each locality are presented in
267 Krause et al. (2010). Mean total parasite number was highest at BEA and IDO, two
268 upstream localities, and lowest at IVT and IBE, both downstream polluted localities
269 ($F_{4,173} = 31.73, p < 0.0001$; Table 1). Mean infracommunity species richness was greatest
270 at BEA, second highest at IPA and IDO, and lowest at the downstream polluted localities,
271 IVT and IBE ($F_{4,173} = 38.48, p < 0.0001$). Standard length was significantly larger for fish
272 from BEA than those from IDO ($F_{4,173} = 3.62, p = 0.007$), but did not differ among fish
273 from other localities.

274 Capture time differed significantly among localities ($F_{4,173} = 6.20, p = 0.0001$),
275 with the longest capture time at IDO and the shortest capture time at IVT (Fig. 2).

276 Capture time did not differ significantly between fish from polluted and reference
277 localities ($F_{1,176} = 1.57, p = 0.12$). Flight initiation distance did not differ significantly
278 between localities ($F_{4,173} = 0.33, p = 0.85$) or between polluted and reference localities
279 ($F_{1,176} = 0.65, p = 0.42$). There was no correlation between capture order and standard
280 length, total parasite number or parasite species richness (all p values ≥ 0.07).

281 Mean total parasite number and mean infracommunity species richness was
282 weakly, but significantly correlated with capture time (total parasite number: $R^2 = 0.03, n$
283 $= 178, p = 0.03$; species richness: $R^2 = 0.03, n = 178, p = 0.03$). The only parasite species
284 that was related to capture time was *Ornithodiplostomum* sp. A nonparametric DISTLM
285 analysis of pooled data showed that the relationship between *Ornithodiplostomum* sp.
286 abundance and capture time was significantly positive ($R^2 = 0.15, n = 178, p = 0.0001$;
287 Fig. 3), suggesting that fish with higher intensity infections might be less susceptible to
288 capture than fish with low or no infection. *Ornithodiplostomum* sp. mean abundance was
289 highest at IDO, followed by IPA, and was lowest at BEA, IVT and IBE ($F_{4,173} = 46.6, p$
290 < 0.0001 ; Fig. 4). It was significantly higher at reference than polluted localities ($F_{1,176} =$
291 $119.40, p < 0.0001$). Capture time was best explained by a PERMANOVA model
292 including *Ornithodiplostomum* sp. abundance (Pseudo- $F = 18.82, p = 0.002, df = 1$) and
293 locality (Pseudo- $F = 2.45, p = 0.039, df = 4$).

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297 **Discussion**

298 Johnny darters with high intensities of the brain-encysting parasite,
299 *Ornithodiplostomum* sp., had longer capture times than fish with low or no infections,
300 perhaps reflecting an increase in activity of infected fish. Johnny darters normally exhibit
301 a cessation of movement when they detect a predator (Smith 1979). Stressors that induce
302 hyperactivity may disrupt adaptive anti-predator behaviour in this species. In this study,
303 darters exhibiting typical antipredator behaviour appeared to be more susceptible to
304 capture, while fish behaving abnormally, by moving quickly and erratically, and
305 swimming to the surface, were more difficult to catch. In natural systems, however,
306 predators such as mergansers and other piscivorous diving birds, the definitive hosts of
307 *Ornithodiplostomum* spp., typically depend on visual cues such as movement to capture
308 their prey, and an increase in activity may make cryptic fish such as johnny darters more
309 susceptible to predation (Ydenberg and Dill 1986). These results should be interpreted
310 with caution, because our sampling design does not allow us to consider parasitism and
311 pollution separately; however, deviations from typical, cryptic anti-predator behaviour of
312 johnny darters caused by high intensities of *Ornithodiplostomum* sp. may reflect an
313 adaptation of the parasite to increase its transmission success. Alternatively, the increased
314 activity observed could simply be a pathogenic by-product of infection (Poulin 1995).

315 Neither parasitism nor pollution could statistically explain observed differences in
316 either of the other two behavioural measures, capture order or flight initiation distance.
317 Capture time has not been used in previous studies; however it was measured because it
318 showed a significant correlation with parasitism in a pilot study. Flight initiation distance
319 is a measure commonly used to assess fish reactions to predation risk (Ydenberg and Dill

320 1986). The lack of response in this study suggest that it may be an inappropriate measure
321 of anti-predator behaviour in a species such as the johnny darter that typically exhibits a
322 cessation of movement in response to perceived predators.

