

PROPERTIES OF MIXTURES OF FATTY ACIDS  
PROPOSED AS LATENT HEAT THERMAL STORAGE  
FOR SPACE HEATING

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A Thesis  
in  
The Centre  
for  
Building Studies  
Faculty  
of  
Engineering

Presented in Partial Fulfillment of the Requirements  
for the degree of Master of Engineering (Building) at  
Concordia University  
Montréal, Québec, Canada

September 1983

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ABSTRACT

An analysis of thermal properties of fatty acids and their binary mixtures has shown that they are attractive candidates for latent heat thermal storage in space heating applications. In this study, the method of differential scanning calorimetry was used to determine the melting point, freezing point, heat of melting and heat of crystallization of the fatty acids and their binary mixtures. These properties are of prime importance in the design of a latent heat thermal storage system.

The melting range of the fatty acids (capric, lauric, palmitic and stearic) was observed to be from approximately 30°C to 65°C. The heat of transition of these acids (melting and crystallization) was observed to have a range from approximately 153J/g to 183J/g.

The eutectic points were determined for the binary mixtures of the fatty acids. The melting points of the eutectics for the binary systems of capric-lauric, lauric-palmitic, lauric-stearic and palmitic-stearic acids were observed to be 18.2°C, 32.7°C, 34.0°C and 51.1°C respectively. The corresponding heats of melting were found to be 120J/g, 147J/g, 152J/g and 159J/g respectively.

The fatty acids and their eutectic mixtures were examined with an X-ray diffractometer and an infrared spectrophotometer to ascertain the polymorphic forms, crystal structures and material purities. The experimental results indicate that the fatty acids are present in the C-polymorphic form.

ACKNOWLEDGEMENTS

I wish to express my gratitude to Professor D. Feldman, who has guided me in my work and has rendered valuable assistance. I am also grateful to Professor M. Shapiro, who has offered thoughtful criticism and provided many worthwhile suggestions. Special thanks are due to Mrs. Dorina Banu, who has aided me on numerous occasions during the experiments.

I also wish to express my thanks to Dr. C. Sandofi of Université de Montréal who has helped me with the interpretation of the infrared spectra and Dr. Bird of Concordia University who has allowed me to use the X-ray diffractometer at Concordia University and later helped me with the interpretation of the X-ray spectra.

To my son

Oren James Fuks

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## I, INTRODUCTION

Man's requirement for energy is increasing at a rapid rate and, in view of the diminishing supply of the non-renewable sources such as fossil fuels, alternative supplies will need to be developed to meet the demand.

Low temperature heat, particularly in the form of solar energy, is abundantly available and can be utilized in various domestic and industrial applications (i.e., waste heat recovery in plants, space heating, greenhouse heating, domestic hot water generation, solar cooling, etc...). Widespread use of solar energy will depend on many factors. The development of a cost-effective compact thermal energy storage system is one of the most important of these factors.

The phase-change heat storage technique is based on the harnessing of the latent heat energy when a substance undergoes melting and freezing. There are various organic and inorganic materials which can be used for low temperature heat storage. Normally, these materials are classified as paraffins, fatty acids, inorganic salt hydrates and eutectic compounds.

This thesis is confined to the discussion of fatty acids (capric, lauric, palmitic and stearic) and their binary mixtures. These acids have melting points from approximately 30°C to 70°C and, therefore, possess a suitable melting range for solar heat storage, particularly in the space heating application. At the present time, there are a few suitable materials which are available on the market

that can effectively store solar energy for space heating. Several inorganic salt hydrates have been employed but these materials have been shown to be generally unsuitable due to the supercooling effect and the segregation of insoluble compounds.

The supercooling phenomenon is undesirable because it gives rise to instability in crystallization. The inorganic salt hydrates possess poor nucleating properties which result in supercooling of the liquid. The supercooling effect can be reduced by several methods which include (1) the addition of a nucleating agent with a similar crystal structure to that of the mother substance, (2) mechanical agitation and (3) by using rough heat exchanging surfaces to promote heterogeneous nucleation. All of these approaches have been used with a limited success. The other limiting factor of the inorganic salt hydrate is the decomposition of the original salt hydrate. The decomposition process is irreversible. The salt hydrate melts incongruently, producing a saturated aqueous phase and a solid phase containing a lower hydrate of the same salt. Due to the specific gravity difference, the solid settles out at the bottom of the container and does not redissolve by simply increasing the temperature.

Fatty acids meet the thermodynamic and kinetic criteria for low temperature latent heat storage. They have melting points in a low temperature range, possess a high latent heat of fusion per unit mass, have a high density, high specific heat for absorbing sensible heat, high thermal conductivity for maintaining low temperature gradients

during charging and discharging the heat storage material (fatty acid), exhibit small volume changes during the phase transition and have little or no supercooling during freezing. The saturated fatty acids appear to have a good chemical stability and do not decompose after hundreds of phase changes.

The objective of this study is to determine thermal properties of a selected group of fatty acids and their binary mixtures and to demonstrate the potential of using these materials for latent heat thermal storage. The thermal properties which are investigated here include (1) melting point, (2) freezing point, and (3) heats of melting and crystallization. In addition, the temperatures for maximum endothermic heat flow (observed during heating) and maximum exothermic heat flow (observed during cooling) are measured. A differential scanning calorimeter (DSC) was used to generate the thermal data.

The thermal data was, subsequently, used to construct phase diagrams of the binary systems of fatty acids to evaluate the freezing and melting characteristics of the substances. The phase behaviour was found to be characterized by the presence of one eutectic and a region of compound formation. The observations are in general agreement with the previously published data. The published data on melting and freezing points of pure fatty acids and their mixtures were to a large extent developed by the classical capillary tube method. The literature data on the heat of fusion of pure substances were measured by cryoscopic and calorimetric methods.

Little work was done previously in examining the properties of eutectic compounds of fatty acids and there is no experimental data available on the heats of melting and crystallization of these compounds. The heats of melting and crystallization were determined with relative ease and reliability by the differential scanning calorimetry technique. This is in contrast to the classical methods which are laborious and time-consuming. The DSC provides a thermogram on a very small quantity of sample (approximately 1-10 mg.), which can be used to evaluate the melting point, freezing point, degree of supercooling, heats of melting and crystallization and specific heat.

The crystal structure of the pure fatty acids and their eutectic compounds was examined by means of X-ray diffraction and infrared spectroscopy. The experimental data shows that the pure fatty acids and their mixtures are present in the stable C-polymorphic form. This is in agreement with results determined by previous workers.

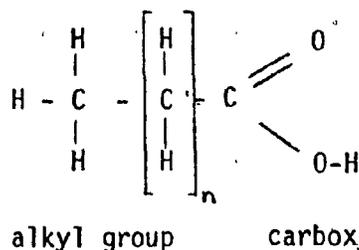
The text of the thesis has been organized to provide a continuity of ideas in a logical sequence. The subject matter is introduced by giving an overview of the general physical and chemical characteristics of fatty acids with a brief mention of the cost and availability. This is followed by discussion of the DSC technique, apparatus, calibration and precision, and definition of technical terms used in the text. The DSC thermograms are then used in constructing graphs and in tabulating data to facilitate the discussion of thermal properties of the fatty acids and their mixtures. A dis-

cussion is later presented on the phase behaviour of a two-component system of fatty acids. The crystal structure of the fatty acids and their eutectic compounds is examined with the aid of infrared and X-ray diffractometric results. The thermal and structural findings are then used in a general discussion of the theory of melting and crystallization of fatty acids.

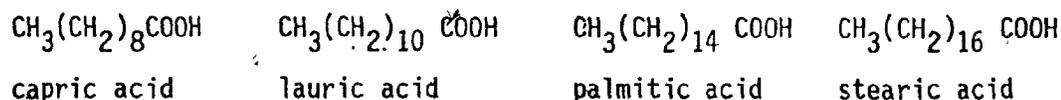
The thesis ends with concluding remarks and recommendations for further work. The original thermograms, X-ray and infrared spectra have been included in the appendix.

## II. CLASSIFICATION OF FATTY ACIDS

The term "fatty acid" is generally applied to a family of organic compounds containing the carboxyl group. The carboxyl group can be attached to an alkyl group as illustrated below:



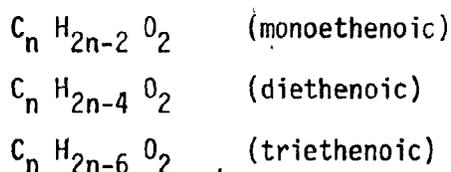
The fatty acid having the above structure is known as a saturated, monobasic, aliphatic acid. The common names and formulas of acids which are considered in this thesis and are part of this group of fatty acids are given below:



The homologous series is characterized by the general formula of  $\text{CH}_3(\text{CH}_2)_{2n}\text{COOH}$ . The straight-chain acids can have an even number of carbon atoms in the chain, in which case the acid is known as n- or even-chained. When the chain contains an odd number of carbon atoms it is known as iso- or odd-chained. The two different groups of straight-chain acids have different crystalline structures and, therefore, possess different physical properties. The saturated straight-chain acids are chemically stable along the carbon chain

and their reactive properties are mainly determined by the functional carboxyl group.

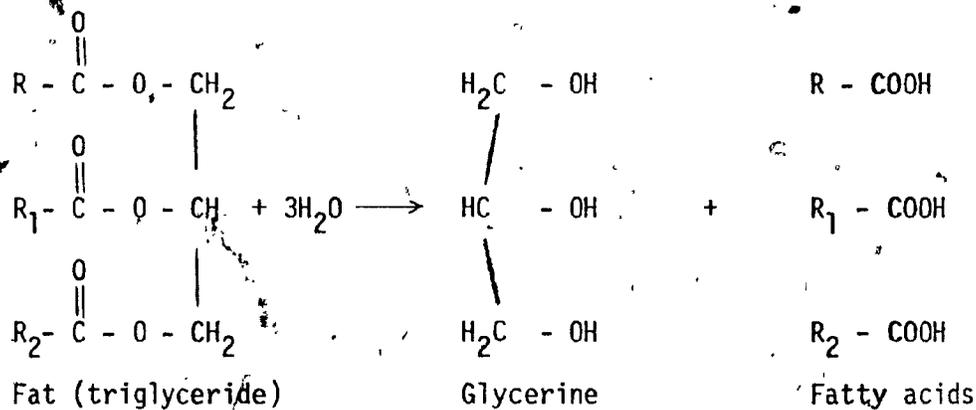
In the straight-chain structure, each carbon atom possesses at least two hydrogen atoms and no carbon atom is bonded to more than two other carbon atoms. In contrast, another group of fatty acids is characterized by double or triple bonds existing between the carbon atoms along the chain. This group is known as unsaturated fatty acids of the ethylenic family and the group consists of three separate series having the composition as shown below:



Due to the presence of the double bond between the adjacent carbon atoms along the carbon chain, the unsaturated fatty acids are chemically reactive and generally lack stability. These acids have a tendency to react with oxygen, thus making them unsatisfactory candidates for phase-change materials. The reaction generally leads to a branched-chain structure where one carbon atom along the chain can be bonded to three other carbon atoms. The double bond between the carbon atoms along the chain can in some cases alternate with single bonds creating additional groups of fatty acid families; namely, conjugated and nonconjugated. Double bonds can also produce different compounds that have the same formula, the concept known as cis and trans isomerism.

III. SOURCES AND COST OF FATTY ACIDS

Most of the fatty acids are produced commercially by hydrolysis which is a process of splitting the fat. The exception is tall oil, which is not a glyceride. The fat is normally characterized by a 3-pronged glycerol which is connected to the monobasic acids. The hydrolysis of a fat to fatty acids (Ref. 1) is illustrated below:



The naturally occurring mixed glycerides provide most of the industrial sources of fatty acids. The glycerides are available in abundant quantities; mostly from animal fat in the form of inedible tallow and vegetable sources. Table I lists the main sources of fatty acids.

TABLE I. Sources of Fatty Acids (Ref. 2)

SATURATED ACIDS	COCONUT OIL	PALM OIL	PALM KERNEL OIL	TALL OIL	BEEF TALLOW
Caproic	0.5%		0.5%		
Caprylic	8%		5%		
Capric	7%		5%		
Lauric	48%		50%		0.1%
Myristic	17%	2%	15%		3%
Palmitic	9%	42%	7%	7%	29%
Stearic	2%	4%	2%		20%

The fatty acids are widely used in the chemical industry. The main uses of fatty acids are listed in Table II.

TABLE II. End-uses of Fatty Acids (Ref. 2)

Fatty Alcohols	12%
Soaps	9%
Emulsion Polymerization	5.5%
Dimer & Trimer	5.5%
Lubricating Grease	5%
Specialty Household Cleaners	5%
Plastic Additives	4%
Food Additives	4%
Cosmetics	4%
Downstream Acids	4%
Ore Flotation	3.5%
Paint & Varnish	3.5%
Other Uses	<u>28%</u>
	100%

The North American production of fatty acids far exceeds the edible and inedible requirements and enjoys an annual growth rate of 2 to 3%. This growth rate is expected to continue for the rest of the decade. The prognosis is that the supply of fatty acids will exceed the demand for many years to come. Therefore, the fatty acid industry appears to be capable of accommodating a higher usage for

the latent heat storage technology.

The cost of fatty acids is generally more expensive than that of inorganic salt hydrates and paraffins. However, with the upsurge in demand and the improved manufacturing economics, the cost of fatty acids is expected to decline. Table III shows the current cost of fatty acids of technical grade in bulk quantities. The prices are on a per kilogram basis prevailing on the Canadian market. The associated latent heats which can be stored per dollar value are also included in the table.

TABLE III. Prices of Fatty Acids

FATTY ACID	PRICE (\$/KG)	LATENT HEAT (KJ/\$)
Capric	\$2.05	76.6
Lauric	\$1.52	107.9
Palmitic	\$1.41	114.5
Stearic	\$0.97	188.7

The cost of material is an important consideration in the selection of materials for latent heat thermal storage applications. The above table illustrates that the latent heat per unit dollar increases with the carbon chain of the fatty acid.

#### IV. DEFINITION OF TEMPERATURES AND LATENT HEATS EMPLOYED IN DIFFERENTIAL SCANNING CALORIMETRY (DSC)

An interactive DSC Data Analysis Program (V2.0) was used in conjunction with the Dupont 1090 Thermal Analyzer Micro Computer. This program calculates the temperatures and heats associated with transitions in materials.

Fig. 1 shows a typical thermogram depicting the heating and cooling cycles of a sample. Following is a list of parameters which are calculated by the interactive program with their corresponding definitions.

Melting Point: The melting point is defined as the temperature of intersection of the base line with the tangent to the inflection point having the greatest rate of change of heat flow in the heat flow versus the temperature curve for a heating cycle.

Freezing Point: The freezing point is generally affected by the sample behavior since it is not an equilibrium measure. In the absence of supercooling, the freezing point is defined as the temperature of intersection of the base line with the tangent to the inflection point having the greatest rate of change of heat flow in the heat flow versus the temperature curve for a cooling cycle.

If the sample supercools and the temperature rises by more than  $0.25^{\circ}\text{C}$  upon crystallization, then the freezing point is the temperature where the recrystallization started.

Sample: 7BL/3BP  
Size: 12.377 MG  
Rate: 2C/MIN  
Program: Interactive DSC V2.8

# DSC

Date: 17-Jun-83 Time: 13:42:58  
File: FUKS.06 DISK TWO  
Operator:  
Plotted: 28-Jun-83 22:25:38

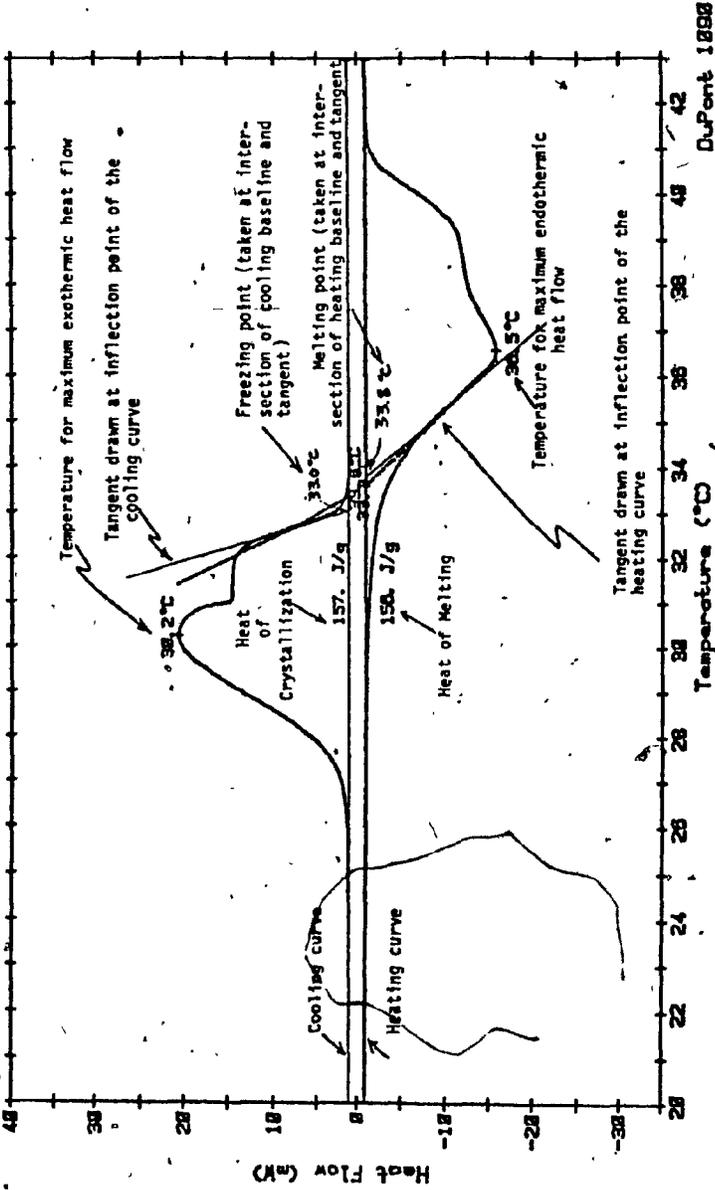


Fig. 1 Typical thermogram showing the parameters computed in DSC.

In both the melting and freezing points calculations, the program uses the least-square fitting method in drawing the tangent line. The program examines the region of the curve of interest and moves along the curve until the inflection point is located. The tangent line is then drawn at the inflection point and is projected to intersect the baseline. To fit the line, the program uses 12.5% of the total number of points between the start and stop limits.

It is apparent from the thermograms that the tangent lines drawn by the computer are skewed and are not true tangents at the inflection points. Consequently, the intersection point of the computer-drawn tangent and the base line does not represent the true transition temperature point. To correct this anomaly, the tangents at the inflection points were drawn by longhand and these points were then used in compiling the data. The fitting of the tangent line is dependent on the data scanning rate, heating or cooling rates and the mass of the sample. The computer program does not appear to be sufficiently responsive to these variables.

When several peaks are present on the thermogram, the tangent is drawn at the peak which has the greatest rate of change of heat flow.

#### Heat of Melting and Crystallization

The heats of melting and crystallization are calculated from calorimetric measurements of heat flux which are based on the following formula:

$$\Delta H = \frac{K}{m} \int_{t_1}^{t_2} \frac{\Delta \dot{Q}}{f(T)} dt$$

where:  $\Delta H$  = enthalpy  
 $m$  = sample mass  
 $\Delta Q$  = heat flux  
 $t$  = time  
 $f(T)$  = temperature dependent heat transport function  
 $K$  = calibration constant of DSC cell

The calibration constant,  $K$ , is dependent on temperature in a non-linear way. The calibration constant is linearized electronically to a temperature-independent straight line by the thermal analyzer system.

The heat is calculated as the integral of the signal over time from the start to stop points. The limit points are indicated on the thermogram by vertical tick marks which are connected by a straight dashed line. The calculated area represents the heat of transition and is reported in J/g.

#### Temperature for Maximum Endothermic Heat Flow

This parameter is defined as the temperature of the point that is located furthest from the baseline in a heating cycle.

#### Temperature for Maximum Exothermic Heat Flow

This parameter is defined as the temperature of the point that is located furthest from the baseline in a cooling cycle.

V. DIFFERENTIAL SCANNING CALORIMETRY TECHNIQUE AND APPARATUS

The pure fatty acids and their binary mixtures were investigated with a Differential Scanning Calorimeter (DSC). Thermograms of each sample were recorded on a Dupont Model 1090 thermal analyzer equipped with a 910 DSC cell. The cell contains a constantan (thermoelectric) disk as a primary heat transfer element that conducts heat to sample and reference pans positioned on the raised portions of the disk. Temperatures of the sample and reference pans are measured by the chromel-constantan area thermocouples. The differential heat flow is calculated using a known sample weight. The cell design is illustrated in Fig. 2. To provide cooling to subambient temperatures, a cooling accessory was improvised and is illustrated in Fig. 3.