323 Studies of fathead minnows (*Pimephales promelas*) with infections of
324 *Ornithodiplostomum ptychocheilus* suggest that behavioural changes may be caused by
325 adaptive manipulation by the parasite or pathology of parasite development in the host.
326 Fathead minnows with mature infections of *O. ptychocheilus* exhibited less compact
327 shoaling behaviour and swam higher in the water column, which may make them more
328 susceptible to predation (Radabaugh 1980). Alternatively, minnows with new infections
329 of *O. ptychocheilus* showed reduced standard optomotor response (OMR), likely due to
330 damage caused at the site of infection, the optic tectum (Shirakashi and Goater 2001,
331 2002). The greatest decrease in OMR occurred during parasite development and subsided
332 after they reached infectivity, reflecting damage to the optic lobes during parasite growth
333 (Shirakashi and Goater 2005). Behavioural changes induced before a parasite becomes
334 infective are considered pathological, while those that ensue following development to
335 the infective forms may be evidence of adaptation (Poulin 1995). The present study does
336 not explore the specific physiological mechanisms of the observed behavioural change,
337 nor does it measure actual predation rates of infected and non-infected fish. However,
338 evidence from other *Ornithodiplostomum*-fish systems, as seen above, suggests that both
339 scenarios are possible. In our study, parasites were encysted and presumably infective,
340 lending support to the idea that the behavioural changes may be adaptive. Further
341 experiments to test the fitness consequences for both the parasite and host are necessary

342 to determine whether the behaviour change seen here is an adaptive modification by the
343 parasite or merely a pathological side effect (Poulin 1995).

344 Locality was also significantly correlated with differences in fish behaviour. This
345 may reflect a tank effect in the experimental design, because fish from each locality were
346 kept and tested in a single tank. However, it may also be due to a parasite effect that was
347 not statistically detectable. Mean capture time of fish from different localities showed
348 patterns similar to patterns of parasite community parameters: fish from BEA and IDO
349 had higher capture times than fish from IVT, and also higher mean species richness and
350 mean total parasite number. Only *Ornithodiplostomum* sp., the parasite in the highest
351 abundance, was significant in the model of capture time, however failure to detect effects
352 of other species may be due to low infection intensities and species richness. However,
353 the fact that species richness was weakly correlated with capture time lends some support
354 to the idea that parasite diversity may have impacts on individual hosts (Bordes and
355 Morrand 2009). There was no interaction between locality and mean abundance of
356 *Ornithodiplostomum* sp., suggesting that the effects of the parasite on behaviour were
357 independent of pollution exposure.

358 A direct, general effect of pollution on fish behaviour was not detected, nor could
359 we detect an interactive effect of pollution and parasitism. However, pollution appears to
360 have a negative effect on *Ornithodiplostomum* sp. infections in johnny darters in this
361 system, through reducing the abundance of this parasite (Krause et al. 2010). Free-living
362 cercariae of digenetic trematodes are sensitive to a variety of types of pollution, including
363 metals, acidification, chemical fertilizers and pesticides, which can reduce their survival,
364 longevity, encystment and infectivity (Morley et al. 2003; Pietrock and Marcogliese

365 2003). Cercariae of *O. ptychocheilus* exposed to cadmium showed decreased infectivity
366 to fish (Pietroock and Goater 2005). Therefore, metal pollution may indirectly affect
367 johnny darter behaviour at contaminated localities, through the reduction of survival
368 and/or infectivity of cercariae of *Ornithodiplostomum* sp.

369 Previous studies of effects of pollution and parasite stress on fish behaviour have
370 focused primarily on single pollutants and single parasite species, and have not tested
371 both stressors together. This study considers these stressors in combination, and tests
372 naturally-infected fish obtained directly from polluted localities. This approach can limit
373 the interpretive power of the study because it does not allow hypotheses regarding effects
374 of specific pollutants to be tested. However this observational approach is nonetheless
375 valuable because it can provide important information about the effects of actual
376 conditions and mixtures. In nature, pollution stress is often due to combinations of many
377 chemicals (Jobling 1995; Lafferty 1997; Marcogliese 2005) and fish are commonly
378 infected with communities of parasites (Barber et al. 2000; Barber and Rushbrook 2008;
379 Bordes and Morand 2009), conditions that are difficult to replicate in laboratory
380 experiments.