The instrument was calibrated using 20 mg of Indium and a heating rate of 10°C/min as recommended by the instrument manufacturer. The observed melting point was 156.6°C with a heat of fusion of 29.1 J/g. This is in agreement with the established values set by the manufacturer. Furthermore, another calibration was performed with Naphtalene ( $C_{10}H_8$ , pure crystal, Fisher, reagent grade with a melting range of 80-81°C). A sample of 2.144 mg of Naphtalene was heated at 2°C/min and the observed melting point was 80.0°C. The sample and reference pans used in all the experiments were made of aluminum and weighed approximately 13 mg. During the experiments the pans were open (there was no need to hermetically seal the pans, as the substances which were measured are considered

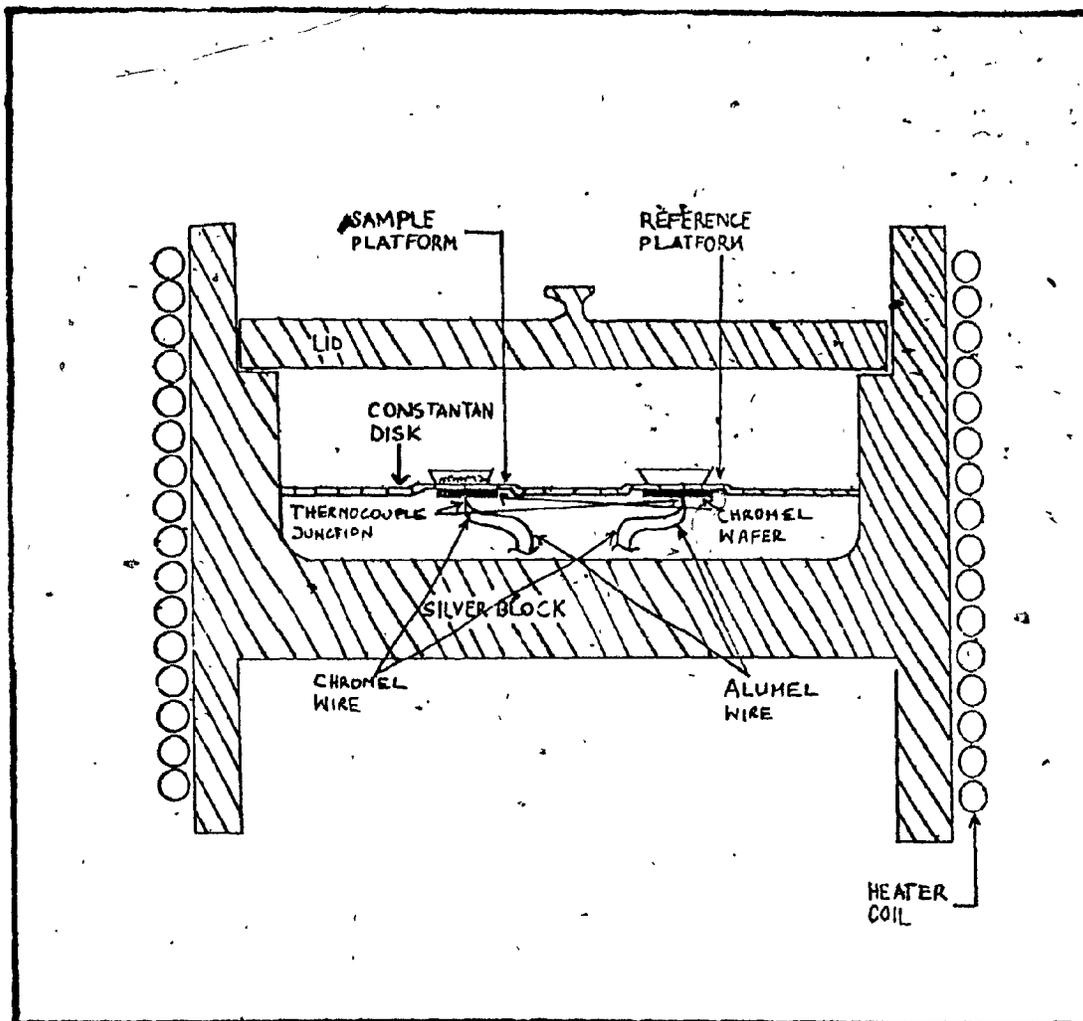


Fig. 2 DSC cell cross section

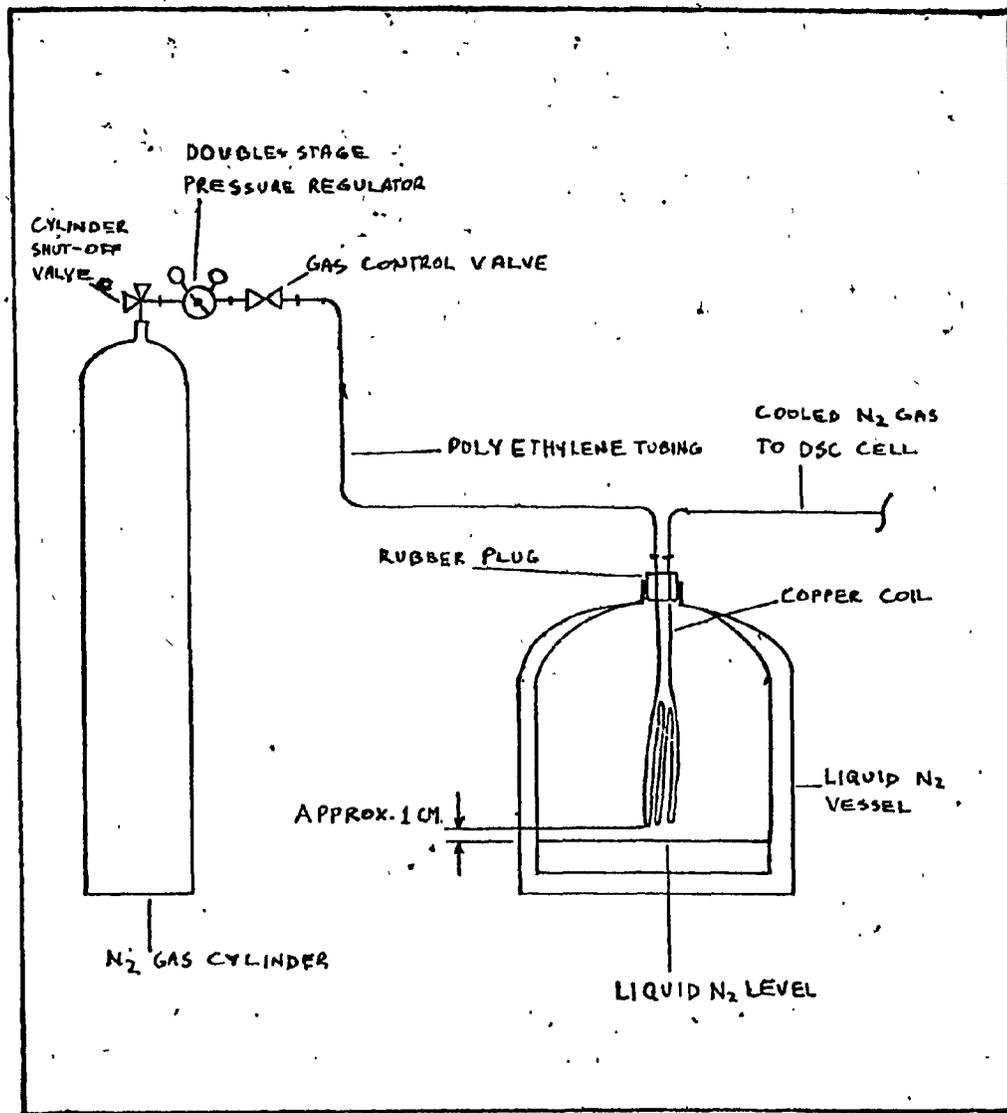


Fig. 3 Cooling accessory for DSC

non-volatile). The samples were weighed after the experiments and no weight loss was observed. The accuracy of the balance was  $\pm 0.005$  mg.

The precision of the instrument was determined by performing several melting and freezing runs on palmitic acid. The results are given in Table A-1 of the appendix. Table IV gives the mean and standard deviation of the observed parameters for melting point, temperature for maximum endothermic heat flow, heat of melting, freezing point, temperature for maximum exothermic heat flow and heat of crystallization

PARAMETER	MEAN	STANDARD DEVIATION
Melting Point ( $^{\circ}\text{C}$ )	59.0	$\pm 0.6$
Temperature for max. endothermic heat flow ( $^{\circ}\text{C}$ )	61.5	$\pm 0.1$
Heat of Melting (J/g)	175	$\pm 1$
Freezing Point ( $^{\circ}\text{C}$ )	58.1	$\pm 0.3$
Temperature for max. exothermic heat flow ( $^{\circ}\text{C}$ )	57.6	$\pm 0.7$
Heat of Crystallization (J/g)	173	$\pm 3$

TABLE IV. Calculated means and standard deviations of measurements done on palmitic acid ( The results were obtained from 4 runs at a data scanning rate of 0.4 seconds per point as listed in table A-1 of the appendix ).

The instrument manufacturer claims that the 910 DSC cell constant is linear to within  $\pm 0.3\%$ , the enthalpy coefficient of variation is less than  $\pm 1\%$  and the temperature has a repeatability of  $1^{\circ}\text{C}$ .

The experimental results indicate that the temperature measurements agree with the manufacturer's claim; however, the standard deviation of the enthalpy (heat of crystallization) is 1.7% and, therefore, slightly higher than the level claimed by the manufacturer.

An optimal heating rate for the experiments was determined by using a sample with a composition of 30 mole % capric: 70 mole % lauric at a data scanning rate of 0.4 seconds/point. The sample was subjected to heating rates from 1°C/min to 15°C/min to ascertain the melting point dependence on the heating rate. The results are tabulated in Table V and plotted in Fig. 4. The thermograms appear in Fig. T-1 to T-5 of the appendix.

Heating Rate (°C/min)	Melting Point (°C)
1	21.0
2	21.0
5	21.0
10	21.2
15	21.5

TABLE V. Melting point dependence on heating rate for a 30 mole % capric: 70 mole % lauric mixture.

An optimal heating and cooling rate of 2°C/min and a data scanning rate of 0.4 seconds per point were selected for all the experiments.



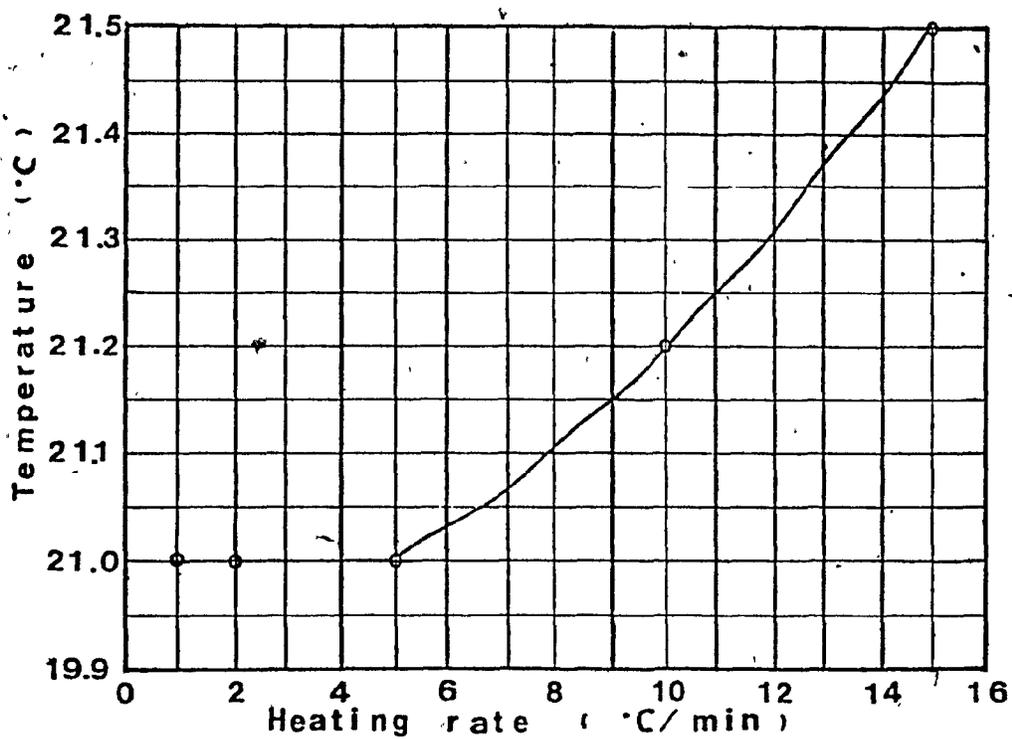


Fig. 4. Melting point dependence on heating rate for 30 mole % capric: 70 mole % lauric mixture.

VI. PHASE BEHAVIOR AND THERMAL PROPERTIES OF  
BINARY SYSTEMS OF FATTY ACIDS

Binary systems of even straight-chain fatty acids of capric-lauric, lauric-palmitic, lauric-stearic and palmitic-stearic were investigated.

The fatty acids used in the experiments were of reagent grade quality, in a flake form and obtained from commercial sources. The suppliers of the fatty acids and the melting points are listed in Table VI.

The fatty acids were mixed in a solid state in 10 molar ratios from 0 to 100% in multiples of 10% of the low to high molecular weight component. Additional molar ratios were prepared around the eutectic point to clearly establish the thermal data in this region. Subsequently, the samples were melted in an oven before being used on the DSC.

Fig. 5 illustrates the freezing behaviour of a two-component system of even straight-chain fatty acids. The binary systems of fatty acids form a single eutectic and a region with an incongruently melting solid-state compound.

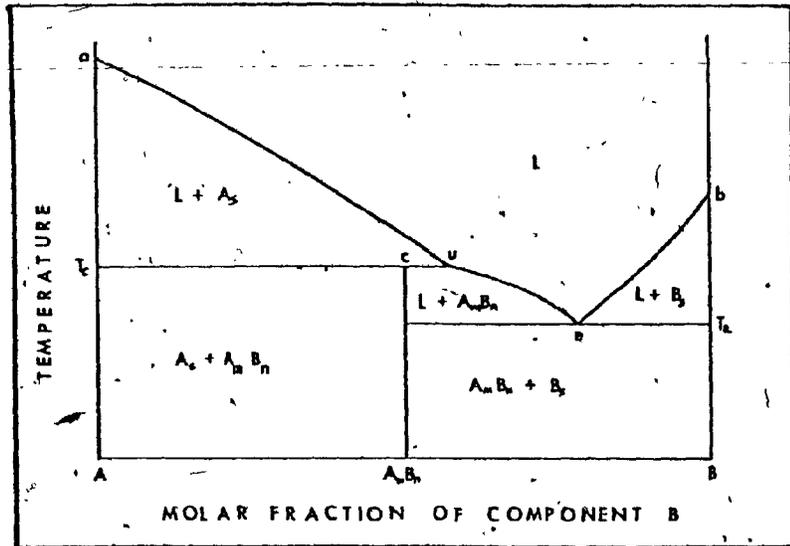
The incongruent compound transforms during melting into two phases; i.e., a higher melting solid phase and a liquid phase in a region containing an excess of the low molecular weight component.

The crystallization process of the system can be explained with the aid of the phase diagram in Fig. 5, which has been developed from freezing point relations (ref. 38 and 39). The liquid solutions, which on cooling, reach the curve line "a u" or "e b" give rise to solids  $A_s$  and  $B_s$  respectively changing the liquid composition.

5

TABLE VI. Sources of Saturated Straight-Chain Fatty Acids Used for Experiments

COMMON NAME	FORMULA	MOLECULAR WEIGHT	ACID SOURCE	MELTING POINT (°C)	MELTING POINT RANGE (°C) Ref-13
Capric	$\text{CH}_3(\text{CH}_2)_8\text{COOH}$	172.27	DX55, P5168 Matheson & Coleman & Bell Reagent	31-32	31-31.35
Lauric	$\text{CH}_3(\text{CH}_2)_{10}\text{COOH}$	200.32	Baker (7-P353) Reagent	43-46	43.92-44.5
Palmitic	$\text{CH}_3(\text{CH}_2)_{14}\text{COOH}$	256.43	Fisher Reagent (A-225)	61-62	62.53-62.82
Stearic	$\text{C}_{18}\text{H}_{36}\text{O}_2$	284.49	Fisher Reagent (A-293)	67-69	69.2-69.9



- L = liquid mixture of A and B
- $A_s$  = solid solution of the shorter-chain fatty acid in the longer-chain fatty acid
- $B_s$  = solid solution of the longer-chain fatty acid in the shorter-chain fatty acid
- $A_n B_n$  = solid compound of A and B
- a = melting point of A
- b = melting point of B
- c = composition of the solid compound
- e = eutectic point
- u = transition point
- $T_c$  = melting temperature of the compound
- $T_e$  = eutectic temperature

Fig. 5. Phase behaviour of a binary system of even straight-chain acids.

Liquid solutions, which on cooling, reach line "a e" give rise to a solid which is a compound containing an equimolar amount of the two components. Crystallization of the liquid solution occurs directly at the eutectic point with no change of composition.

For even straight-chain fatty acids which differ in chain length by two carbon atoms (capric-lauric, palmitic-stearic systems) the eutectic occurs at approximately 73 mole % of the lower-melting component. The hump in the freezing curve, representing the component formation region, occurs at about 50 to 73 mole % of the lower-melting component. These curve characteristics confirm earlier investigations by Francis et al (Ref. 3), Shuette et al (Ref. 4-8), Smith (Ref. 9), Kulka et al (Ref. 10) and Grondal et al (Ref. 11).

Table VII shows the observed eutectics of the binary systems of fatty acids. The eutectic for the capric-lauric system ( $C_{10}-C_{12}$ ) was found to have a composition of 73 mole % capric: 27 mole % lauric on both the melting and freezing curves. The melting and freezing points of the eutectic are  $18.2^{\circ}\text{C}$  and  $16.6^{\circ}\text{C}$  respectively. The freezing point is slightly lower than the melting point due to the effect of supercooling. Kulka and Sandin (Ref. 10) found the eutectic of the  $C_{10}-C_{12}$  system to be  $19.6^{\circ}\text{C}$  at a composition of 72.5 mole % capric 27.5 mole % lauric using the capillary tube method. The small difference in the melting point can be ascribed to the impurities found in the reagent grade substances. Kulka and Sandin used highly purified acids which were repeatedly recrystallized from acetone.

The data for the melting and freezing points, latent heats of transition and temperatures for maximum exothermic and endothermic

heat flow for the C<sub>10</sub>-C<sub>12</sub> binary system have been plotted in Fig. 6 to Fig. 11.

TABLE VII. Observed eutectics of binary systems of fatty acids (Taken from phase diagrams).

PARAMETER	LAURIC-CAPRIC SYSTEM	LAURIC-PALMITIC SYSTEM	LAURIC-STEARIC SYSTEM	PALMITIC-STEARIC SYSTEM
Melting Point (°C)	18.2 @ 73C/2L	32.7 @ 80L/20P	34.0 85L/15S	51.1 @ 72.5P/27.5S
Temperature for max. endothermic flow (°C)	20.8 @ 73C/27L	35.6 @ 80L/20P	36.7 @ 85L/15S	53.8 @ 72.5P/27.5S
Heat of Melting (J/g)	120 @ 72.5C/27.5L	147 @ 80L/20P	152 @ 86L/14S	159 @ 71P/29S
Freezing Point (°C)	16.6 @ 73C/27L	30.3 @ 80L/20P	31.3 @ 86L/14S	50.8 @ 72.5P/27.5S
Temperature for max. exothermic heat flow (°C)	16.2 @ 72.5C/27.5L	29.1 @ 80L/20P	30.8 @ 86L/14S	49.7 @ 72.5P/27.5S
Heat of Crystallization (J/g)	113 @ 72.5C/27.5L	149 @ 73 L/21P	155 @ 86L/14S	161 @ 72.5P/27.5S

The profiles of the curves for the temperature for maximum exothermic heat flow, heat of melting and heat of crystallization resemble the profile of the curve for freezing points.

This is in agreement with the freezing curve profile developed by Kulka and Sandin. However, the profiles of the curves for melting points and maximum endothermic heat flow points differ in shape from the freezing curve. This occurs in the concentration

range of about 0-40 mole % capric acid and to a lesser degree in the compound formation region of about 40-73 mole % capric acid. In this concentration range, the melting points and the maximum endothermic heat flow points are lower than their corresponding freezing and maximum exothermic heat flow points. This difference in temperature can be explained in terms of the existence of multiple peaks on the thermograms and the method by which these parameters are calculated.

In the DSC calculation, the melting point is located at the peak which possesses the greatest rate change of heat flow. When a mixture contains two components, several peaks are present on the thermogram and the melting point and the maximum endothermic heat flow point can be located at different peaks depending on the concentration of the two components. The shifting of the melting point and the maximum heat flow point between the peaks can result in marked differences in these parameters. For example, for the mixture containing 10 mole % capric: 90 mole % lauric the melting point and the maximum endothermic heat flow point are 35.7°C and 40.1°C respectively and both are located on the second peak of the thermogram. However, for the next higher mixture containing 20 mole % capric: 80 mole % lauric the melting point is 20.9°C, having shifted to the first peak and the maximum endothermic heat flow point is 27.0°C continuing to remain on the second peak. Furthermore, going to the next higher concentration of 30 mole % capric: 70 mole % lauric, the melting point and the maximum endothermic heat flow point of the mixture are 20.4°C and 22.3°C respectively, both of which are found

on the first peak. As can be seen, the shifting between peaks can produce marked temperature differences in adjacent mixtures having different concentrations of the two components. This is also illustrated by the curves of these parameters which show that when shifting occurs between the peaks, the points generally do not fall directly on the curve. The significance of the peaks is discussed in a later section titled "Discussion of Thermograms of Binary Mixtures of Fatty Acids".

The eutectic composition for the palmitic-stearic ( $C_{16}-C_{18}$ ) system was found to have a concentration of 72.5 mole % palmitic: 27.5 mole % stearic. The melting and freezing points at this concentration are  $51.1^{\circ}\text{C}$ . and  $49.7^{\circ}\text{C}$  respectively. Francis et al (Ref. 3) have studied the  $C_{16}-C_{18}$  system and found the same composition of the eutectic point having a freezing point of  $54.5^{\circ}\text{C}$ . Another study of the  $C_{16}-C_{18}$  system was done by Shriner et al (Ref. 12) who also found the eutectic to have the same composition with a freezing point of  $53.6^{\circ}\text{C}$ . Therefore, the eutectic location appears to be in agreement with the published data. The small difference in the freezing point temperature found by the various workers can be attributed to the presence of reagent impurities which can produce a freezing point depression. The  $C_{16}-C_{18}$  binary system, similar to the  $C_{10}-C_{12}$  system which differs in chain length by two carbon atoms, contains a hump in the freezing curve in the same concentration range of 50-73 mole % palmitic acid demarcating the compound formation region. The data for the freezing and cooling parameters of the  $C_{16}-C_{18}$  system is represented graph-

ically in Fig. 12 to Fig. 17.

For even straight-chain fatty acids which differ by more than two carbon atoms in the chain length such as lauric-palmitic ( $C_{12}$ - $C_{16}$ ) and lauric-stearic ( $C_{12}$ - $C_{18}$ ) systems, the eutectic location is shifted to a higher molar concentration of the lower-molecular weight component. Also, the compound formation region is displaced to a concentration range containing a higher molar concentration of the lower-molecular weight component.

For the  $C_{12}$ - $C_{16}$  binary system, the eutectic occurs at a concentration of 80 mole % lauric: 20 mole % palmitic. The melting and freezing points at this concentration are 32.7°C and 30.3°C respectively. This is in agreement with the results obtained by Waentig and Pescheck (Ref. 13) and Efremov et al (Ref. 13) who observed the eutectic at 80 mole % lauric: 20 mole % palmitic and 77.5 mole % lauric: 22.5 mole % palmitic respectively. The corresponding freezing points were 35.5°C and 31°C respectively. The small difference in freezing temperatures which were determined by the different workers can be explained by the degree of impurities found in the reagent materials. The results of the thermal experiments for the  $C_{12}$ - $C_{16}$  binary system are depicted graphically in Fig. 18 to Fig. 23.

For the  $C_{12}$ - $C_{18}$  binary system, the melting point of the eutectic was found to be 34°C, having a concentration of 85 mole % lauric: 15 mole % stearic. The freezing point of the eutectic was found to be 31.3°C at a concentration of 86 mole % lauric: 14 mole % stearic. This agrees with the eutectic composition of 82.5 mole % lauric:

12.5 mole % stearic having a freezing point of 36.5°C. The slight difference in the eutectic composition and freezing points are due to impurities in the reagent materials. The thermal data for the C<sub>12</sub>-C<sub>18</sub> binary system is shown graphically in Fig. 24 to Fig. 29.

The results of all the DSC runs on C<sub>10</sub>-C<sub>12</sub>, C<sub>12</sub>-C<sub>16</sub>, C<sub>12</sub>-C<sub>18</sub> and C<sub>16</sub>-C<sub>18</sub> are tabulated in the appendix tables A-2, A-3, A-4 and A-5 respectively. Thermograms of the samples, T-6 to T-81, also appear in the appendix.

FIG.6 MELTING POINTS OF BINARY SYSTEM  
CAPRIC - LAURIC ACIDS

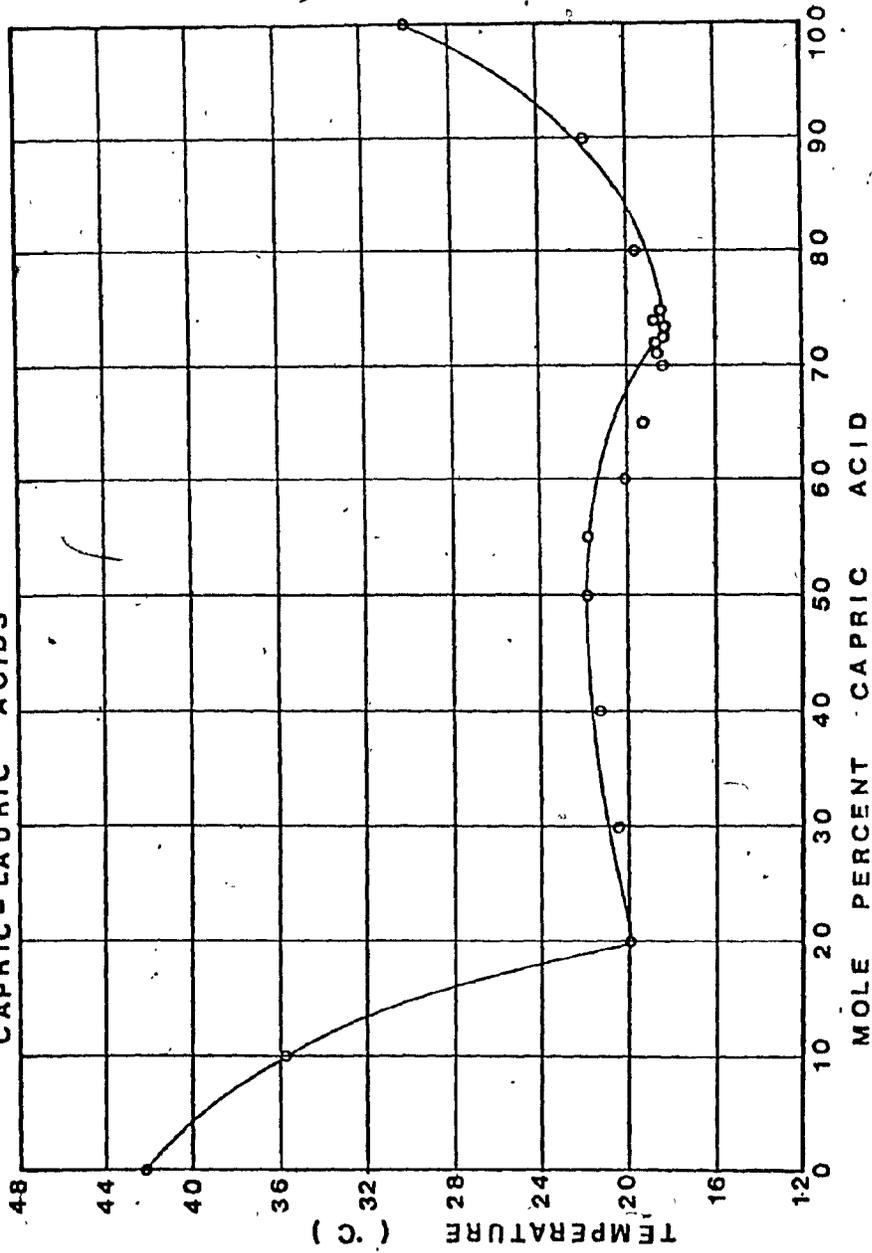


FIG.7 FREEZING POINTS OF BINARY SYSTEM  
CAPRIC-LAURIC ACIDS

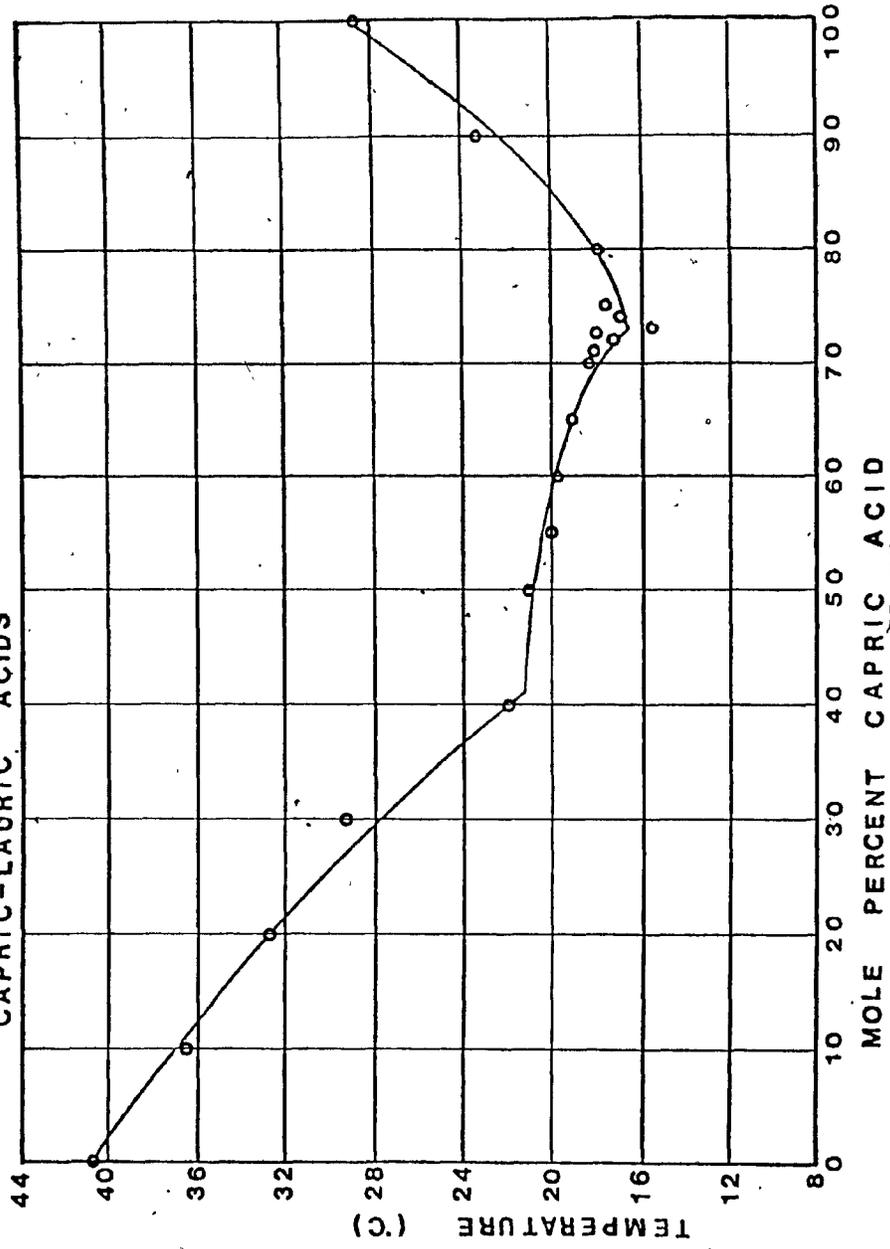


FIG.8 HEAT OF CRYSTALLIZATION OF BINARY SYSTEM CAPRIC-LAURIC ACIDS

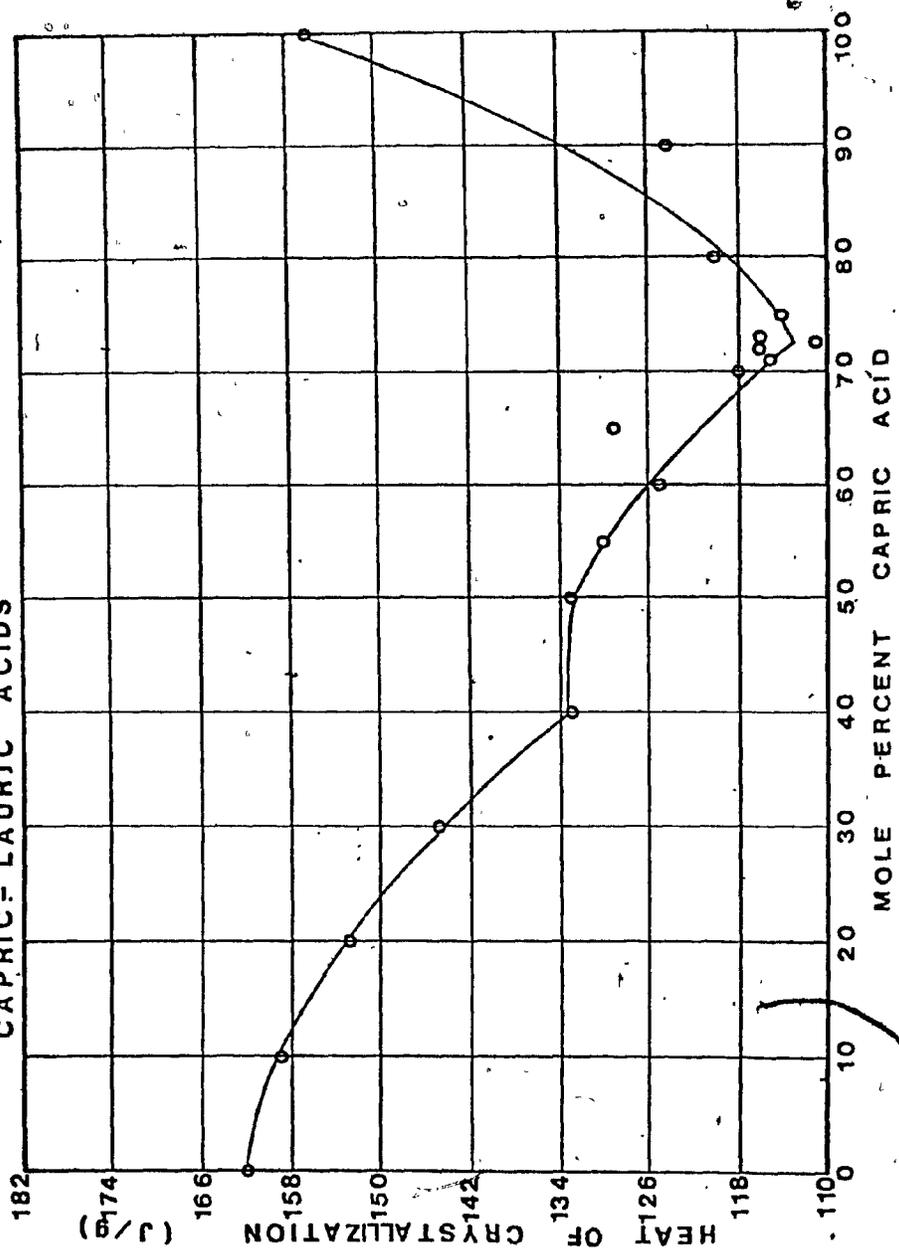
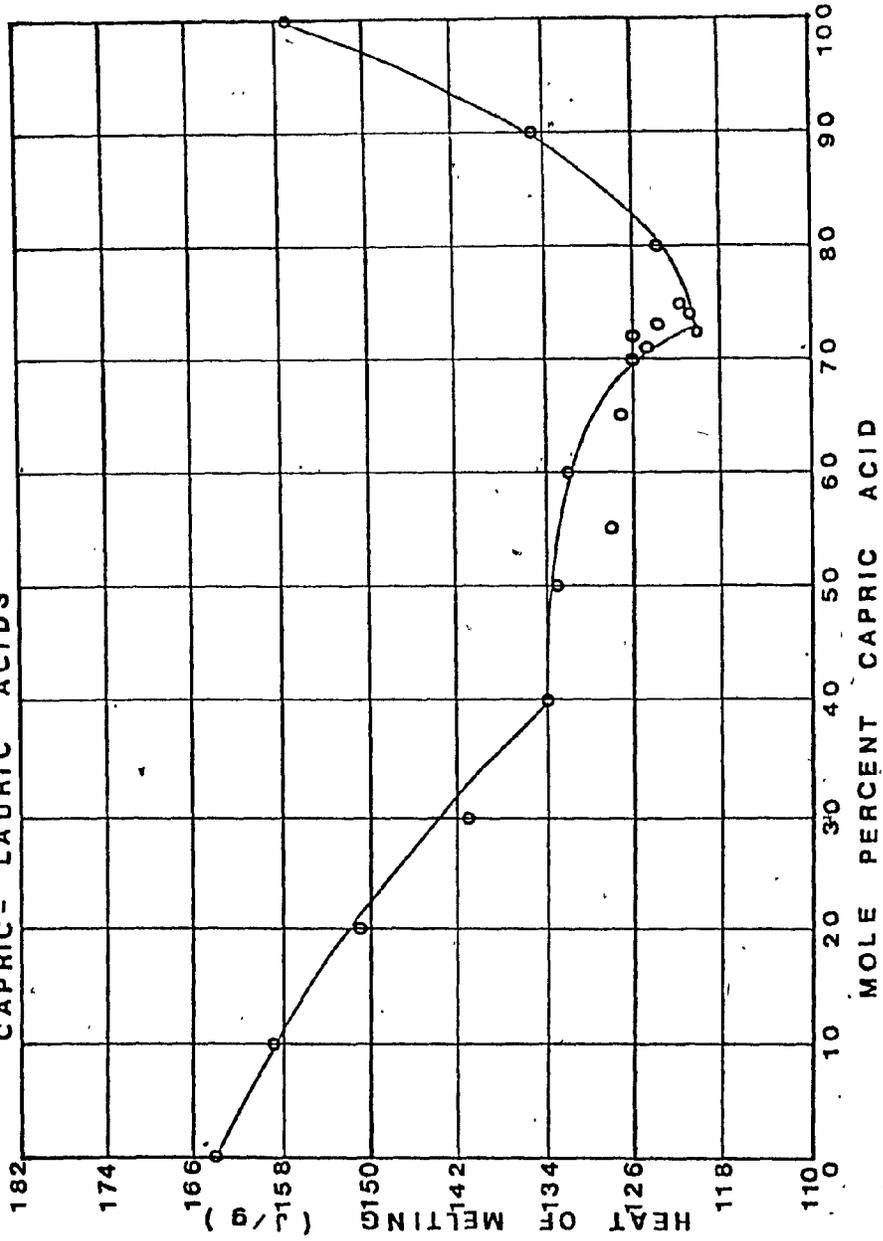
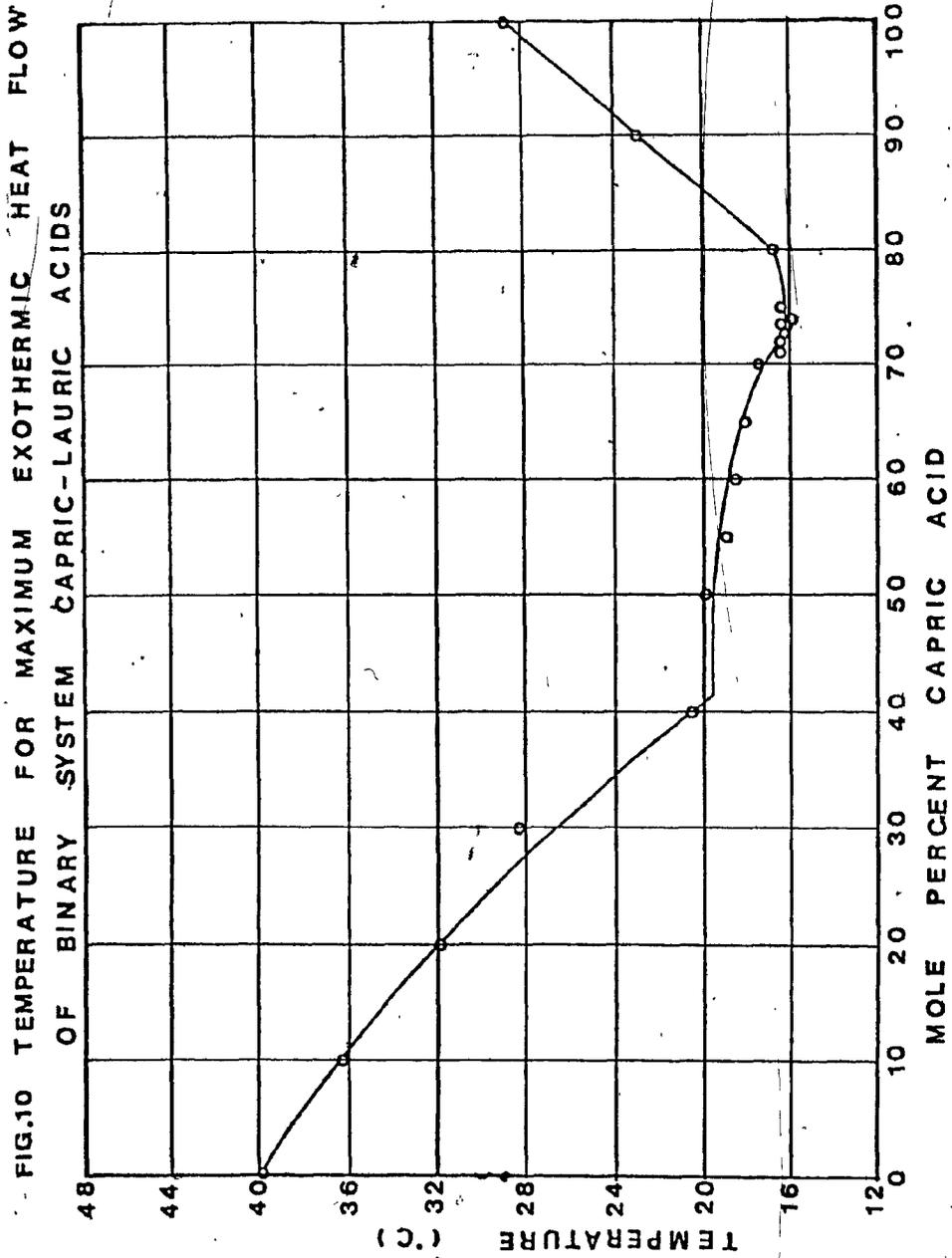


FIG.9 HEAT OF MELTING OF BINARY SYSTEM  
CAPRIC- LAURIC ACIDS





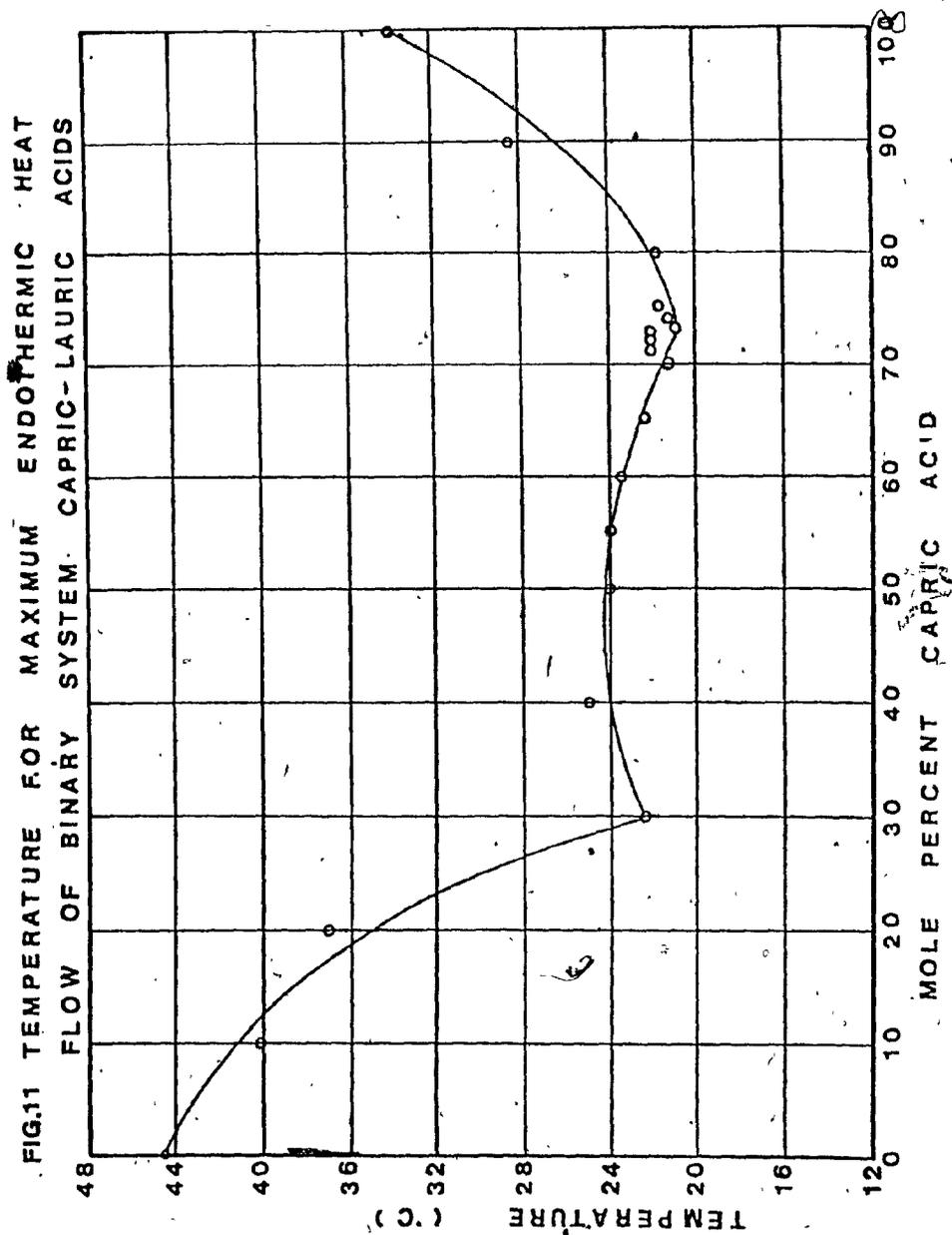
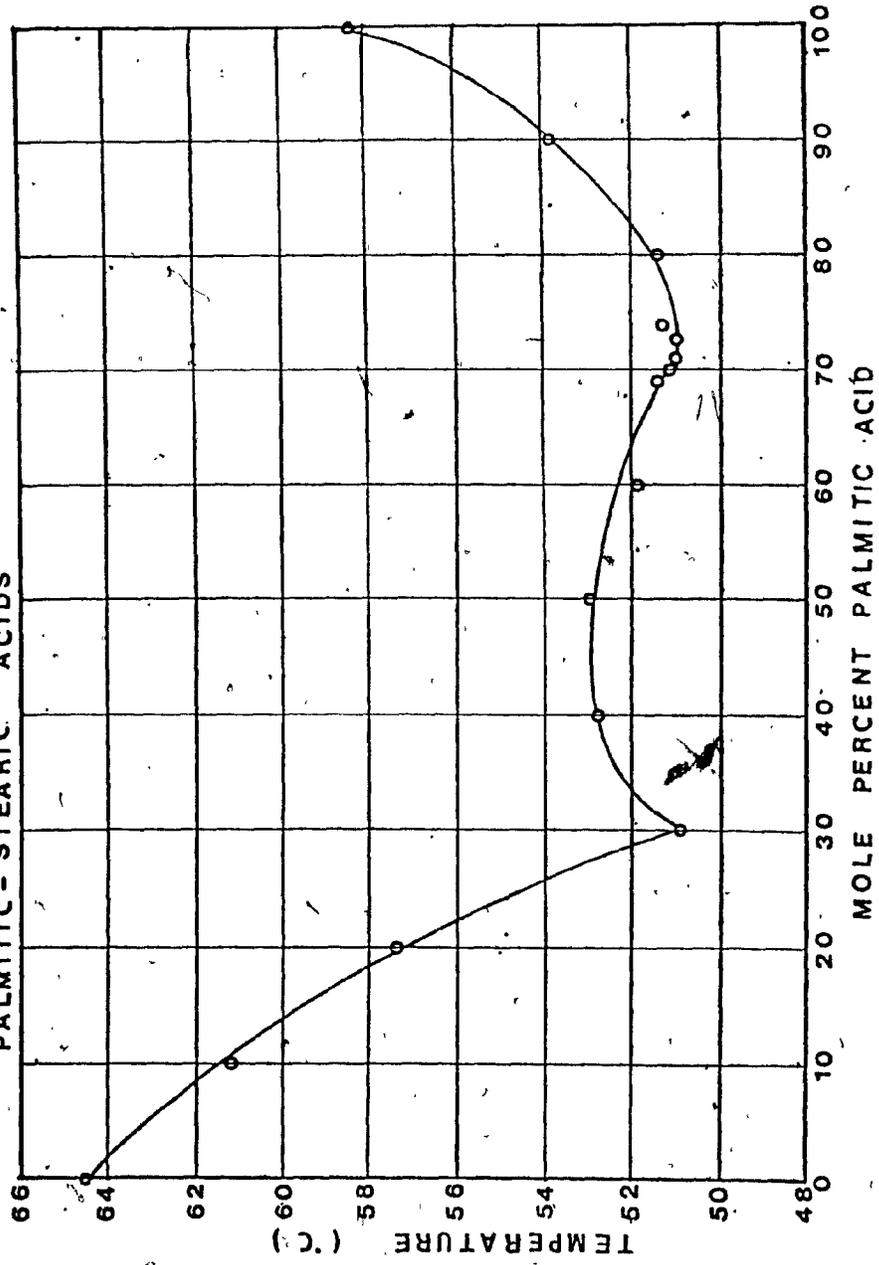