381

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394

395 **References**

- 396 Atchison, G.J., Henry, M.G., and Sandheinrich, M.B. 1987. Effects of metals on fish
397 behavior: a review. *Environ. Biol. Fishes.*, **18**(1): 11-26.
- 398 Bagenal, T.B., and Tesch, F.W. 1978. Age and growth. *In* Methods for assessment of fish
399 production in fresh waters. *Edited by* T. Bagenal. Blackwell Scientific
400 Publications, Oxford, NY. pp. 101-136.
- 401 Barber, I., and Rushbrook, B.J. 2008. Parasites and fish behaviour. *In* Fish Behaviour.
402 *Edited by* C. Magnhagen, V.A. Braithwaite, E. Forsgren, and B.G. Kapoor. Science
403 Publishers, Enfield, NH. pp. 525-561.
- 404 Barber, I., Hoare, D., and Krause, J. 2000. Effects of parasites on fish behaviour: a review
405 and evolutionary perspective. *Rev. Fish Biol. Fish.* **10**(2): 131-165.
406 doi:10.1023/A:1016658224470.
- 407 Beverley-Burton, M. 1984. Monogenea and Turbellaria. *In* Guide to the Parasites of
408 Fishes of Canada. Part I. *Edited by* L. Margolis and Z. Kabata. Can. Spec. Publ. Fish.
409 Aquat. Sci. No. 74. pp. 1-209.

410 Bordes, F., and Morand, S. 2009. Parasite diversity: an overlooked metric of parasite
411 pressures? *Oikos*, **118**(6): 801-806. doi:10.1111/j.1600-0706.2008.17169.x.

412 Brown, G.E., Bongiorno, T., DiCapua, D.M., Ivan, L.I., and Roh, E. 2006. Effects of
413 group size on the threat-sensitive response to varying concentrations of chemical
414 alarm cues by juvenile convict cichlids. *Can. J. Zool.* **84**(1): 1-8. doi:10.1139/Z05-
415 166.

416 Bush, A.O., Lafferty, K.D., Lotz, J.M., and Shostak, A.W. 1997. Parasitology meets
417 ecology on its own terms: Margolis et al. revisited. *J. Parasitol.* **83**(4): 575-583.

418 Cairn, J.N. 1989. A revision of the North American papillose Allocreadiidae (Digenea)
419 with independent cladistic analyses of larval and adult forms. *Bull. Univ. Nebr. State*
420 *Mus.* **11**: 58.

421 Clotfelter, E.D., Bell, A.M., and Levering, K.R. 2004. The role of animal behaviour in
422 the study of endocrine-disrupting chemicals. *Anim. Behav.* **68**: 665-676.
423 doi:10.1016/j.anbehav.2004.05.004.

424 Dautremepuits, C., Marcogliese, D.J., Gendron, A.D., and Fournier, M. 2009. Gill and
425 head kidney antioxidant processes and innate immune system responses of yellow
426 perch (*Perca flavescens*) exposed to different contaminants in the St. Lawrence
427 River, Canada. *Sci. Total Environ.* **407**(3): 1055-1064.
428 doi:10.1016/j.scitotenv.2008.10.004.

429 Gibson, D.I. 1996. Trematoda. *In* Guide to the Parasites of Fishes of Canada. Part IV.
430 *Edited by* L. Margolis and Z. Kabata. *Can. Spec. Publ. Fish. Aquat. Sci.* No. 124. pp.
431 1-373.

432 Hoffman, G.L. 1999. Parasites of North American freshwater fishes. 2nd ed. Comstock
433 Publishing Associates, Ithaca, N.Y.

434 Jacobson, K.C., Arkoosh, M.R., Kagley, A.N., Clemons, E.R., Collier, T.K., and Casillas,
435 E. 2003. Cumulative effects of natural and anthropogenic stress on immune function
436 and disease resistance in juvenile Chinook salmon. J. Aquat. Anim. Health, **15**(1): 1-
437 12. doi:10.1577/1548-8667(2003)015<0001:CEONAA>2.0.CO;2.