FIG.12 MELTING POINTS OF BINARY SYSTEM  
PALMITIC - STEARIC ACIDS



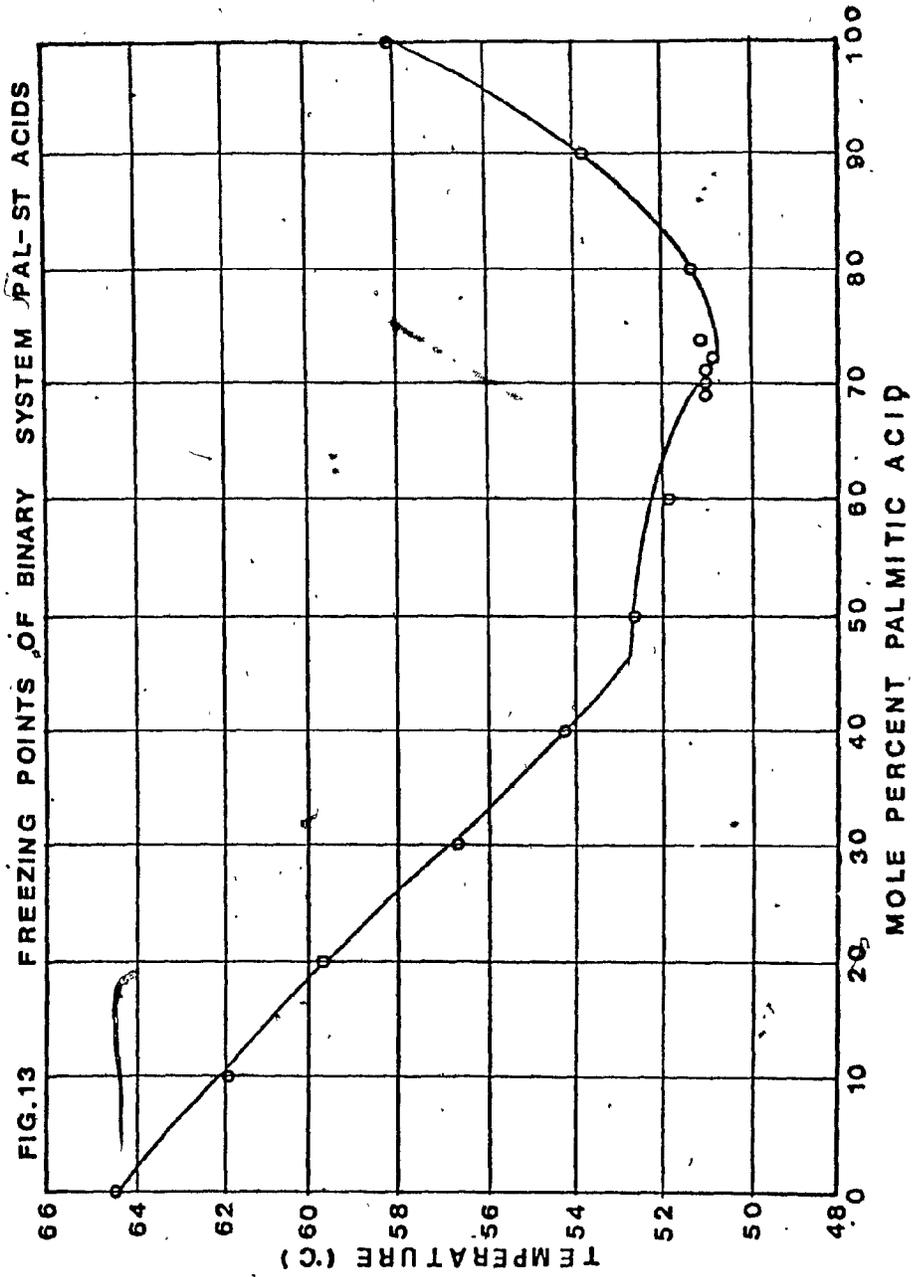


FIG.14 HEAT OF CRYSTALLIZATION OF BINARY SYSTEM  
PALMITIC-STEARIC ACIDS

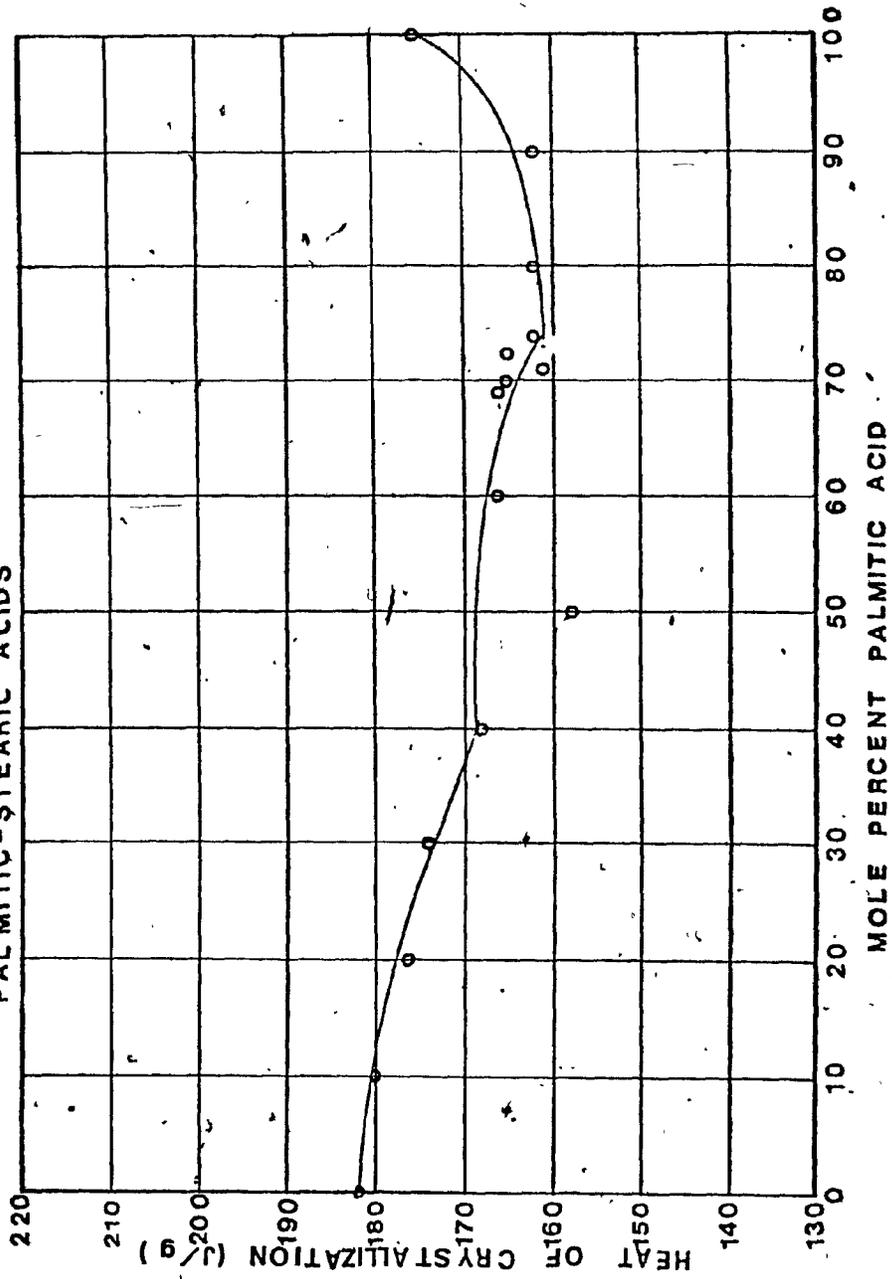
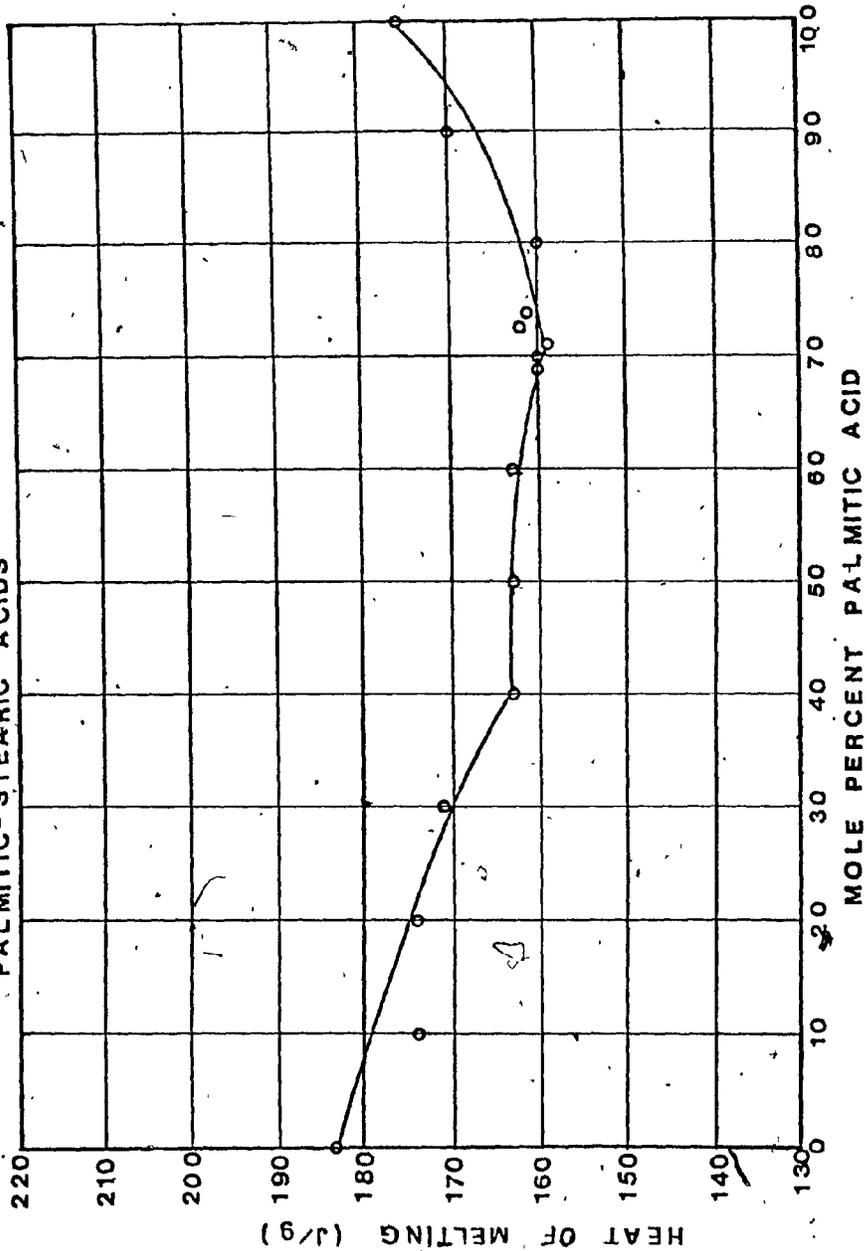
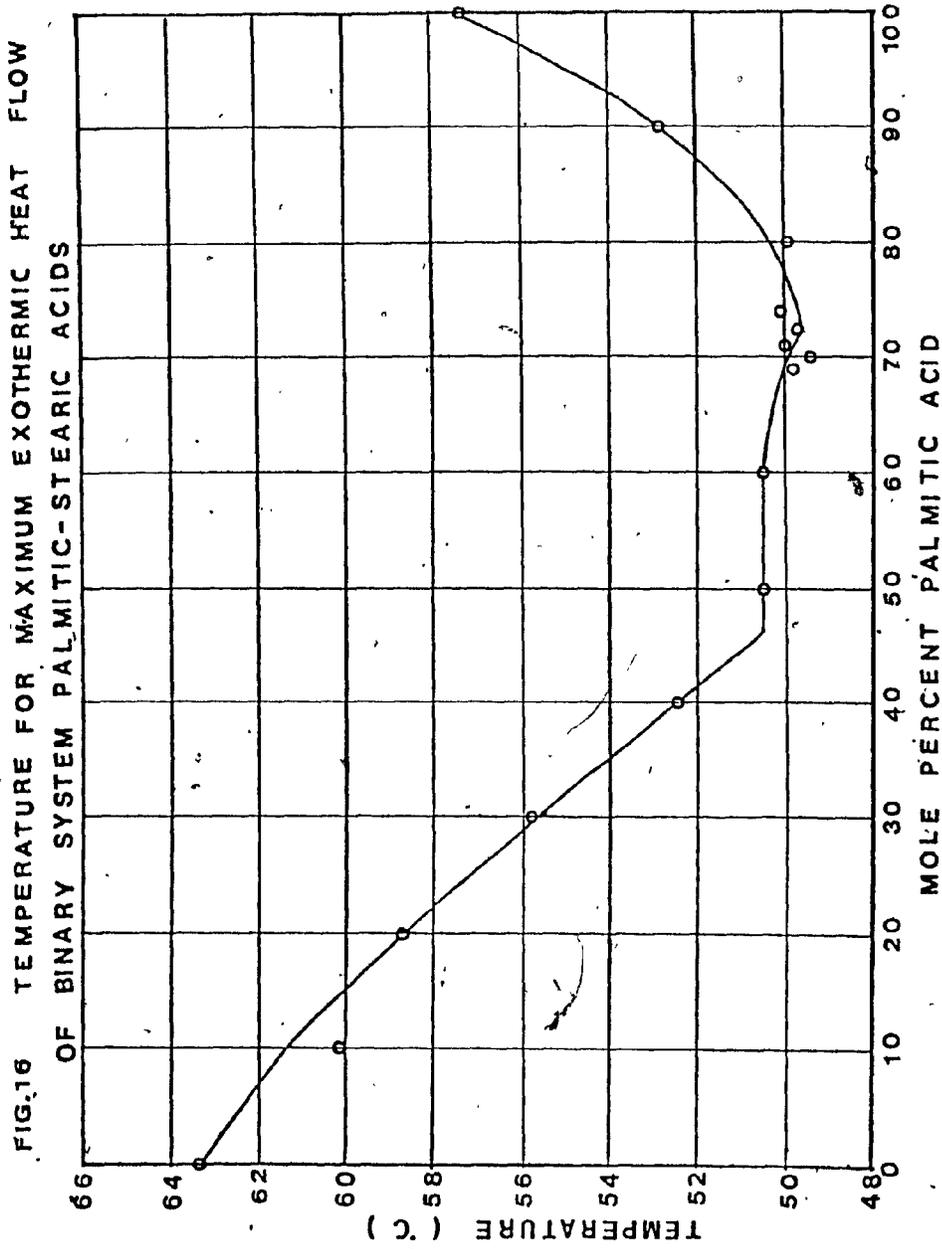