438 Jobling, M. 1995. Environmental biology of fishes. Chapman and Hall, London, UK.

439 Kabata, Z. 1988. Crustacea. *In* Guide to the Parasites of Fishes of Canada. Part II -
440 Copepoda and branchiura. *Edited by* L. Margolis and Z. Kabata. Can. Spec. Publ.
441 Fish. Aquat. Sci. No. 101. pp 3-127.

442 Krause, R.J., McLaughlin, J.D., and Marcogliese, D.J. 2010. Parasite fauna of
443 *Etheostoma nigrum* (Percidae: Etheostomatinae) in localities of varying pollution
444 stress in the St. Lawrence River, Quebec, Canada. Parasitol. Res. **107**(2): 285-294.
445 doi: 10.1007/s00436-010-1862-6.

446 Lafferty, K.D. 1997. Environmental parasitology: what can parasites tell us about human
447 impacts on the environment? Parasitol. Today, **13**(7): 251-255. doi:10.1016/S0169-
448 4758(97)01072-7.

449 Loiselle, C., Fortin, G.R., Lorraine, S., and Pelletier, M. 1997. Dynamics and
450 contamination of the St. Lawrence River sediment. Environment Canada, Montréal,
451 Que.

452 Marcogliese, D.J. 2002. Protocols for measuring biodiversity: parasites of fishes in fresh
453 water. Ecological Monitoring and Assessment Network, Environment Canada.
454 <http://www.eman-rese.ca/eman/ecotools/protocols/freshwater/parasites>.

455 Marcogliese, D.J. 2005. Parasites of the superorganism: are they indicators of ecosystem
456 health? *Int. J. Parasitol.* **35**(7): 705-716. doi:10.1016/j.ijpara.2005.01.015.

457 Marcogliese, D.J., Brambilla, L.G., Gagne, F., and Gendron, A.D. 2005. Joint effects of
458 parasitism and pollution on oxidative stress biomarkers in yellow perch *Perca*
459 *flavescens*. *Dis. Aquat. Org.* **63**(1): 77-84. doi:10.3354/dao063077

460 Marcogliese, D.J., Dautremepuits, C., Gendron, A.D., and Fournier, M. 2010.
461 Interactions between parasites and pollutants in yellow perch (*Perca flavescens*) in
462 the St. Lawrence River, Canada: implications for resistance and tolerance to
463 parasites. *Can. J. Zool.* **88**(3): 247-258. doi:10.1139/Z09-140.

464 Marcogliese, D.J., Gendron, A.D., Plante, C., Fournier, M., and Cyr, D. 2006. Parasites
465 of spottail shiners (*Notropis hudsonius*) in the St. Lawrence River: effects of
466 municipal effluents and habitat. *Can. J. Zool.* **84**(10): 1461-1481. doi:10.1139/Z06-
467 088.

468 Moore, J. 2002. Parasites and the behavior of animals. Oxford University Press, Oxford,
469 NY.

470 Moravec, F. 1994. Parasitic nematodes of freshwater fishes of Europe. Kluwer Academic
471 Publishers, Dordrecht, Germany.

472 Morley, N.J., Irwin, S.W.B., and Lewis, J.W. 2003. Pollution toxicity to the transmission
473 of larval digeneans through their molluscan hosts. *Parasitology*, **126**: S5-S26.
474 doi:10.1017/S0031182003003755.

475 Pietrock, M., and Goater, C.P. 2005. Infectivity of *Ornithodiplostomum ptychocheilus*
476 and *Posthodiplostomum minimum* (Trematoda : Diplostomidae) cercariae following
477 exposure to cadmium. *J. Parasitol.* **91**(4): 854-856. doi: 10.1645/GE-473R.1.

478 Pietrock, M., and Marcogliese, D.J. 2003. Free-living endohelminth stages: at the mercy
479 of environmental conditions. *Trends Parasitol.* **19**(7): 293-299. doi:10.1016/S1471-
480 4922(03)00117-X.