FIG.15 HEAT OF MELTING OF BINARY SYSTEM  
PALMITIC-STEARIC ACIDS





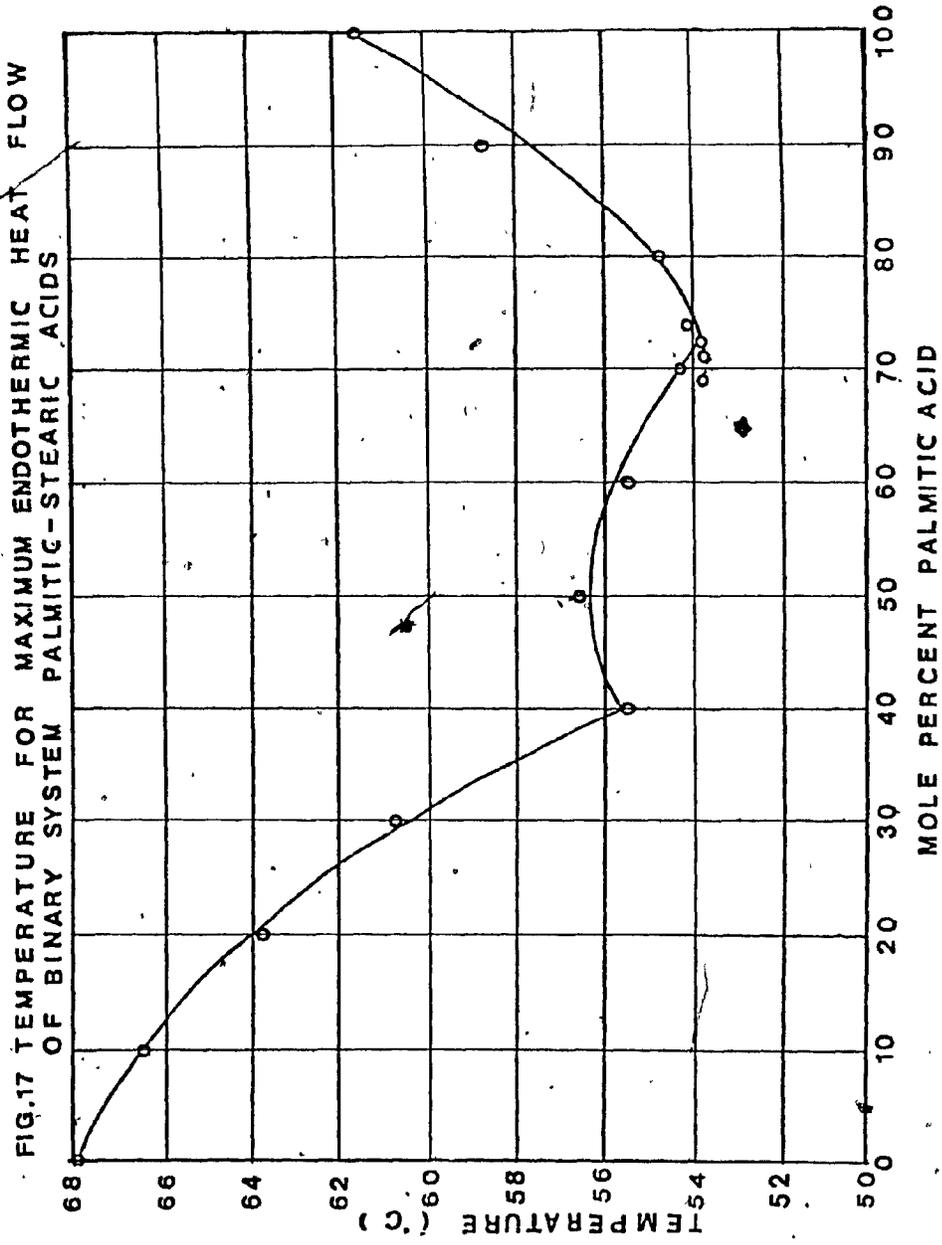


FIG.18 MELTING POINTS OF BINARY SYSTEM  
LAURIC-PALMITIC ACIDS

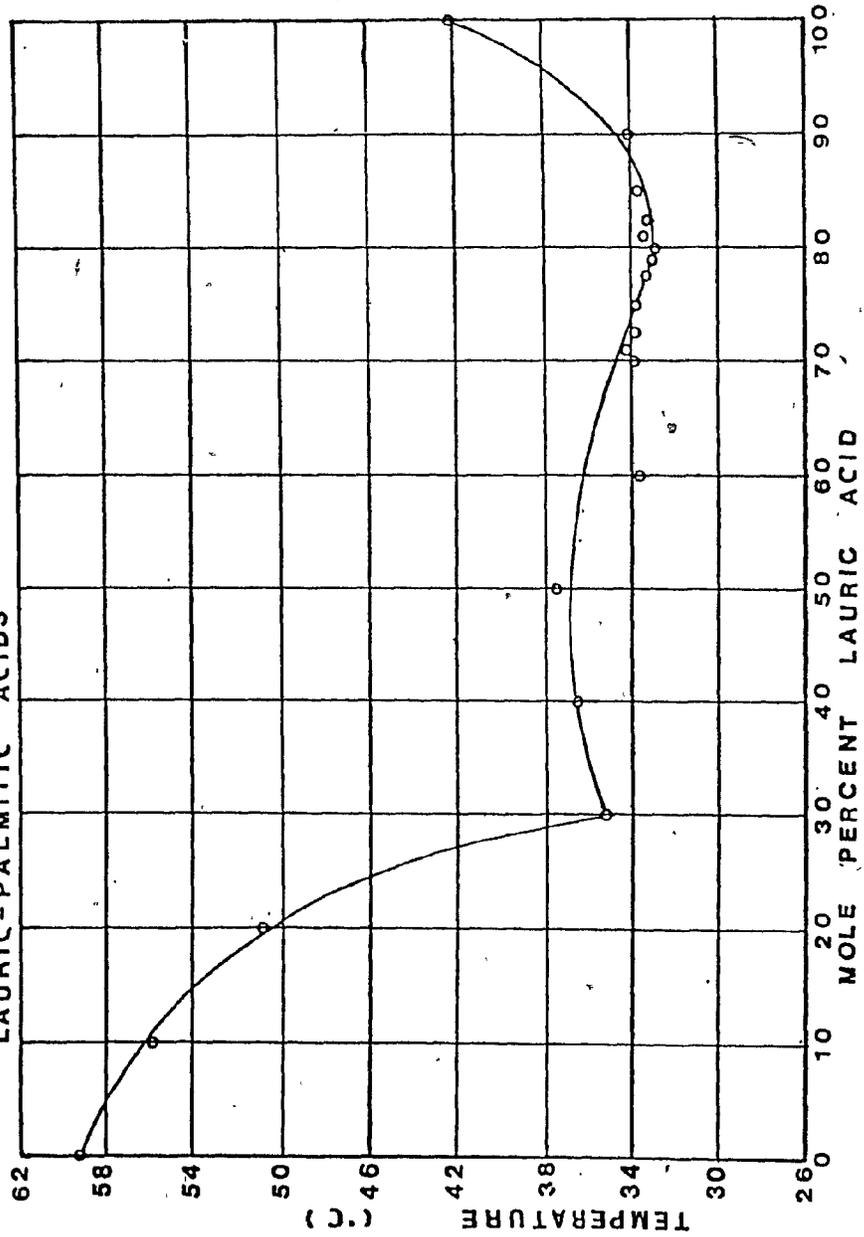


FIG.19 FREEZING POINTS OF BINARY SYSTEM  
LAURIC - PALMITIC ACIDS

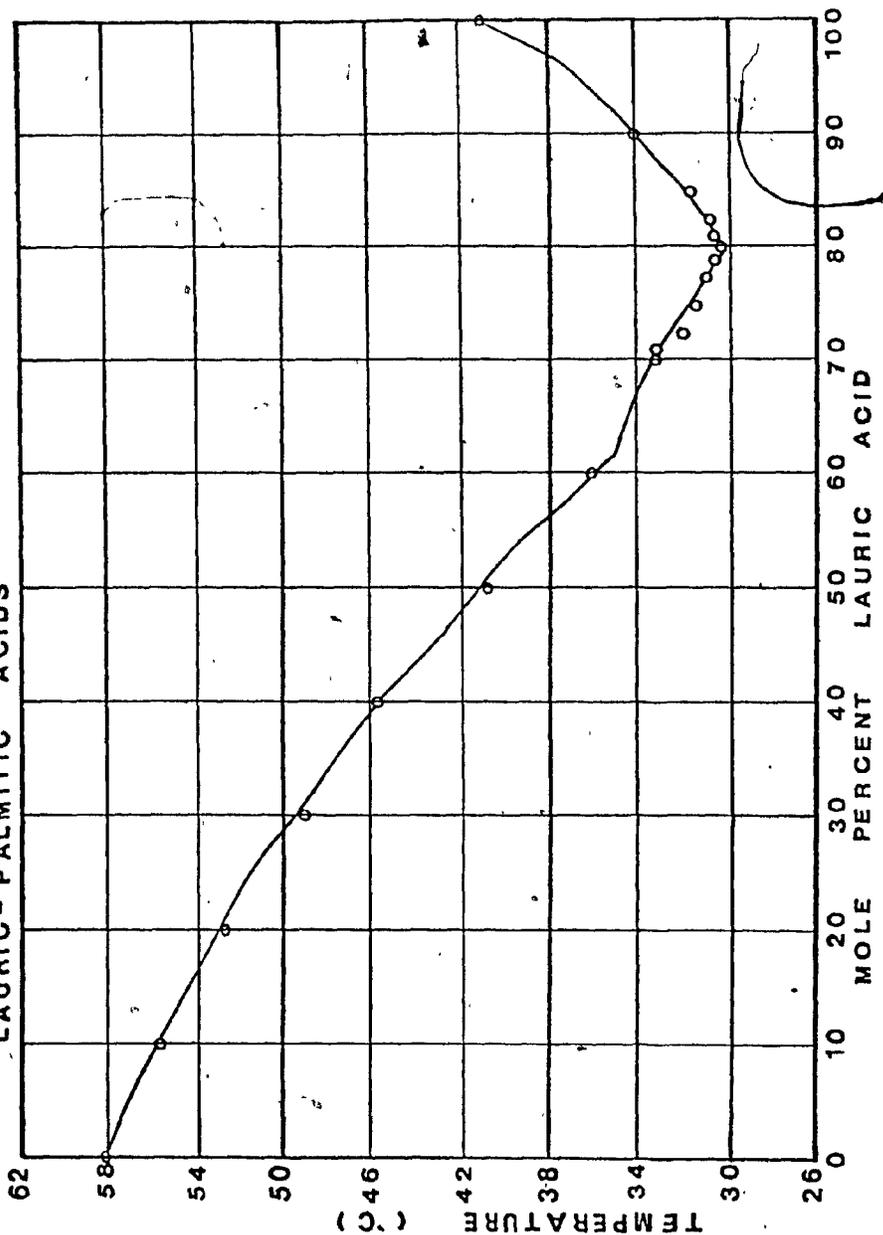


FIG.20 HEAT OF CRYSTALLIZATION OF BINARY SYSTEM,  
LAURIC-PALMITIC ACIDS

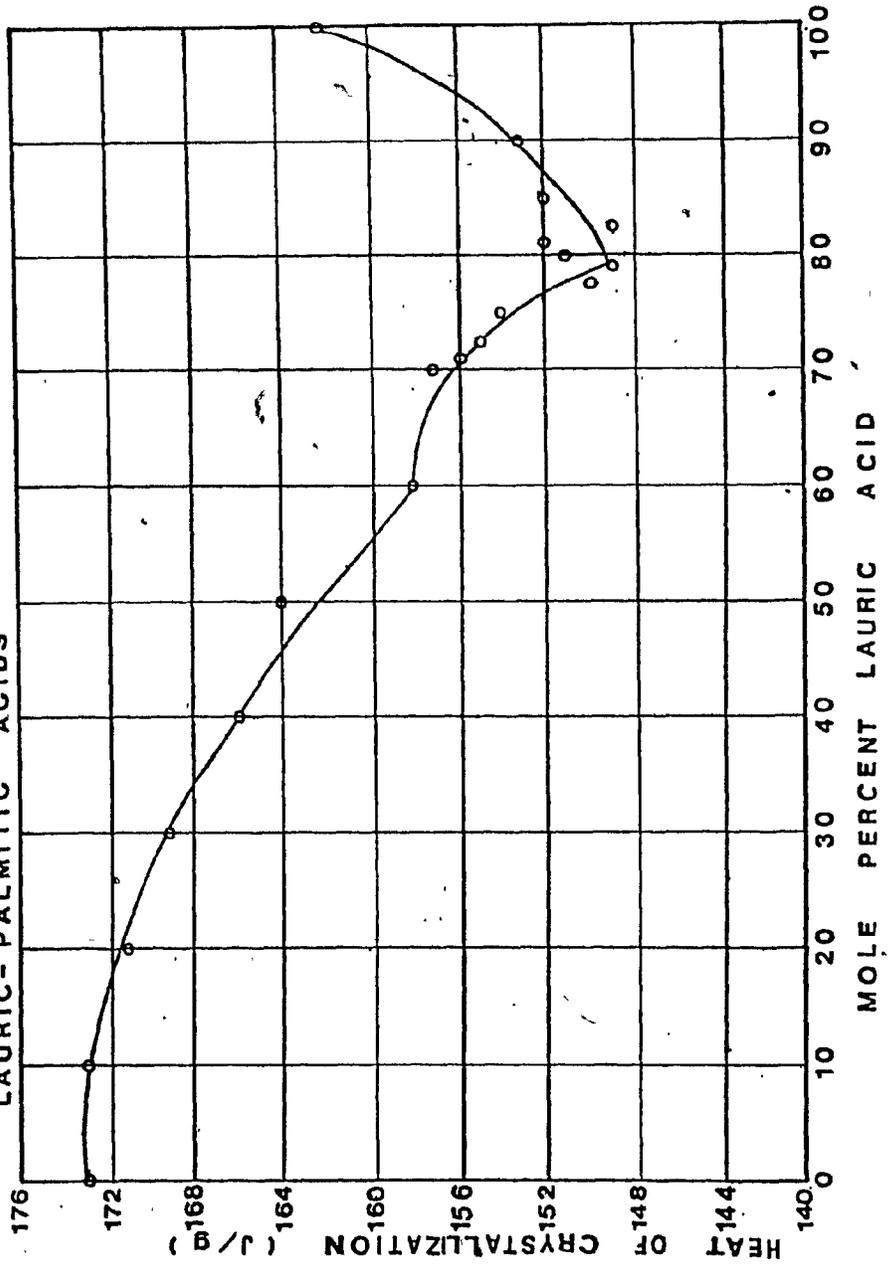


FIG.21 HEAT OF MELTING OF BINARY SYSTEM  
LAURIC- PALMITIC

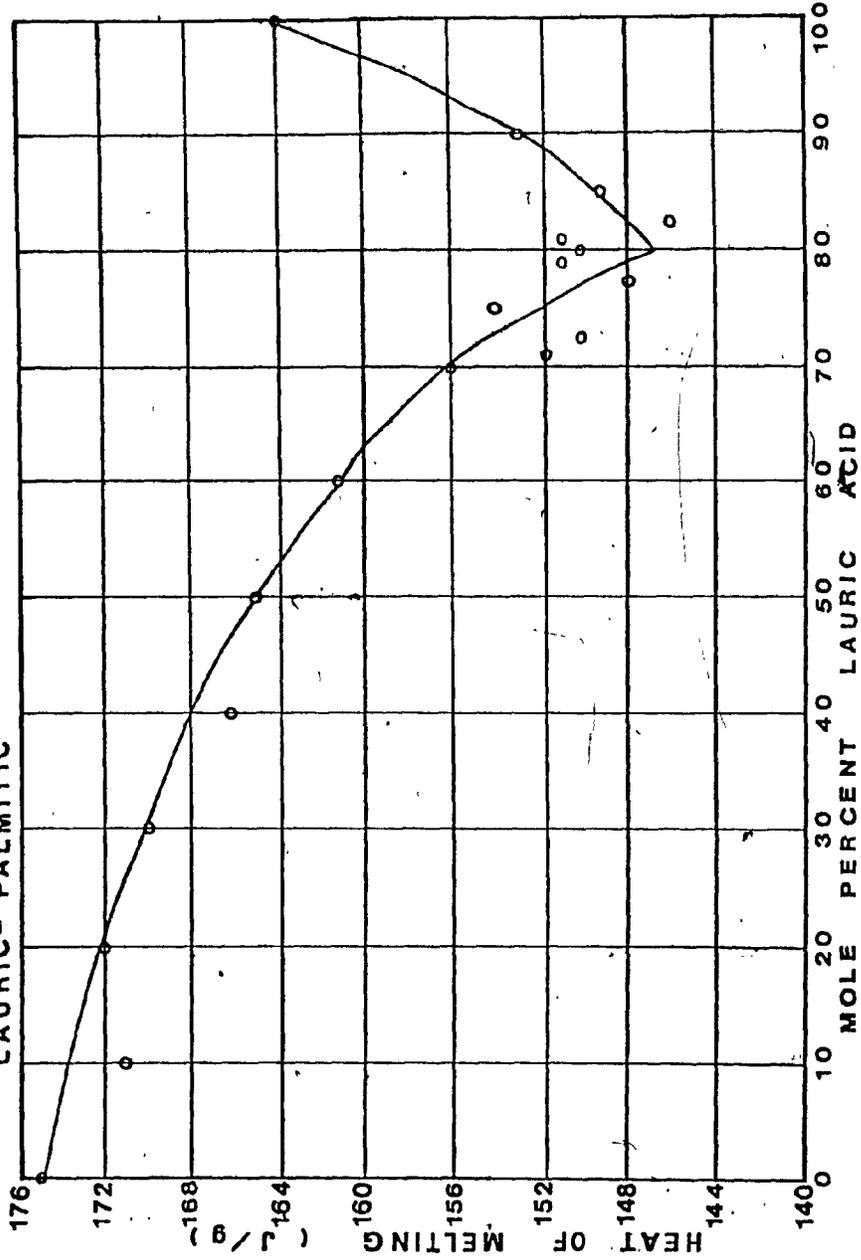
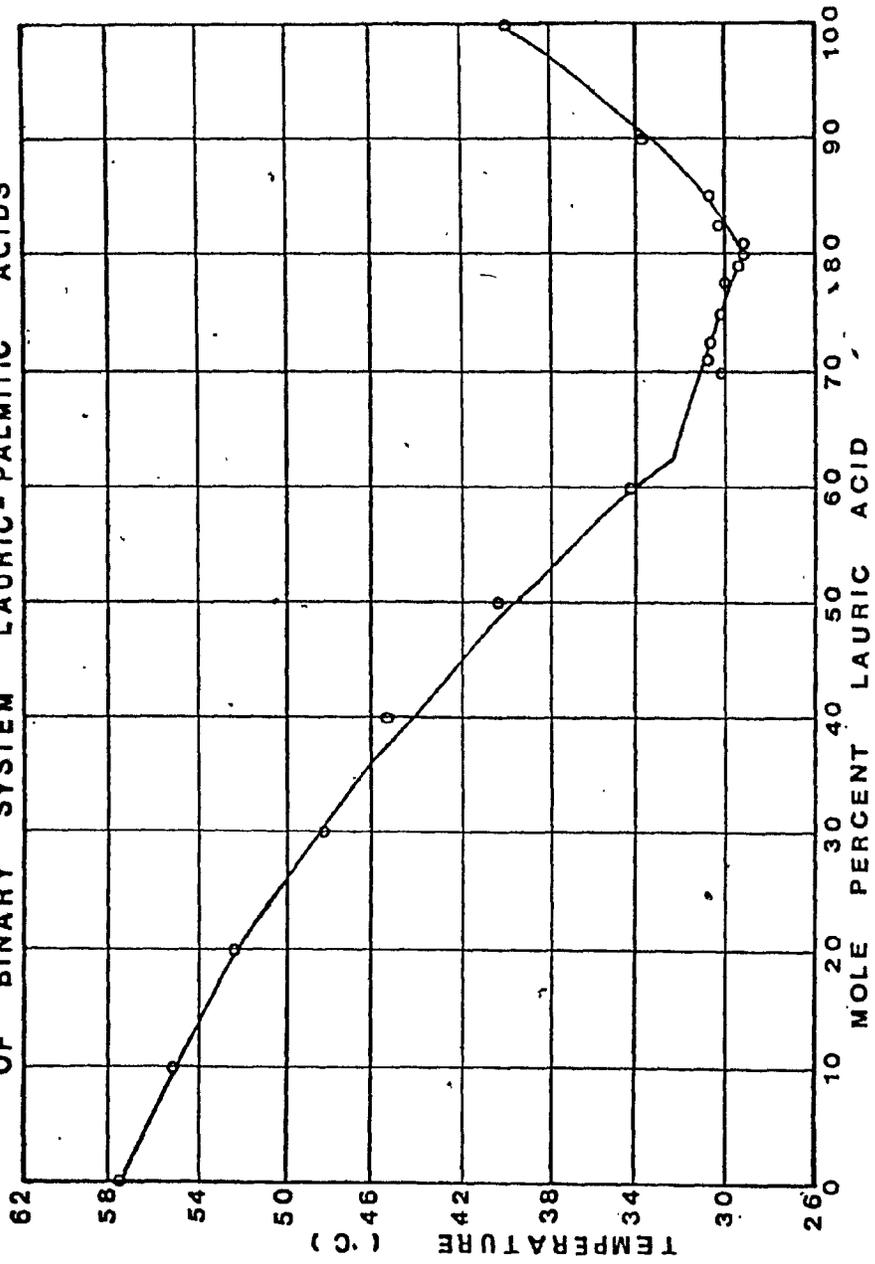


FIG.22 TEMPERATURE FOR MAXIMUM EXOTHERMIC HEAT FLOW OF BINARY SYSTEM LAURIC-PALMITIC ACIDS



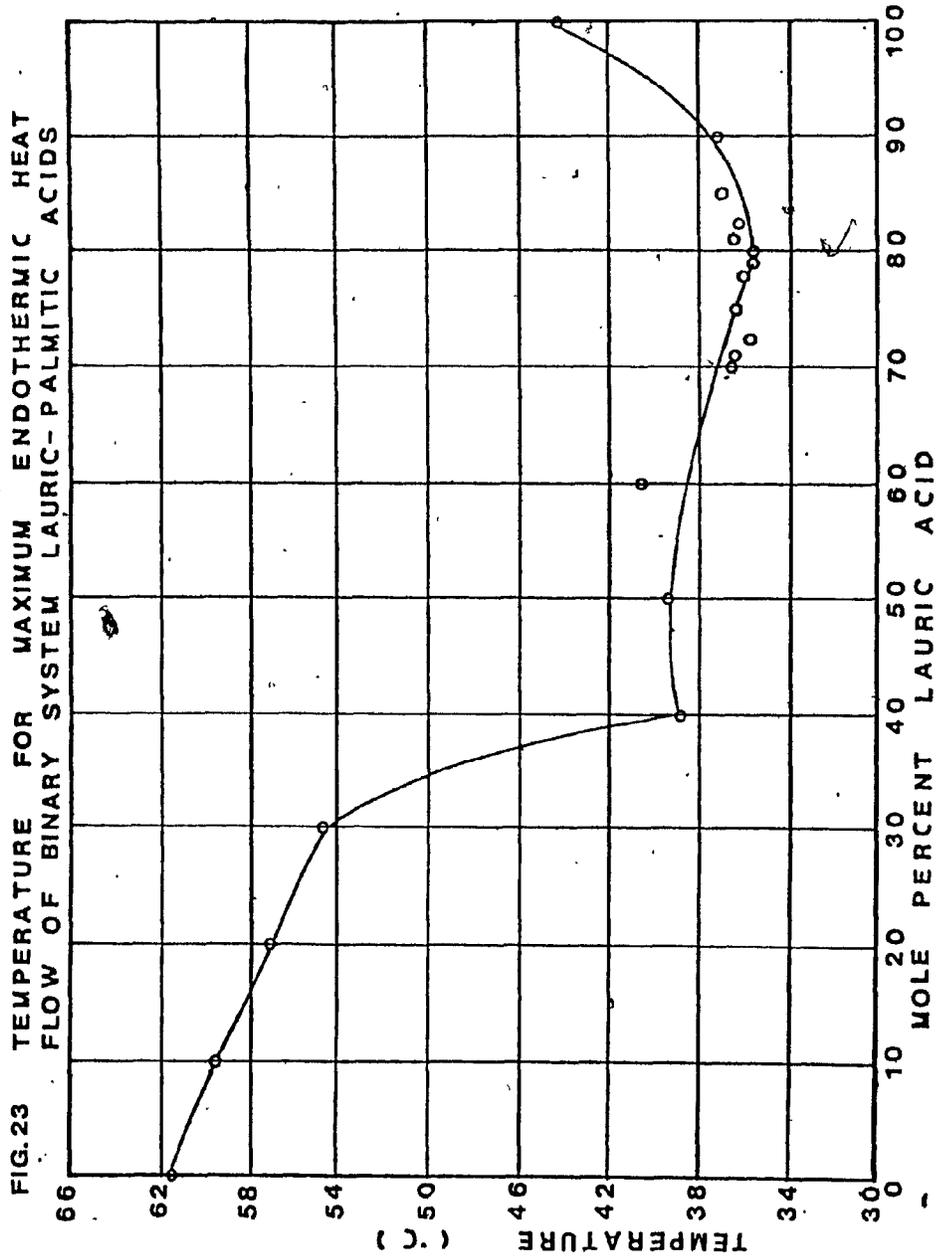


FIG.24 MELTING POINTS OF BINARY SYSTEM  
LAURIC-STEARIC ACIDS

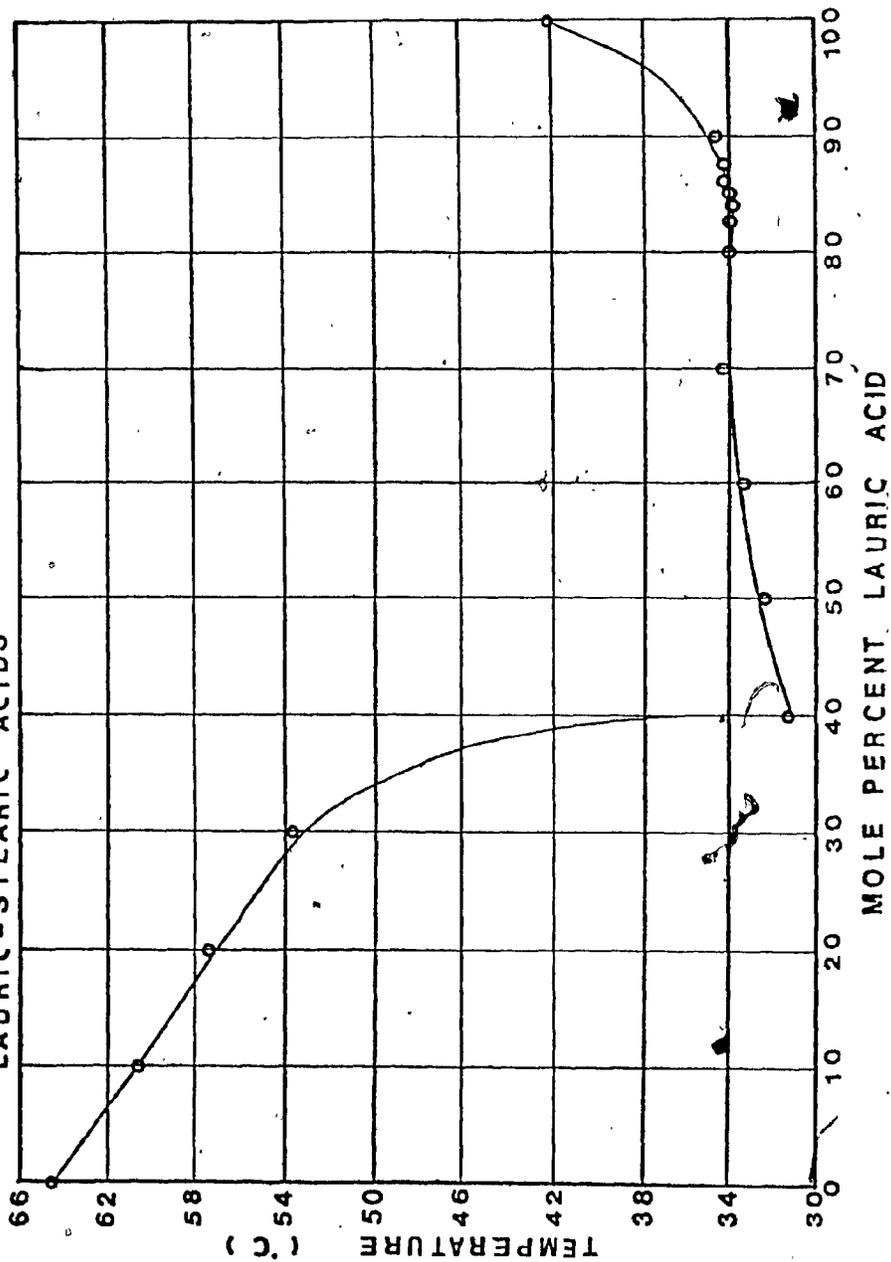
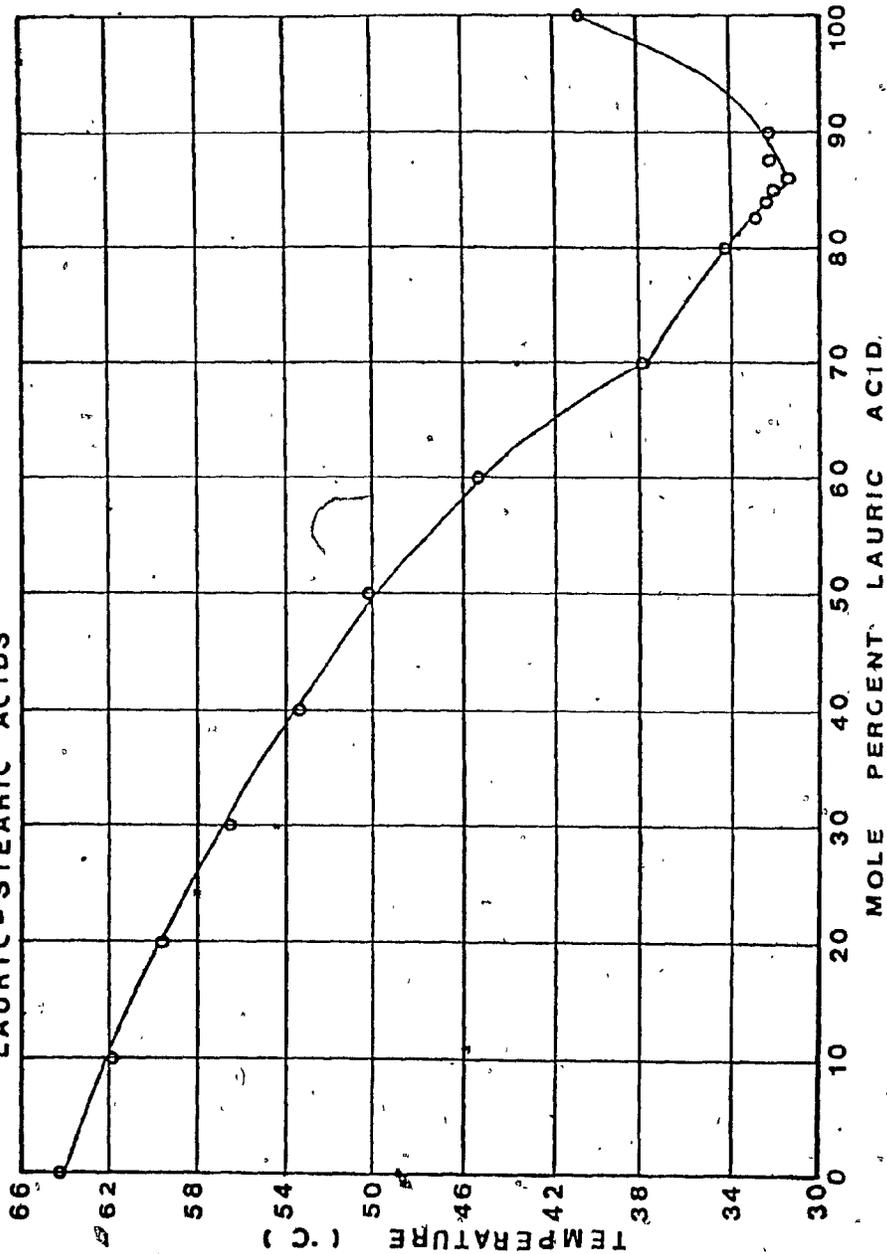