481 Poulin, R. 1995. "Adaptive" changes in the behaviour of parasitized animals: a critical
482 review. *Int. J. Parasitol.* **25**(12): 1371-1383. doi:10.1016/0020-7519(95)00100-X.

483 Radabaugh, D.C. 1980. Changes in minnow, *Pimephales promelas* Rafinesque, schooling
484 behavior associated with infections of brain encysted larvae of the fluke,
485 *Ornithodiplostomum pychocheilus*. *J. Fish Biol.* **16**(6): 621-628. doi:10.1111/j.1095-
486 8649.1980.tb03741.x.

487 Scheirer, C.J., Ray, W.S., and Hare, N. 1976. The analysis of ranked data derived from
488 completely randomized factorial designs. *Biometrics*, **32**(2): 429-434.

489 Scholz, T. 1997. A revision of the species of *Bothriocephalus* Rudolphi, 1808 (Cestoda:
490 Pseudophyllidea) parasitic in American freshwater fishes. *Syst. Parasitol.* **36**(2): 85-
491 107. doi:10.1023/A:1005744010567.

492 Scott, G.R., and Sloman, K.A. 2004. The effects of environmental pollutants on complex
493 fish behaviour: integrating behavioural and physiological indicators of toxicity.
494 *Aquat. Toxicol.* **68**(4): 369-392. doi:10.1016/j.aquatox.2004.03.016.

495 Shirakashi, S., and Goater, C.P. 2001. Brain-encysting parasites affect visually-mediated
496 behaviours of fathead minnows. *Ecoscience*. **8**(3): 289-293.

497 Shirakashi, S., and Goater, C.P. 2002. Intensity-dependent alteration of minnow
498 (*Pimephales promelas*) behavior by a brain-encysting trematode. *J. Parasitol.* **88**(6):
499 1071-1074. doi: 10.1645/0022-3395(2002)088[1071:IDAOMP]2.0.CO;2.

500 Shirakashi, S., and Goater, C.P. 2005. Chronology of parasite-induced alteration of fish
501 behaviour: effects of parasite maturation and host experience. *Parasitology*, **130**(2):
502 177-183. doi:10.1017/S0031182004006432

503 Sloman, K.A. 2007. Effects of trace metals on salmonid fish: the role of social
504 hierarchies. *Appl. Anim. Behav. Sci.* **104**(3-4): 326-345.
505 doi:10.1016/j.applanim.2006.09.003.

506 Smith, R.J.F. 1979. Alarm reaction of Iowa darter, *Etheostoma exile*, and johnny darters,
507 *Etheostoma nigrum*, (Percidae, Pisces) to chemicals from injured conspecifics. *Can.*
508 *J. Zool.* **57**(6): 1278-1282.

509 Strange R. M. 1991. Diet selectivity in the johnny darter, *Etheostoma nigrum*, in Stinking
510 Fork, Indiana. *J. Freshw. Ecol.* **6**: 377-381.

511 Sures, B. 2004. Environmental parasitology: relevancy of parasites in monitoring
512 environmental pollution. *Trends Parasitol.* **20**(4): 170-177.
513 doi:10.1016/j.pt.2004.01.014.

514 Thilakaratne, I.D.S.I.P., McLaughlin, J.D., and Marcogliese, D.J. 2007. Effects of
515 pollution and parasites on biomarkers of fish health in spottail shiners, *Notropis*
516 *hudsonius* (Clinton). *J. Fish Biol.* **71**(2): 519-538. doi:10.1111/j.1095-
517 8649.2007.01511.x.

518 Ydenberg, R.C., and Dill, L.M. 1986. The economics of fleeing from predators. *Adv.*
519 *Stud. Behav.* **16**: 229-249.

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523 **Table 1.** Mean total parasites, parasite infracommunity species richness and total length
 524 \pm SD of johnny darters from five localities in June 2008 in the St. Lawrence River in
 525 Quebec, Canada: Beauharnois (BEA), an upstream polluted locality, Île Beauregard
 526 (IBE) and Îlet Vert (IVT), downstream polluted localities, and Île Dorval (IDO) and Îles
 527 de la Paix (IPA).