FIG.25 FREEZING POINTS OF BINARY SYSTEM  
LAURIC-STEARIC ACIDS



2

FIG.26 HEAT OF CRYSTALLIZATION OF BINARY SYSTEM  
LAURIC-STEARIC ACIDS

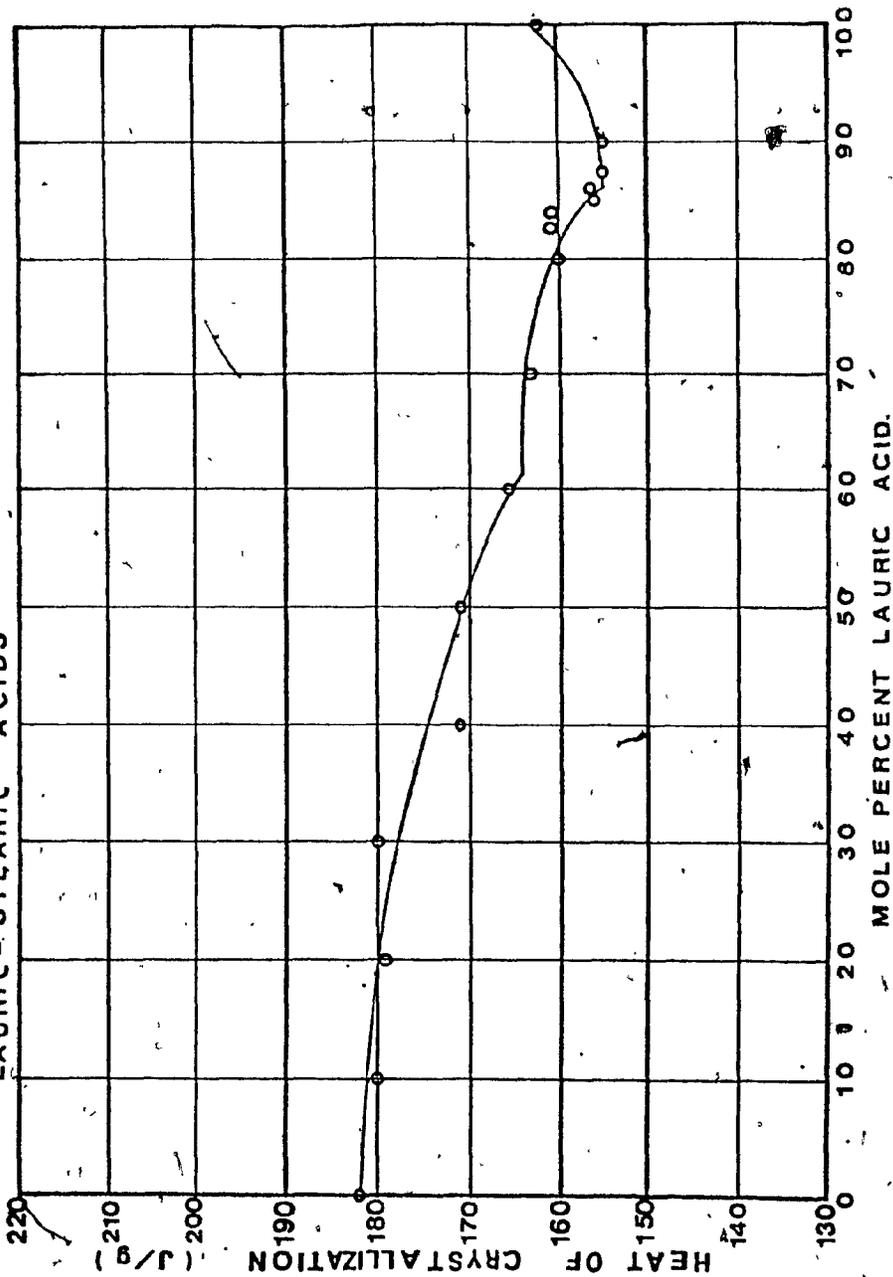
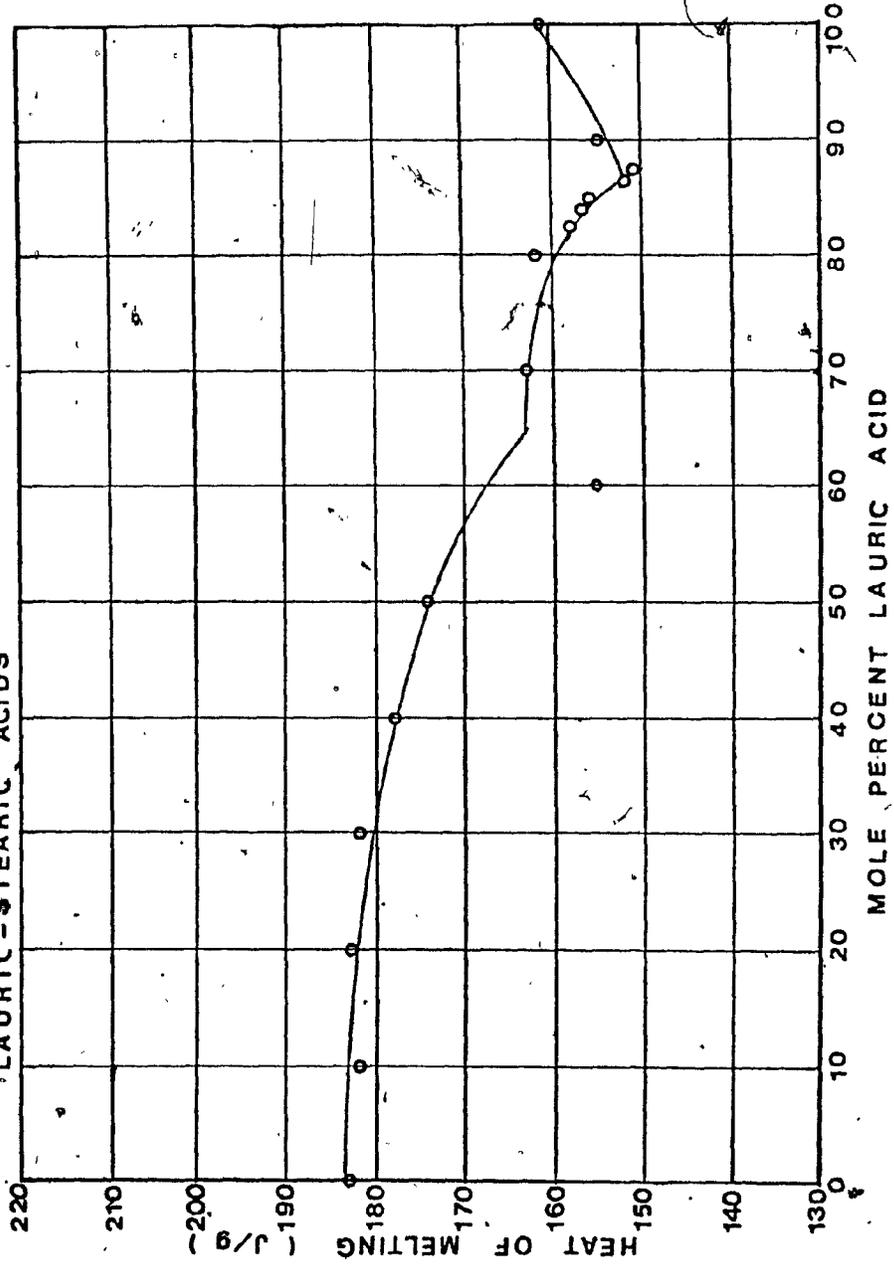


FIG.27 HEAT OF MELTING OF BINARY SYSTEM  
LAURIC-STEARIC ACIDS



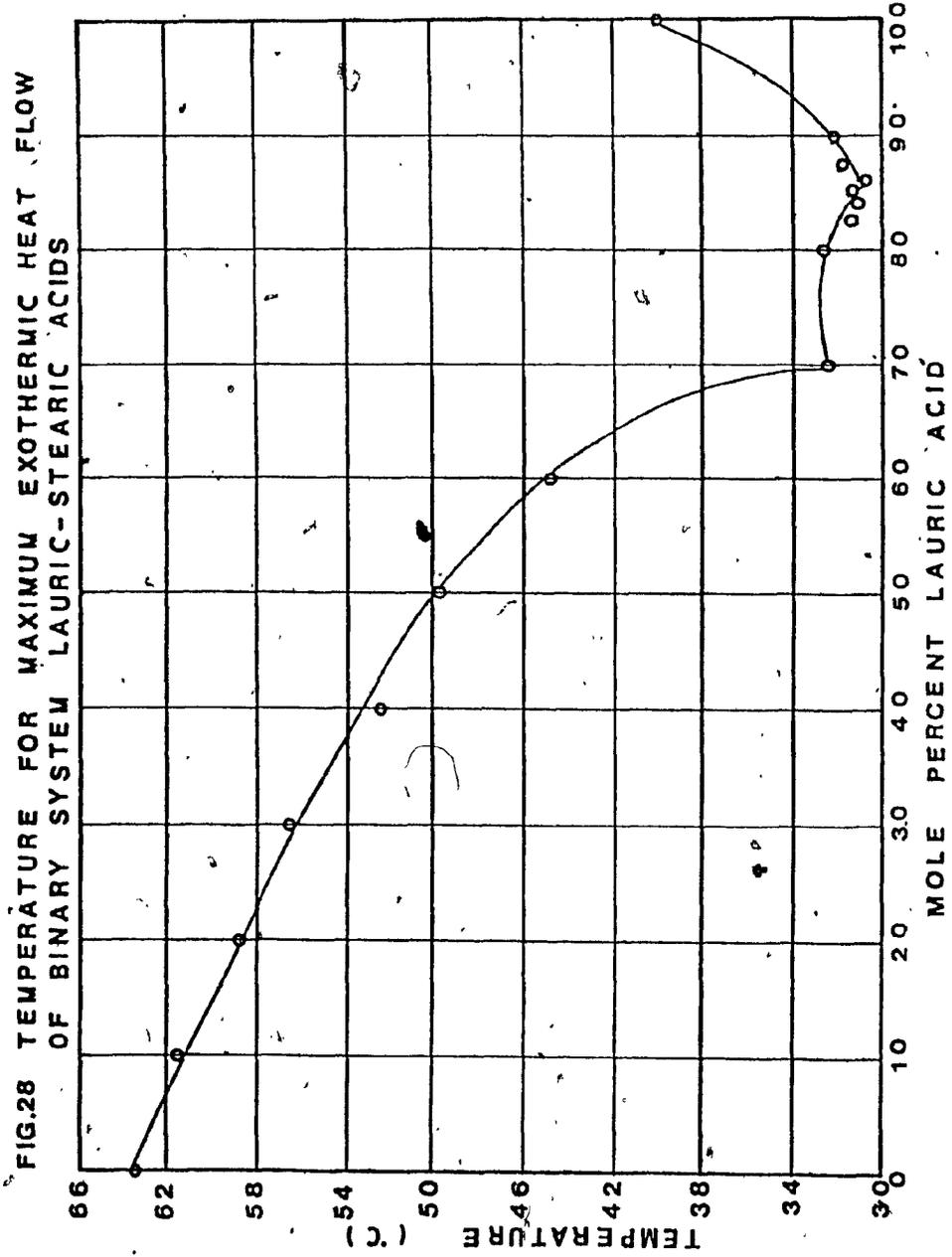
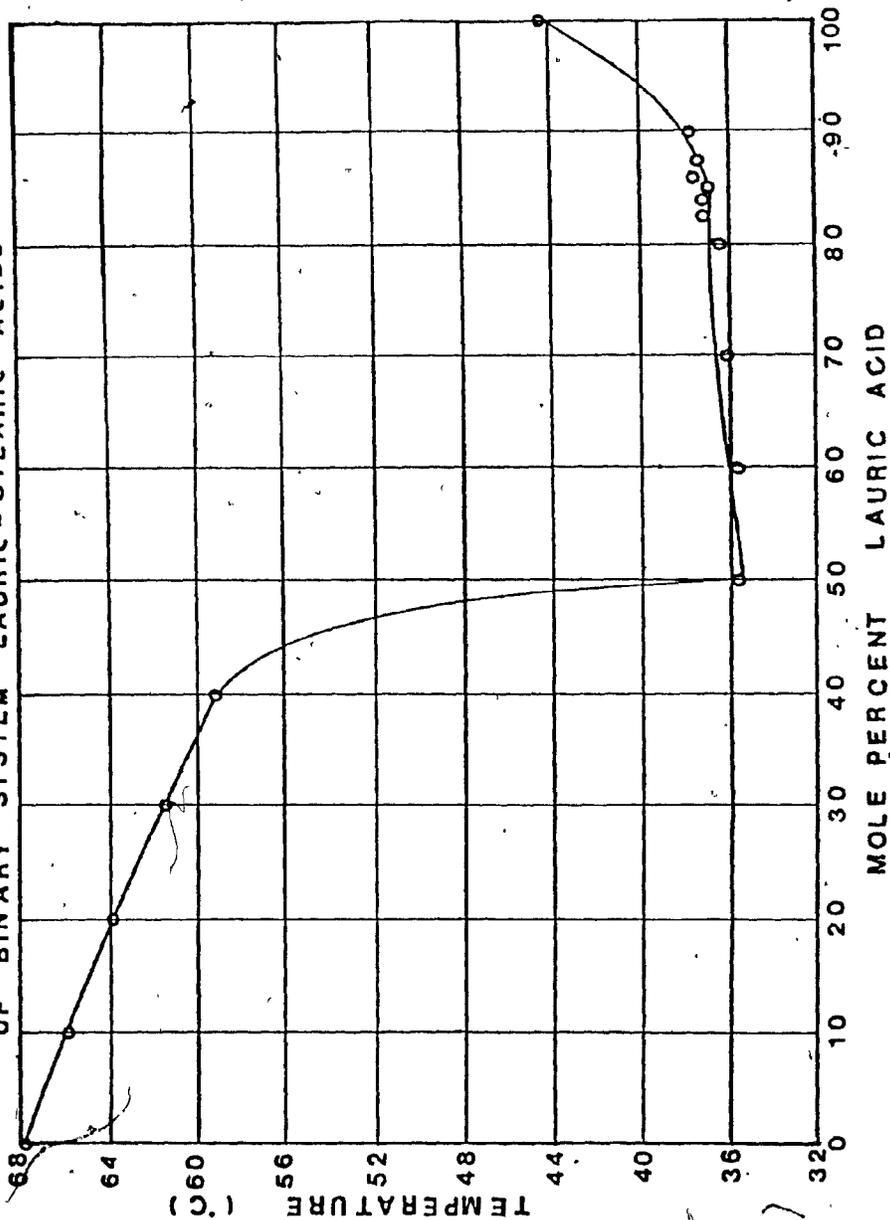


FIG.29 TEMPERATURE FOR MAXIMUM ENDOTHERMIC HEAT FLOW OF BINARY SYSTEM LAURIC - STEARIC ACIDS



## VII. THERMAL PROPERTIES OF FATTY ACIDS

### Melting Points

The melting points of capric, lauric, palmitic and stearic acids were observed to be  $30.0 \pm 0.4^\circ\text{C}$ ,  $42.2 \pm 0.1^\circ\text{C}$ ,  $59.0 \pm 0.6^\circ\text{C}$  and  $65.6 \pm 0.1^\circ\text{C}$  respectively. The observed means and standard deviations of the melting points and other parameters are listed in Table VIII. The melting points of capric and lauric acids are about  $1^\circ\text{C}$  lower than the published values (Ref. 13) and those of palmitic and stearic acids are about  $4^\circ\text{C}$  lower. The generally accepted melting range of these acids is shown in Table VI. However, the observed melting points were close to the points found by Berchiesi (Ref. 14) which were  $30.5^\circ\text{C}$ ,  $42.8^\circ\text{C}$ ,  $61.4^\circ\text{C}$  and  $66.2^\circ\text{C}$  for capric, lauric, palmitic and stearic acids respectively. The discrepancy in the results appears to be due to the degree of impurities found in the reagent grade substances. Also, in the DSC calculations the melting point is taken at the inflection point of the curve which has the greatest rate of change of heat flow. The tangent drawn at the inflection point and projected to the base line can produce a value of the melting point which may differ from the value determined by other methods. In addition, in the DSC method the heating rate and mass of sample affect the melting point. The relationship between the melting point and the chain length (number of carbon atoms in the acid) is shown in Fig. 30. From the figure, it can be seen that the melting point increases directly with the chain length.

TABLE VIII. Observed Thermal Properties of Fatty Acids\*

COMMON NAME	MELTING POINT (°C)	TEMPERATURE FOR MAX. ENDOTHERMIC HEAT FLOW (°C)	FREEZING POINT(°C)	TEMPERATURE FOR MAX. EXOTHERMIC HEAT FLOW (°C)	HEAT OF MELTING		HEAT OF CRYSTALLIZATION		ENTROPY OF MELTING J/g - °K
					J/g	Kcal/Mole	J/g	Kcal/Mole	
Capric	30.0 ± 0.4	31.8 ± 0.8	28.7 ± 0.3	28.6 ± 0.1	157 ± 6	6.5	153 ± 7	6.3	0.518
Lauric	42.2 ± 0.1	44.2 ± 0.4	40.7 ± 0.1	39.8 ± 0.1	164 ± 4	7.8	162 ± 7	7.8	0.520
Palmitic	59.0 ± 0.6	61.5 ± 0.1	58.1 ± 0.3	57.6 ± 0.7	175 ± 1	10.7	173 ± 3	10.6	0.527
Stearic	64.6 ± 0.1	67.9 ± 0.1	64.5 ± 0.2	63.4 ± 0.1	183 ± 3	12.4	182 ± 2	12.4	0.542

\*. The results were obtained from 4 different runs on palmitic acid, 2 runs on capric acid, 2 runs on lauric acid and 2 runs on stearic acid. All determinations were done at a data scanning rate of 0.4 seconds per point and are listed in table A-1 of the appendix.

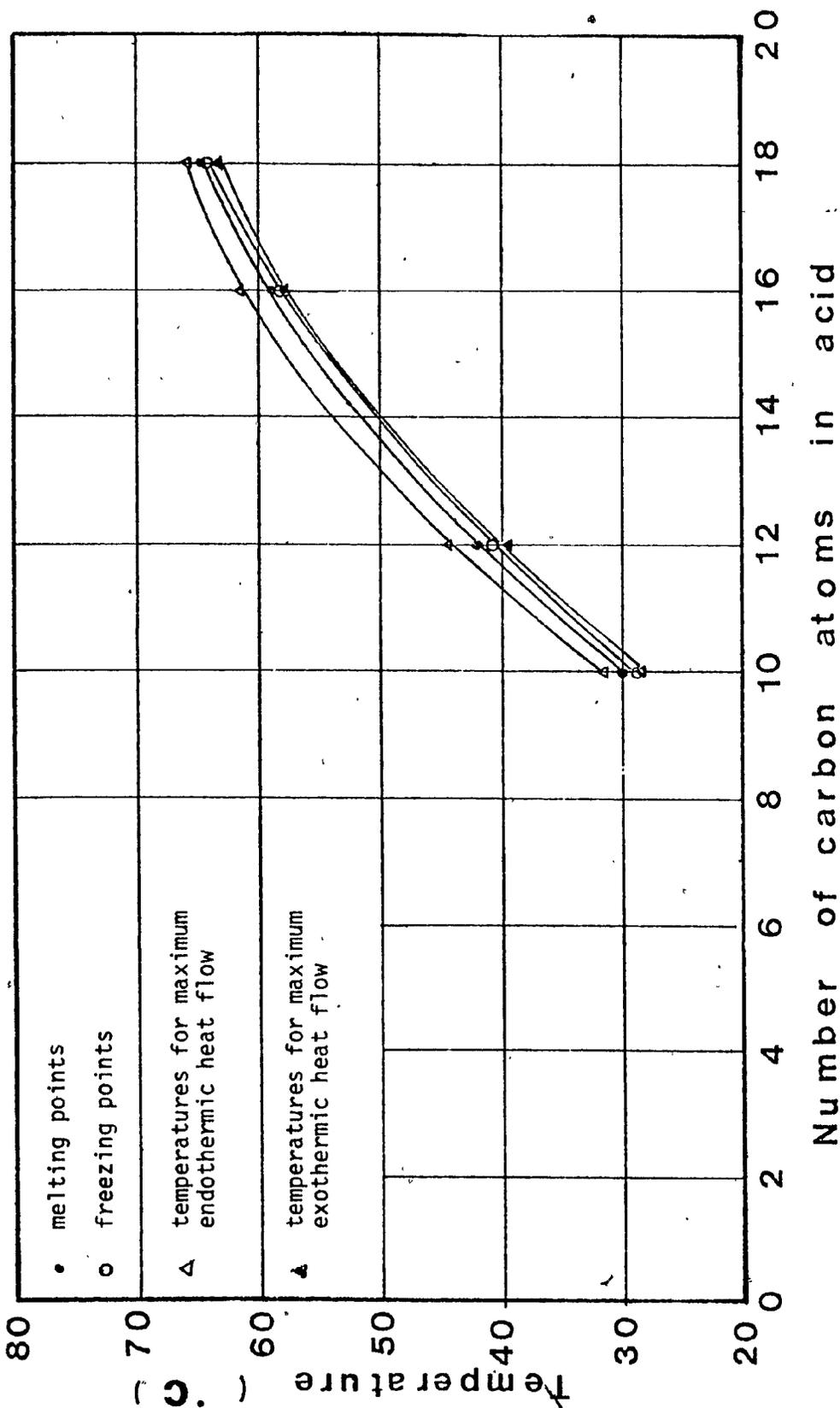


Fig. 30. Melting and freezing points and temperatures for maximum endothermic and exothermic heat flows of fatty acids.

### Freezing Points

The freezing points of capric, lauric, palmitic and stearic acids were observed to be  $28.7 \pm 0.3^\circ\text{C}$ ,  $40.7 \pm 0.1^\circ\text{C}$ ,  $58.1 \pm 0.3^\circ\text{C}$  and  $64.5 \pm 0.2^\circ\text{C}$  respectively. The freezing points are slightly lower than the melting points. In contrast to the melting point, the freezing point is not an equilibrium measure and the supercooling effect is an important factor. Also, the cooling rate and mass of sample can affect the freezing point. Kulka et al (Ref. 10) found the freezing point of pure capric acid to be  $31.2^\circ\text{C}$ . Efremov et al (Ref. 13) observed freezing points of lauric, palmitic and stearic acids at  $42.7^\circ\text{C}$ ,  $59.2^\circ\text{C}$  and  $67.7^\circ\text{C}$  respectively.

The discrepancy between the literature and experimental values is due to the presence of impurities in the reagents. Also, the literature values are based on the capillary tube method which has a lower error (max.  $0.05^\circ\text{C}$ ) in reproduction of the freezing point (Ref. 10 and 12). In addition, the capillary tube method requires a larger sample weight than the DSC method and, therefore, the supercooling effect is smaller in the capillary tube method.

The freezing point dependence on the carbon chain length in the acid is illustrated in Fig. 30. The profile of the freezing curve is similar to the melting curve.

### Temperatures for Maximum Endothermic and Exothermic Heat Flows

The temperatures for maximum endothermic and exothermic heat flows when plotted against the number of carbon atoms in the acid show a close resemblance to the melting and freezing curves.

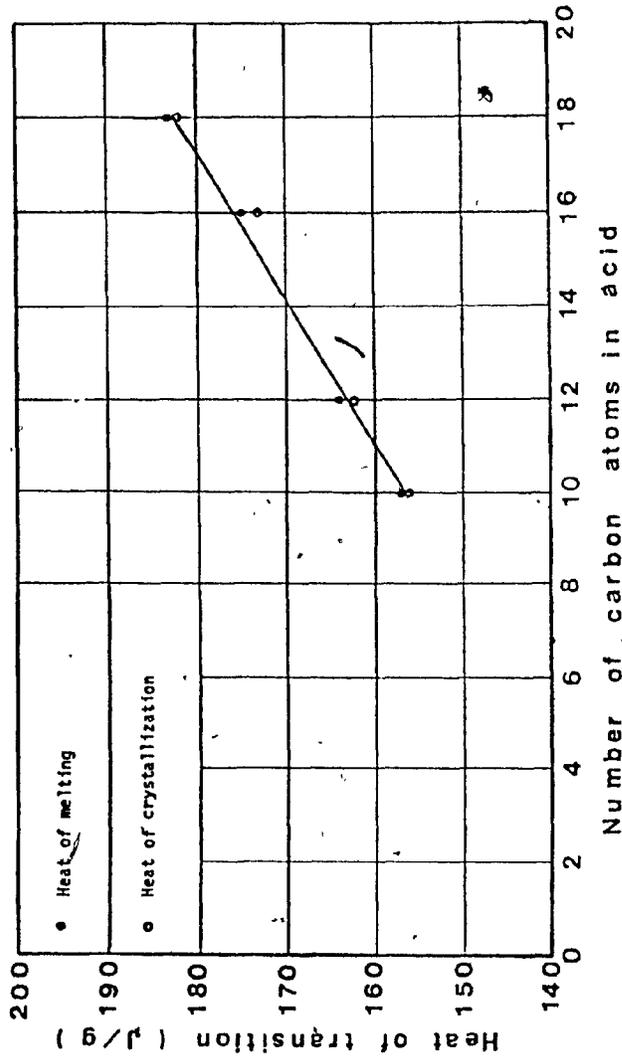


Fig. 31. Heats of melting and crystallization of fatty acids.

This is illustrated in Fig. 30. These points are highly dependent on the heating and cooling rates as shown in Fig. T-1 to T-5 of the appendix.

#### Heats of Melting and Crystallization

The observed heats of melting for capric, lauric, palmitic and stearic acids are  $157 \pm 6$  J/g,  $164 \pm 4$  J/g,  $175 \pm 1$  J/g and  $183 \pm 3$  J/g respectively. The corresponding heats of crystalli-

zation.  $153 \pm 7$  J/g,  $162 \pm 7$  J/g,  $173.3 \pm 3$  J/g and  $182 \pm 2$  J/g, respectively. The heats of melting and crystallization appear to be essentially the same. These transition heats have been plotted versus the number of carbon atoms in the acid chain in Fig. 31. The resulting curve shows that the heats of melting and crystallization increase with an ascending number of carbon atoms.

The observed heats of melting of the pure acids are slightly lower than the published data (Ref. 13 and 15). However, Berchiesi et al (Ref. 14) found the heat of melting to be 8.2, 12.3, and 13.8 Kcal/mole for lauric, palmitic and stearic acids respectively. This compares to 7.8, 10.7 and 12.4 Kcal/mole for lauric, palmitic and stearic acids obtained in the DSC results after applying a conversion factor. The discrepancy in the observed values appears to be due to the degree of impurities found in the reagent substances and different measurement techniques.

#### Entropy of Melting

The entropy of melting of the pure fatty acids is shown in Table VIII. The relationship of the entropy with the number of carbon atoms in the acid chain is illustrated graphically in Fig. 32. The entropy of melting of the fatty acids has the same order of magnitude for this particular homologous series but tends to increase with an ascending number of carbon atoms in the acid chain.

The entropies of melting were calculated with the use of the following equation:

$$\Delta S = \frac{\Delta H_m}{T_{m.p.}}$$

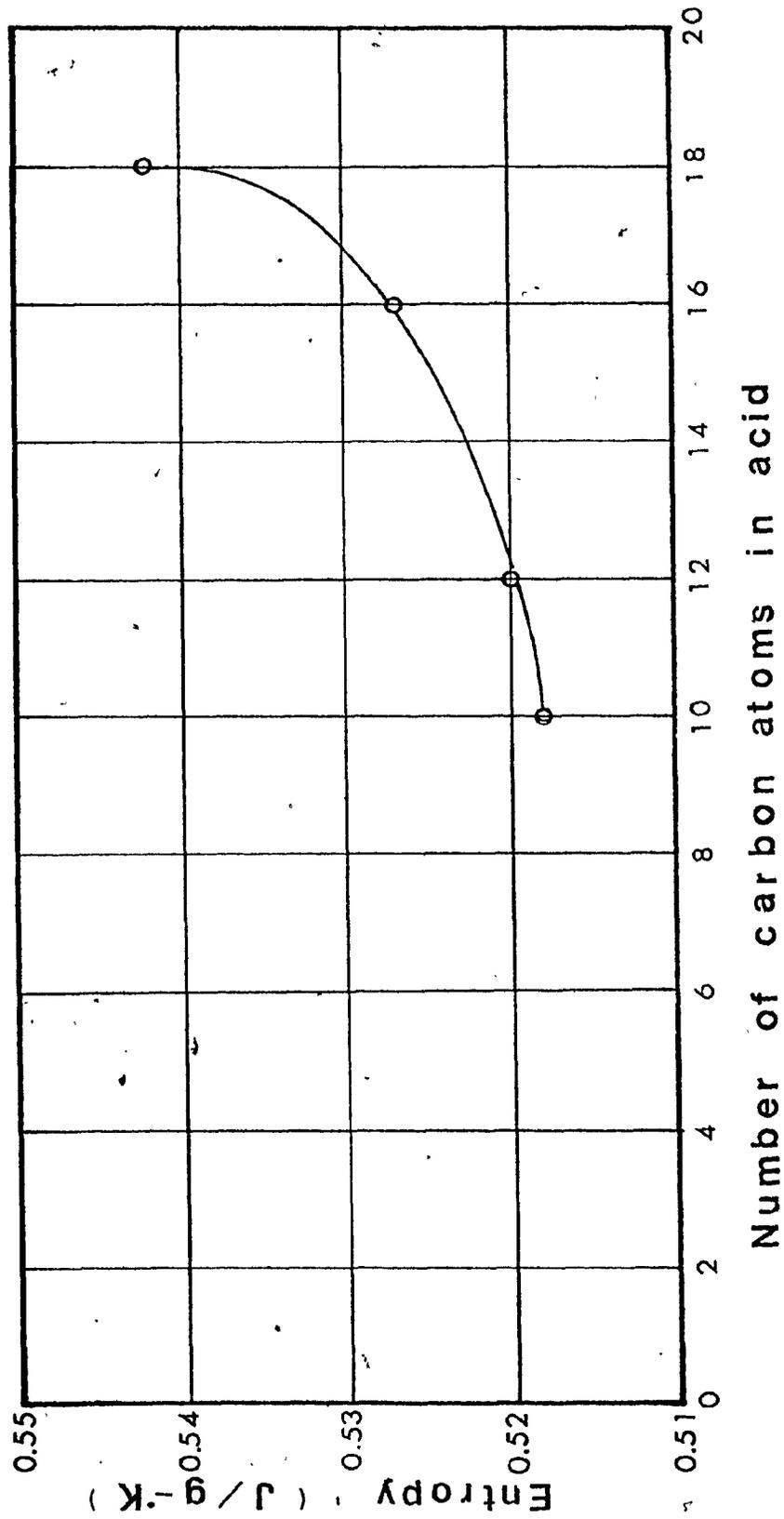


Fig. 32. Entropy of melting of fatty acids.

where:  $\Delta S$  = entropy of melting in J/g - °K

$\Delta H_m$  = heat of melting in J/g

Tm.p. = melting point temperature in °K (°C + 273.15)

Internal Energy Change of Melting

The change in internal energy during melting can be calculated using the enthalpy function at constant pressure.

$$\Delta H = \Delta E + P\Delta V$$

where:  $\Delta H$  = heat absorbed by the system (heat of melting)  
in J/g

$\Delta E$  = change in internal energy of the system in J/g

P = pressure at which melting occurs in N/M<sup>2</sup>

$\Delta V$  = molar volume change between the liquid and solid  
in cm<sup>3</sup>/g-mole

Rearranging the above equation and introducing the terms for the molar volume of liquid and solid, the equation becomes

$$\Delta E = \Delta H - P (V_L - V_s)$$

where:  $V_L$  = molar volume of liquid in cm<sup>3</sup>/g-mole

$V_s$  = molar volume of solid in cm<sup>3</sup>/g-mole

Using capric acid as an example with the following data:

$V_L$  = 192.4 cm<sup>3</sup>/g-mole (Ref. 16)

$V_s$  = 170.2 cm<sup>3</sup>/g-mole (Ref. 16)

P = 1 atmosphere

$\Delta H$  = 157 J/g

$$\begin{aligned} \Delta E &= 157 \text{ J/g} - [1 \text{ ATM}] \left[ \frac{1.0133 \times 10^5 \text{ N/m}^2}{1 \text{ ATM}} \right] \left[ \frac{22.2 \text{ cm}^3}{\text{g-mole}} \right] \left[ \frac{1 \text{ g-mole}}{172.27 \text{ g}} \right] \left[ \frac{1 \text{ m}^3}{10^6 \text{ cm}^3} \right] \left[ \frac{1 \text{ J}}{\text{N-m}} \right] \\ &= 157 - 0.013 \text{ J/g} \qquad \qquad \qquad = 156.987 \text{ J/g} \end{aligned}$$

The above calculation illustrates that very little work is expended when the fatty acid undergoes melting and the internal energy change is essentially the same as the heat of melting.

### XIII. CRYSTAL STRUCTURE OF FATTY ACIDS AND THEIR BINARY SYSTEMS

Knowledge of the crystal structure of fatty acids and their solid solutions is essential in the understanding of the phase behaviour and polymorphism.

Extensive work has been done to determine the crystallographic structure of pure fatty acids and their binary systems using the X-ray powder and single crystal techniques (Ref. 17, 18, 19 and 20). As a result, a considerable amount of data has been obtained about the packing of the hydrocarbon chains, orientation of the carboxyl group and polymorphism.

The X-ray powder patterns of the compounds reveal the existence of two group spacings. These group spacings are illustrated in Fig. 33. The first group is referred to as a long spacing and the second group consists of two short spacings. The long spacing increases proportionally with the number of  $\text{CH}_2$  groups in a given homologous series. The two short spacings have approximately the same value for a particular homologous series and are associated with the manner of packing of the hydrocarbon chains.

X-ray diffractometric results have shown the existence of three polymorphic forms for the even straight chain fatty acids (Ref. 35). These forms are known as A, B and C modifications with the long spacing diminishing in that order. The C form is the most stable modification (Ref. 35). When the A and B modifications are heated to 10-15°C below their melting point, the two modifications transform irreversibly to the C modification with little heat of transition (Ref. 22). Consequently, the C polymorph of the

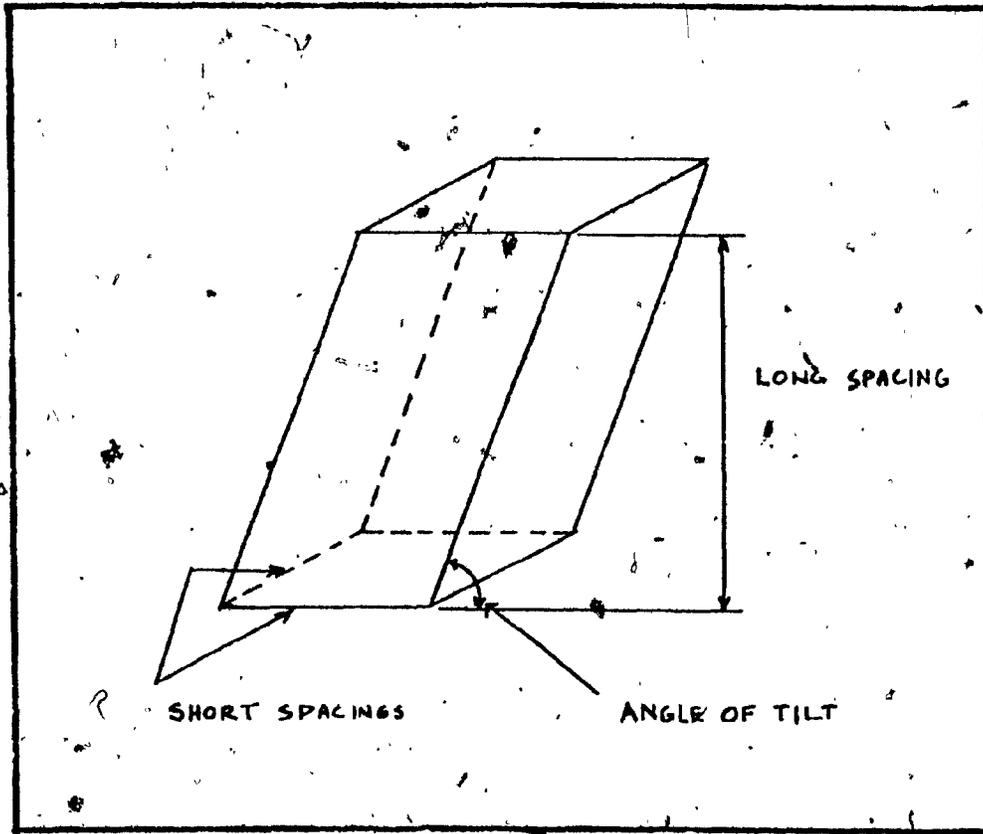


Fig. 33. Group spacings of unit crystal cell of fatty acids (Ref. 21).

acid is always obtained from a melt and the acids exhibit the property of monotropy.

The C<sub>2</sub> form has a monoclinic prismatic structure with four molecules to the unit cell. In this arrangement, the ends of the molecules are found at the corners and in the centre of the parallelepiped. Shearer (Ref. 23) found that the fatty acids crystallize with two molecules between successive layers. The active groups COOH of the molecules are located in the centre and the inactive groups CH<sub>3</sub> at the corners of the prism.

Müller (Ref. 29) showed that the carbon atoms in the chain are arranged in a zig-zag pattern and are joined to each other at a tetrahedral angle. Subsequent experimental work by Piper (Ref. 25) and other workers have found the angle to be 116°. The distance between the adjacent carbon atoms was found to be 2.6Å (Ref. 25).

The lattice dimensions of the C-form of the fatty acids are given in Table IX.

Name	Short Spacing a (Å)	Short Spacing b (Å)	Long Spacing c (Å)	Angle of Tilt β	Ref.
Capric			29.7		26
Lauric	9.524	4.965	35.39	129°13'	27
Palmitic	9.41	5.0	45.9	129°10'	27
Stearic	9.357	4.956	50.76	128°14'	27

TABLE IX. Crystal lattice dimensions of fatty acids

The unit cell dimensions vary slightly with chain length and, therefore, the series is not strictly homologous. The cross section of the unit cell decreases with a longer chain indicating that the molecules are packed closer together in the longer chain acid.

Fig 34 shows a diagrammatic representation of the unit cell of stearic acid.

The crystal structures of binary mixtures of fatty acids are well documented in the work of Frede et al (Ref. 28) and Baun (Ref. 26).

Frede et al examined different mixtures of the palmitic-stearic system by means of X-ray and electron diffraction and found that the crystal form occurring in all concentrations conforms to the C-form of the pure compounds. In the concentration range of 95-60 mole % of stearic acid, the shorter palmitic acid molecules are incorporated into the crystal lattice of stearic acid without affecting the long spacing,  $c$ , of the unit cell. In the concentration range of 50 to 20 mole % of stearic acid, the crystal structure is characterized by constancy of nearly all lattice parameters. The long spacing is approximately the arithmetic mean of the spacings of the pure components. Frede et al depict the mixed crystal form in terms of double molecules, which can be formed from two stearic acid molecules, two palmitic acid molecules and one stearic and one palmitic acid molecule. Within the concentration range of 0-10 mole % stearic acid, the longer stearic acid molecules can be incorporated into the crystal lattice of the palmitic acid. The unit cell of the palmitic acid is slightly affected by a decrease in the angle of tilt  $\beta$ , by 1-2° and an enlargement of the distance between the neighbouring atoms.

Saguchi and Asada (Ref. 18) determined unit crystal cell dimensions of binary systems of  $C_{12}-C_{16}$  and  $C_{16}-C_{18}$  by means of X-ray diffraction. The systems were classified into three groups:

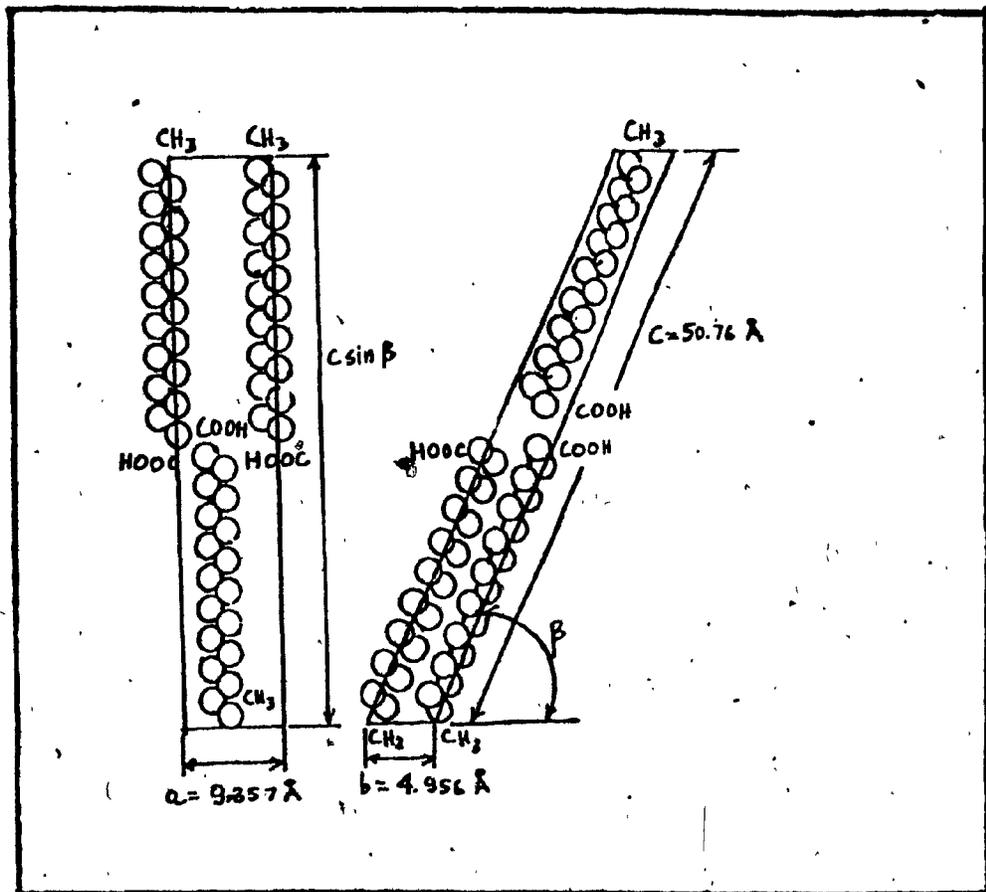


Fig. 34. Diagrammatic representation of the unit cell of a stearic acid crystal (Ref. 37).

1. A solid solution in which the lower-chain acid is dissolved by the higher-chain acid.
2. An equimolar compound composed from the two components containing one of the components in small excess.
3. A solid solution in which the higher-chain acid is dissolved by the lower-chain acid.

The stability of the crystal lattice is affected by the difference between the number of carbon atoms in the fatty acid molecules. As the difference rises, the stability of the lower acid in the higher acid increases, whereas the stability of the higher acid in the lower acid decreases (Ref. 18).

Baun (Ref. 26) has examined the binary system of  $C_{10}-C_{12}$  by X-ray diffraction. His results of mixed crystal formation are in general agreement with the other workers.

IX. ANALYSIS OF FATTY ACIDS AND THEIR EUTECTICS  
WITH AN X-RAY DIFFRACTOMETER

An X-ray diffractometer (manufactured by Picker) equipped with a Chromium tube (manufactured by Macklett) was used to obtain the X-ray patterns. Measurements were made of pure fatty acids and their eutectics (except for capric-lauric eutectic, which is liquid at ambient temperature). The samples were melted in an oven and later crystallized before being used on the instrument. Subsequently, the samples were ground on a watchglass with a pestle to obtain a fine homogeneous powder. The material was then pressed in a thin layer on a strip of glass to obtain sufficient adhesion. Due to the waxy nature of the compound, the samples easily adhered to the glass strip.

The X-ray patterns were recorded in an angular range of 6 to 50° at an angular velocity of 1°/min. A strip chart recorder was used to record the diffraction pattern.

All pure acids and their mixtures were found to be present in the C-form having a monoclinic crystal lattice. This form is characterized by short spacings at  $2\theta=19.4^\circ$ ,  $2\theta=21.5^\circ$  and  $2\theta=24^\circ$ . This is in agreement with the results found by Frede et al (Ref. 28) except for the angle of  $2\theta=19.4^\circ$ , which Frede et al found to be  $20.2^\circ$ .

The long spacings were found to be located at  $2\theta=11.5^\circ$  for capric acid,  $2\theta=9.7^\circ$  for lauric acid,  $2\theta=7.4^\circ$  for palmitic acid and  $2\theta=6.6^\circ$  for stearic acid. The values for palmitic and stearic acids are in agreement with the results of Frede et al.

The intensity of the long spacings essentially disappear in the binary mixtures of acids. The X-ray patterns conform to the results obtained by Frede et al, Van (Ref. 29) and Malta et al (Ref. 30).

The diffractometric results indicate that during crystallization of a binary mixture of fatty acids, the mixture does not decompose into the pure acid components (Ref. 28).

The scattering of X-rays depends on the concentration of electrons in a given area (Ref. 41). The terminal methyl groups in the fatty acids contain a low electron density due to the high concentration of hydrogen and, therefore, the methyl plane has a lower scattering power than the main body of the crystal. In contrast, the presence of a carboxyl group gives rise to planes of higher scattering power than the main body of the crystal. With double molecules, the strongly diffracting and the weakly diffracting planes are separated by the same distance. This is illustrated in Fig. 35.

The results of the X-ray spectra are shown in Fig. S-1 to Fig. S-9 in the appendix. The vertical lines which appear on the spectra denote the location of the short and long spacings. These lines were drawn skewed rather than vertical as a result of a misaligned strip chart paper caused by a fault on the X-ray diffractometer recorder. The actual peak readings on the spectra were taken directly from the angle indicator on the diffractometer.

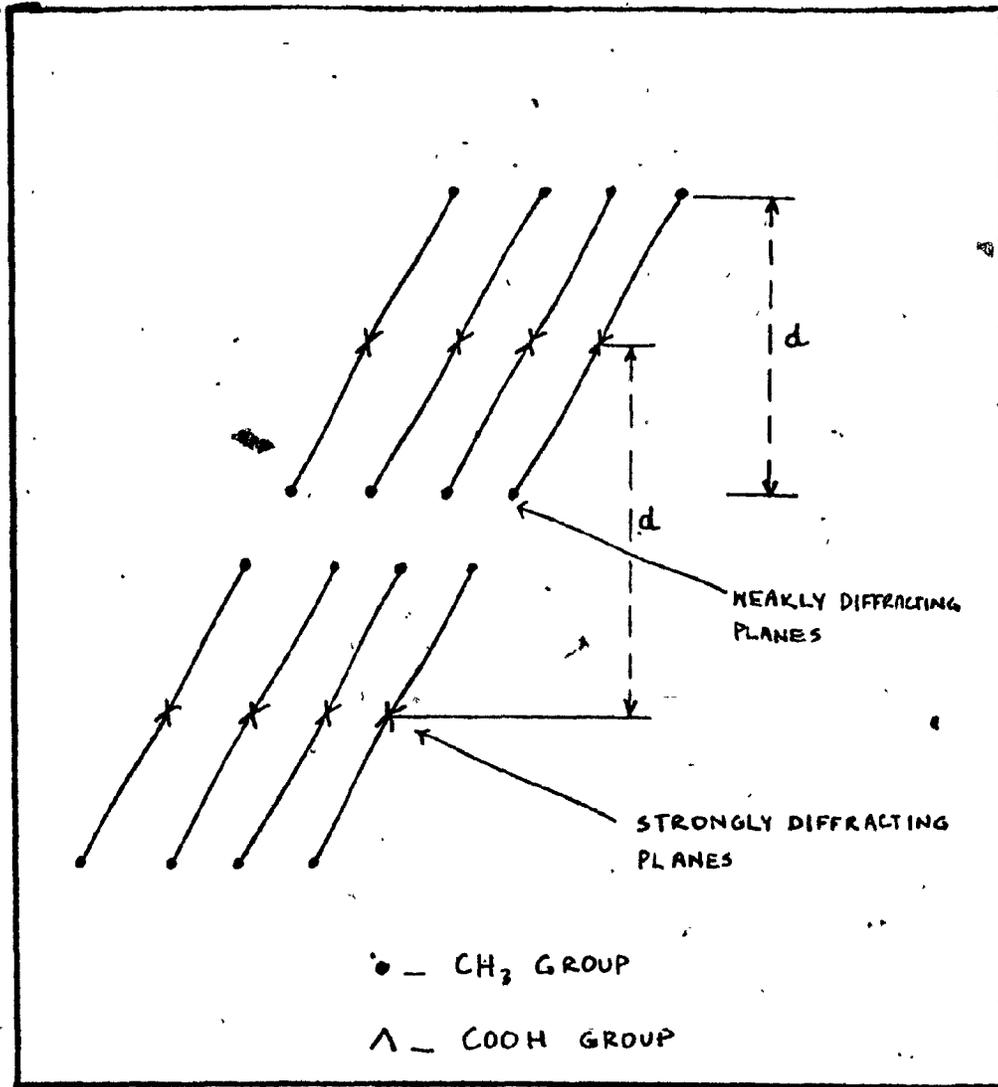


Fig. 35. X-ray diffracting planes of fatty acids (Ref. 41).