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Locality	Mean total parasites \pm SD	Mean parasite infracommunity species richness \pmSD	Mean total length (mm) \pm SD
BEA	45.6 \pm 30.5	7.7 \pm 1.9	51.0 \pm 6.9
IVT	14.17 \pm 11.7	3.3 \pm 1.3	47.9 \pm 4.3
IBE	13.5 \pm 10.8	3.9 \pm 1.5	50.0 \pm 4.4
IPA	32.0 \pm 28.5	5.6 \pm 2.0	46.9 \pm 5.3
IDO	54.6 \pm 41.4	5.2 \pm 1.5	48.1 \pm 5.1

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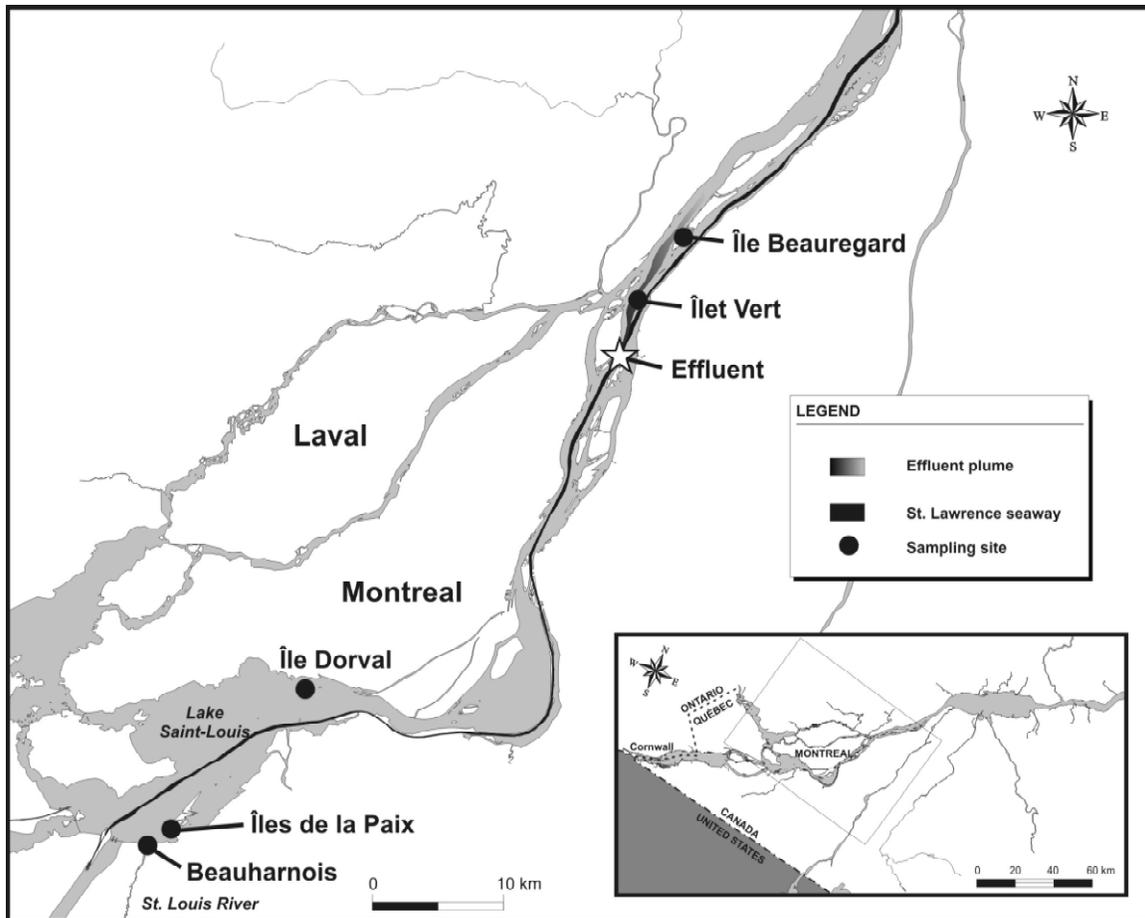
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540 **Fig. 1.** Map of the St. Lawrence River in southwestern Quebec, Canada, showing the five
541 localities sampled in June 2008: one upstream polluted locality, Beauharnois (BEA); two
542 downstream polluted localities, Îlet Vert (IVT) and Île Beaugard (IBE); and two
543 reference localities, Îles de la Paix (IPA) and Île Dorval (IDO).

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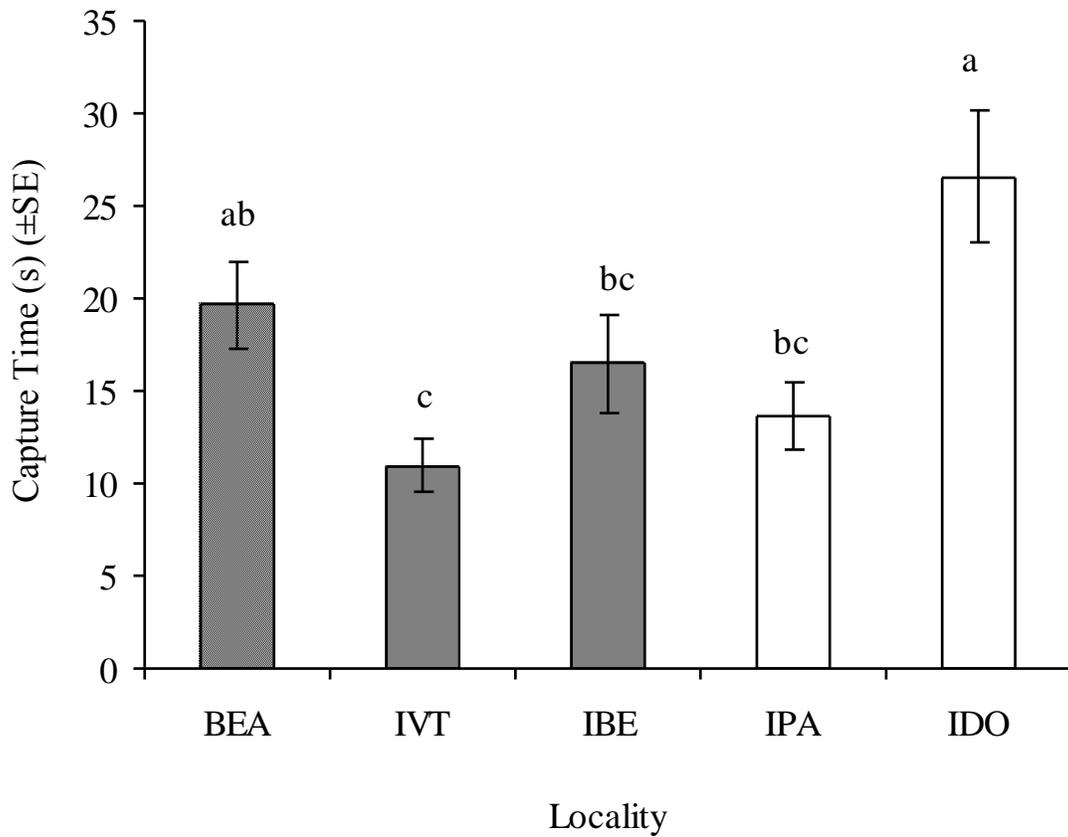
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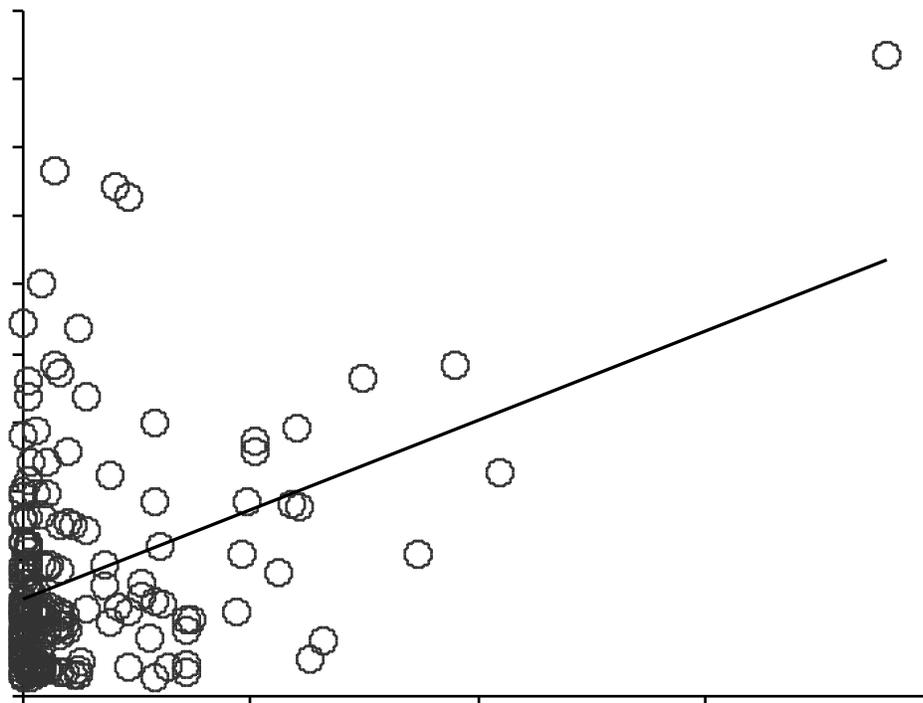
551 **Fig. 2.** Mean capture time (s) \pm standard error of johnny darters from five localities in
552 June 2008 in the St. Lawrence River in southwestern Quebec, Canada: one upstream
553 polluted locality (light grey), Beauharnois (BEA); two downstream polluted localities
554 (dark grey), Île Beauregard (IBE), Îlet Vert (IVT); and two reference localities (white),
555 Île Dorval (IDO) and Îles de la Paix (IPA). Different letters indicate significant
556 differences between localities.
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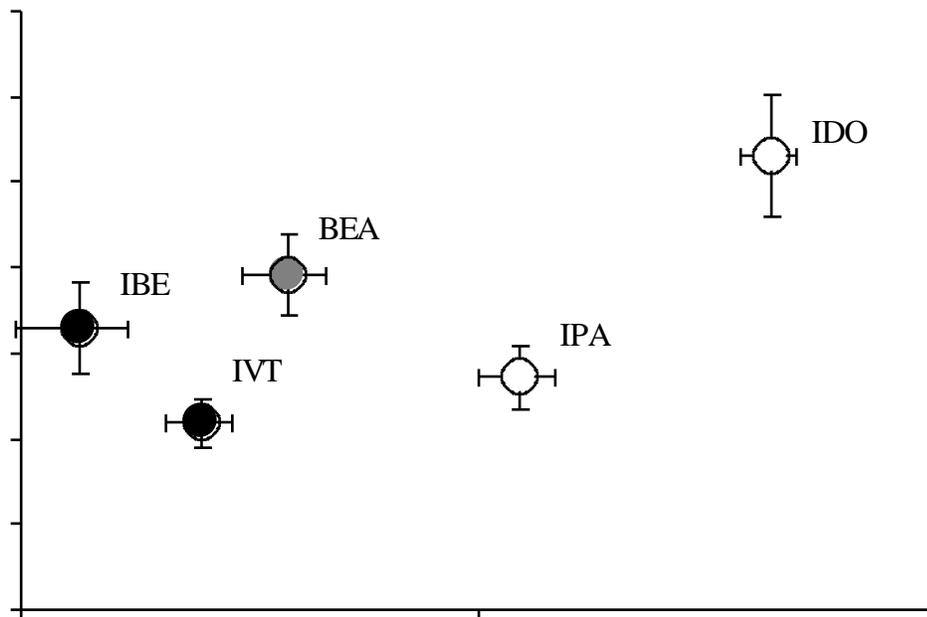
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560 **Fig. 3.** Scatter plot of capture time (s) versus *Ornithodiplostomum* sp. abundance for
561 johnny darters from five localities in June 2008 in the St. Lawrence River, Quebec,
562 Canada.
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570 **Fig. 4.** Mean capture time (s) and mean *Ornithodiplostomum* sp. abundance in johnny
571 darters from five localities in June 2008 in the St. Lawrence River in Quebec, Canada:
572 one upstream polluted locality (grey circle), Beauharnois (BEA); two downstream
573 polluted localities (black circles), Île Beauregard (IBE) and Îlet Vert (IVT); and two
574 reference localities (white circles), Île Dorval (IDO) and Îles de la Paix (IPA). Error bars
575 represent standard errors.
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