X. INFRARED SPECTRA OF FATTY ACIDS AND THEIR EUTECTICS

The fatty acids used for the experiments were obtained from commercial sources as given in Table VI. The samples were prepared by mixing about 10 mg of the compounds with about 1000 mg of potassium bromide in a mortar dish. Subsequently, the mixtures were ground with a pestle to a fine homogeneous powder. The powder was then placed in a die which was subjected to a force of 10 metric tons to produce a potassium bromide disk. The only sample which remained in a liquid form consisted of 74 mole % capric: 26 mole % lauric. Consequently, the IR analysis on this sample was performed by placing a capillary film of the sample between two potassium bromide disks. A Beckman spectrophotometer, Model IR 4240, was used to measure the IR spectra over a range of 4000 to 400  $\text{cm}^{-1}$ .

The IR spectra of the pure fatty acids and their binary mixtures are shown in Fig. I-1 and I-2 in the appendix. Comparison of the spectra of the pure acids with standard published spectra (Ref. 31) show that the acids are present in the C-polymorphic form. This is particularly apparent at a wave number of about 930  $\text{cm}^{-1}$  where the O-H out-of-plane deformation occurs and no two crystal forms have the same absorption. This is consistent with the X-ray diffraction results.

The IR spectra of the even straight-chain fatty acids in crystalline form show a series of bands of uniform spacing in the region of 1300 to 1180  $\text{cm}^{-1}$ . This is due to interaction of rocking and/or twisting vibrations of the  $\text{CH}_2$  groups. The number of bands increases with the chain length of the molecule and the number of carbon

atoms can be calculated by multiplying the number of bands for a particular compound by a factor of 2. The spectra of the fatty acids in the 1300 to 1180  $\text{cm}^{-1}$  region show 5 bands for capric acid, 6 bands for lauric acid, 8 bands for palmitic acid and 9 bands for stearic acid.

Table X shows the band positions for fatty acids and their binary mixtures. The bands were designated in a numerical order with decreasing wavelengths. The locations of the functional groups are in agreement with published data.

It is noteworthy that for the binary mixtures the region of 1300 to 1180  $\text{cm}^{-1}$  is significantly altered from the one for pure fatty acids. The vibrations of the  $\text{CH}_2$  groups are markedly different indicating a different order of molecular packing in the crystal lattice.

TABLE X. Infrared Spectra Band Positions for Fatty Acids and Their Binary Mixtures.

FUNCTIONAL GROUP	CAPRIC ACID		LAURIC ACID		PALMITIC ACID		STEARIC ACID		74% CAPRIC 26% LAURIC		85% LAURIC 15% STEARIC		72.5% PALMITIC 27.5% STEARIC		80% CAPRIC 20% PALMITIC	
	GRAPH ID	BAND	GRAPH ID	BAND	GRAPH ID	BAND	GRAPH ID	BAND	GRAPH ID	BAND	GRAPH ID	BAND	GRAPH ID	BAND	GRAPH ID	BAND
C-H <sub>3</sub> Assym. Stretching	1C	2955	1L	2940	1P	2940	1S	2955	1CL	2944	1LS	2940	1PS	2938	1LP	2940
		2920		2910		2900		2920		2910		2905		2900		2910
C-H <sub>2</sub> Assym. Stretching	2C	2870	2L	2860	2P	2855	2S	2870	2CL	2860	2LS	2860	2PS	2840	2LP	2860
		2850		2840		2835		2845		2845		2840		2830		2840
C-H <sub>3</sub> Sym. Stretching	3C	1708	3L	1695	3P	1690	3S	1700	3CL	1698	3LS	1695	3PS	1690	3LP	1700
		1410		1404		1400		1410		1405		1402		1400		1402
C-H Bending	4C	1435	4L	1425	4P	1420	4S	1430	4CL	1430	4LS	1425	4PS	1420	4LP	1425
		1462		1460		1455		1465		1460		1458		1455		1455
C=O Stretching	5C	1235	5L	1235	5P	1235	5S	1293	5CL	1295	5LS	1290	5PS	1285	5LP	1280
		1190		1185		1180		1182		1180		1175		1175		1175
C-H Wagging and/or Bending	6C	935	6L	925	6P	925	6S	930	6CL	925	6LS	925	6PS	925	6LP	925
		718		712		712		718		712		712		712		712
O-H Vibration	7C	718	7L	712	7P	718	7S	725	7CL	712	7LS	715	7PS	705	7LP	708
		718		712		712		718		712		712		712		712
C-H <sub>2</sub> Rocking	8C	718	8L	712	8P	718	8S	725	8CL	712	8LS	715	8PS	705	8LP	708
		718		712		712		718		712		712		712		712
C-H <sub>2</sub> Rocking	9C	718	9L	712	9P	718	9S	725	9CL	712	9LS	715	9PS	705	9LP	708
		718		712		712		718		712		712		712		712

THEORY OF MELTING AND CRYSTALLIZATION OF FATTY ACIDS

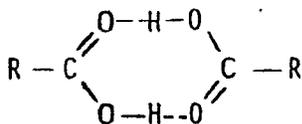
The melting process is a function of the structure of the crystalline state and can be visualized as the structural breakdown of the crystal lattices. In the crystalline solid the molecules are arranged in a regular and symmetrical way. The process of melting causes the highly ordered arrangement to break down to a more random arrangement which normally characterizes a liquid.

The measure of the randomness of the molecular disorder of the system is expressed by a change of entropy which accompanies a phase change in a material. The change of entropy of fatty acids increases during melting as illustrated in Fig. 32.

The melting proceeds at a temperature level at which the thermal energy of the molecules is sufficiently high to overcome the crystalline forces holding the molecules together in the crystal lattice. First, the surface molecules acquire sufficient vibration energy to escape the lattice. Also, on a smaller scale, interior molecules can rupture the crystal lattice when sufficient vibrational energy is acquired. When the fatty acids undergo melting, the heat of melting is essentially converted to the internal energy of the molecules with little work done by the system during the phase-change process. This is illustrated by the calculation of internal energy change shown on pg. 61. The heat of melting depends on the molecular weight and tends to increase with an increase in the carbon chain length of the acid as shown in Fig. 31.

The intermolecular forces holding the molecules together in the crystal lattice can be viewed as composed of two types of forces; namely, the dipole-dipole interaction due to the presence of the polar OH group and the Van der Waals forces (Ref. 16). The latter forces are small, momentary polarizations of part of one molecule by part of another near-by molecule.

The monocarboxylic acid molecules are polar and are normally called associated liquids due to the presence of the hydrogen bond. These molecules are dimerized in a liquid phase and held together by two hydrogen bonds as shown below.



The hydrogen bond has a strength of about 5Kcal/mole (Ref. 40) and is the main factor for the higher than expected melting and boiling points of the fatty acids. The melting points of the acids depend on polarity and molecular weight. For a particular homologous series, the melting points increase with an increase in the molecular weight as illustrated in Fig. 30. The polarity of the acid molecules imposes on them an arrangement which gives the greatest attractive forces between molecules. This is counter-balanced by the kinetic energy of the molecules.

Crystallization is the reverse of melting and occurs when a number of molecules at a low energy level are grouped together to form a nucleus. Supercooling, a metastable state, can proceed in the absence of a nucleus on which the crystal growth is dependent. The acids are susceptible to the supercooling phenomenon as shown in Fig. T-6 and T-8 in the appendix.

## XII. DISCUSSION OF THERMOGRAMS OF BINARY MIXTURES OF FATTY ACIDS

The analysis of multiple peaks on thermograms of binary mixtures of fatty acids can provide useful information about phase transitions and the possible structure of molecular crystals.

The nature of the peaks can be examined in terms of the solid solutions that can be formed by the two components. Analysis of the melting and freezing curves shows that an additional compound is formed at the hump in the curve. Therefore, the binary mixture of fatty acids can be considered to contain three types of solid solutions. These solutions have been reported by Saguchi and Asada (Ref. 18) and can be described as (1) solid solution of the shorter-chain fatty acid in the longer-chain fatty acid (2) equimolar compound of the two components containing one of them in small excess (3) solid solution with small amount of the longer-chain acid dissolved in the shorter-chain acid.

Binary mixtures of even straight-chain acids contain up to four discernible peaks on the thermograms depending on the concentration of the two components. The pure even straight-chain acids and their eutectic compounds are characterized by a single, sharp and well-defined peak. The number of peaks appearing on the thermograms for the binary systems of capric-lauric, lauric-palmitic, lauric-stearic and palmitic-stearic acids have been plotted in Fig. 36.

Fig. 36 shows that the region containing up to four peaks is rich in the high-molecular weight component. The X-ray diffractometric results clearly show that the crystal lattice in this concen-

- Binary mixtures of palmitic-stearic acids
- Binary mixtures of lauric-stearic acids
- × Binary mixtures of capric-lauric acids
- ◇ Binary mixtures of lauric-palmitic acids

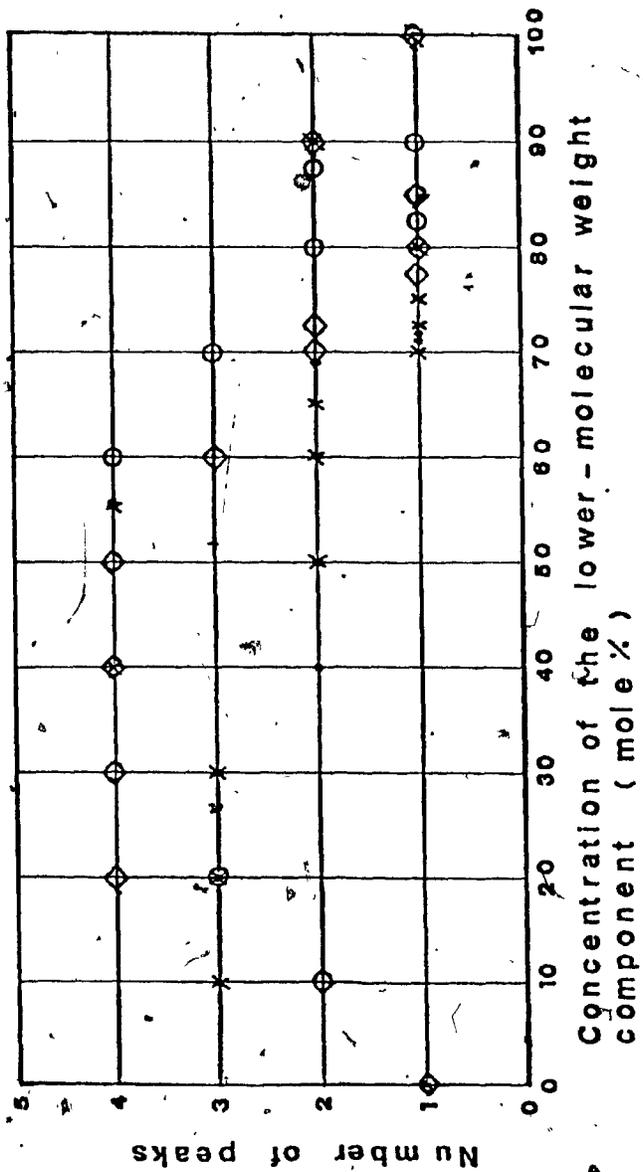


Fig. 36. Plot of number of peaks versus concentration for thermograms of binary mixtures of fatty acids.

tration region has the cell dimensions of the high-molecular weight component. Taking the example of the palmitic-stearic system for the purpose of discussion, it can be seen that in the region which is rich in the high-molecular weight component, stearic acid in this instance, the crystal lattice must conform to the stearic acid cell dimensions (Ref. 28).

The possible crystal lattice arrangements that can conform to the stearic acid cell dimensions are shown in Fig. 37 and consist of lattice structures Type 1, Type 2, Type 3 and Type 4. Fig. 37 shows that the acids crystallize in double-chain length molecules. The crystal lattices possess different stabilities (Ref. 18 and 28) which require different energy formation levels. When the crystal lattice absorbs sufficient thermal energy to disorder the molecular arrangement in the lattice, the crystal lattice is expected to rupture. This type of breakdown can be associated with the appearance of the peak on the thermogram. Therefore, the four peaks on the thermogram can be explained in terms of rupture of the four types of crystal lattices:

In the compound formation region, Fig. 36 shows that up to two peaks can appear on the thermograms. In this concentration range, all the crystal lattices must conform to a cell dimension corresponding to the arithmetic mean of the dimensions of the pure components (Ref. 28). Using again the example of palmitic-stearic system, the possible crystal lattice that can satisfy this constraint is crystal lattice type V. Since this compound is of the incongruent type, it transforms during melting to crystal type IV which is associated with the second peak.

In the region containing an excess of the lower-molecular weight

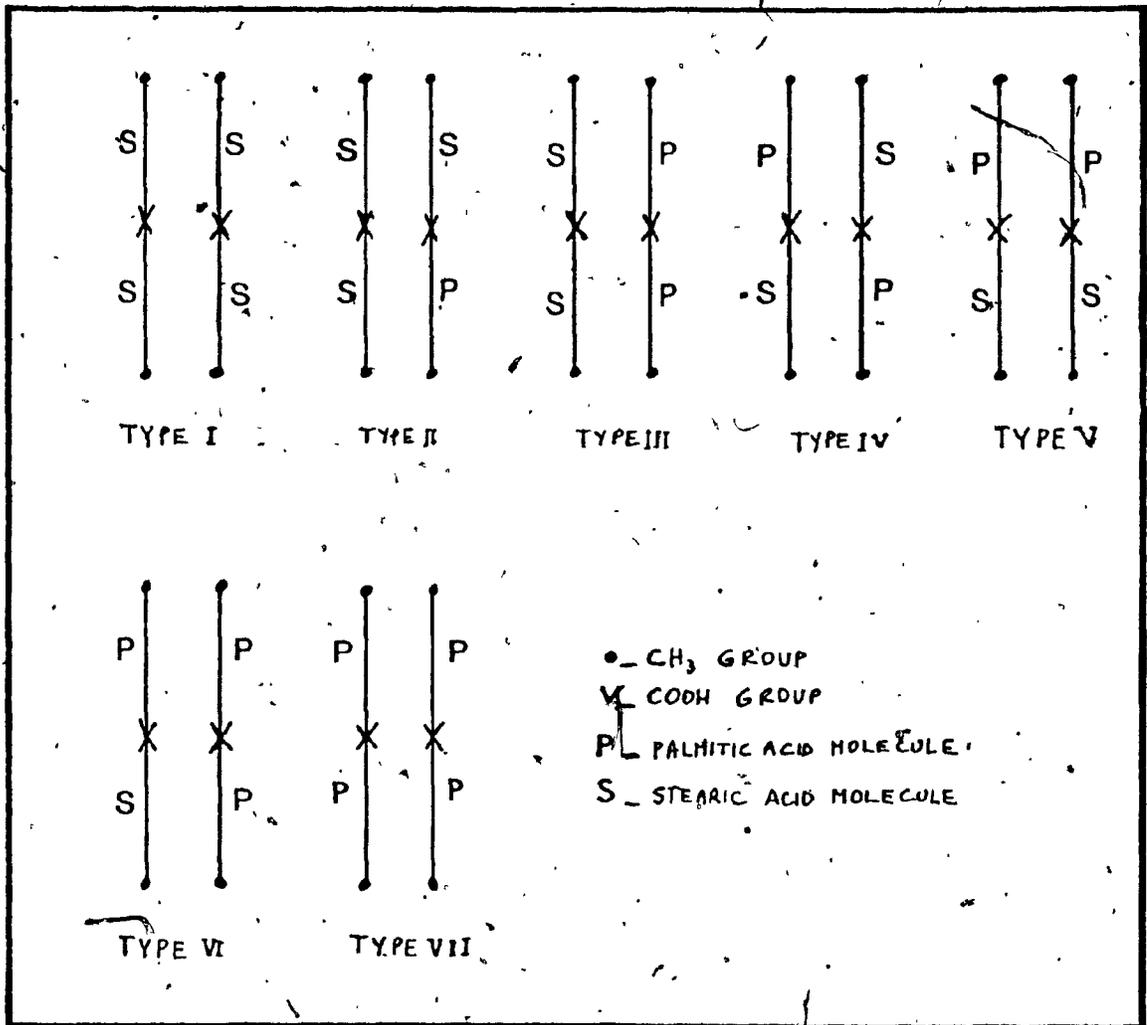


Fig. 37. Possible crystal lattice arrangements of binary mixtures of palmitic-stearic acids molecules.

component, i.e. between the eutectic point and 100% low-molecular weight component, Fig. 36 shows that up to two peaks are present on the thermograms. For the palmitic-stearic system Frede et al (Ref. 28) found that up to a concentration of 10 mole % palmitic: 90 mole % stearic, the longer stearic acid molecules can be incorporated in the crystal lattice of the palmitic acid molecule. In this concentration range, the two possible crystal lattices are types VI and VII in Fig. 37.

A similar analogy can be extended to the discussion of thermograms of other binary mixtures of fatty acids.

The analysis of the multiple peaks on the thermograms can also be considered from the standpoint of the possible influences of impurities present in the starting materials. However, due to the very low concentration of the impurities, it is possible that these effects are superimposed on the sharper peaks which are associated with the first-order transitions of materials.

XIII. CONCLUDING REMARKS

The melting point, freezing point, heat of melting and heat of crystallization of capric, lauric, palmitic and stearic acids and their binary mixtures have been determined by the Differential Scanning Calorimetry method. The melting points of capric, lauric, palmitic and stearic acids were observed to be  $30.0 \pm 0.4^\circ\text{C}$ ,  $42.2 \pm 0.1^\circ\text{C}$ ,  $59.0 \pm 0.6^\circ\text{C}$  and  $65.6 \pm 0.1^\circ\text{C}$  respectively. The corresponding freezing points of the fatty acids were found to be  $28.7 \pm 0.3^\circ\text{C}$ ,  $40.7 \pm 0.1^\circ\text{C}$ ,  $58.1 \pm 0.3^\circ\text{C}$  and  $64.5 \pm 0.2^\circ\text{C}$  respectively. The observed heats of melting of capric, lauric, palmitic and stearic acids were  $157 \pm 6$  J/g,  $164 \pm 4$  J/g,  $175 \pm 1$  J/g and  $183 \pm 3$  J/g. The corresponding heats of crystallization were  $153 \pm 7$  J/g,  $162 \pm 7$  J/g,  $173.3 \pm 3$  J/g and  $182 \pm 2$  J/g respectively.

The eutectic points were determined for binary mixtures of capric-lauric acids, lauric-palmitic acids, lauric-stearic acids and palmitic-stearic acids. The eutectic mixtures were examined by Differential Scanning Calorimetry to ascertain the thermal properties of melting point, freezing point, heat of melting and heat of crystallization. The eutectic compositions of the binary systems were found to be 73 mole % capric: 27 mole % lauric, 80 mole % lauric: 20 mole % palmitic, 85 mole % lauric: 15 mole % stearic and 73 mole % palmitic: 27 mole % stearic. The corresponding melting points of the eutectics were observed to be  $18.2^\circ\text{C}$ ,  $32.7^\circ\text{C}$ ,  $34.0^\circ\text{C}$  and  $51.1^\circ\text{C}$  respectively. The corresponding freezing points were observed to be  $16.6^\circ\text{C}$ ,  $30.3^\circ\text{C}$ ,  $31.3^\circ\text{C}$  and  $50.8^\circ\text{C}$  respectively.

The heats of melting of the eutectic compositions of binary systems of capric-lauric acids, lauric-palmitic acids, lauric-stearic acids and palmitic-stearic acids, lauric-stearic acids and palmitic-stearic acids were found to be 120 J/g, 147 J/g, 152 J/g and 159 J/g respectively. The corresponding heat of crystallization of the eutectics were found to be 113 J/g, 149 J/g, 155 J/g and 161 J/g respectively.

The pure fatty acids and their eutectic mixtures were examined with an X-ray diffractometer and an infrared spectrophotometer to establish the polymorphic form and crystal structure. The experimental results show that the acids are present in the stable C - polymorphic form and possess a monoclinic crystal structure.

The thermal properties of the fatty acids and their eutectics appear to be propitious for space heating applications. However, utilization of these materials for space heating and other applications depends on the development of an efficient and economic thermal energy storage system.

XIV. RECOMMENDATIONS FOR FURTHER WORK

Utilization of fatty acids for latent heat thermal storage applications will depend to a large extent on an efficient and economic design of the thermal storage system.

Saturated fatty acids possess a good chemical stability but, nonetheless, long-term stability ought to be verified by thermal-cycling (repeated melting and freezing of the material) to ensure the required longevity of the system.

The fatty acids possessing different grades of purity (such as research, reagent and technical grades) can be subjected to the thermal cycling technique and the constancy of the thermal parameters ought to be verified over a long period of time. If the chemical stability of the technical grade material is found comparable to the research grade material then the former should be selected due to a lower cost.

The supercooling effect is an important phenomenon and further work is required to characterize the liquid behavior at the freezing point. Several cooling rates can be used on samples having different purity levels and the relation between the cooling rate and supercooling be thus determined. For this purpose, the cooling accessory for the DSC cell should be fitted with a flow-meter to accurately measure the flow of the cooling gas.

The compatibility of fatty acids with conventional materials of construction is another area which requires further investigation. The evaluation of the material compatibility factor should have a

two-fold objective; namely, it should be considered from a corrosion point of view and the possible contamination of the fatty acid. Leached impurities from the container can combine with the fatty acids producing objectionable effects such as chemical decomposition. The decomposition, in turn, can affect the thermal properties of the acids as well as produce toxic or poisonous by-products. Some data has been published on the corrosion rates of fatty acids on conventional materials of construction (Ref. 1, 32, and 34). The materials which have a good corrosion resistance to fatty acids include aluminum, 304 stainless steel, fiberglass reinforced epoxy and polyester resin.

In the building construction industry, the flammability criterion is a crucial factor in the selection of materials. The widespread use of fatty acids will depend on how well these materials can meet the industry standards and municipal by-laws. The use of fire-retardant additives ought to be explored. To contain small fires, carbon dioxide gas, carbon tetrachloride and finely powdered sodium bicarbonate have been found effective. However, for large fires the use of foam is required.

The design and construction of heat exchangers requires further analysis. The geometry factor of the container must account for volume changes which occur during the phase transitions. Also, the factor of air-tightness should be examined to prevent water vapour and impurities from entering the system.

The experimental results on the binary mixtures of fatty acids indicate the formation of an incongruently melting compound.

Further work is required to determine the compound suitability for the thermal storage system.

Finally, the total package design of the thermal storage system must be cost-effective to favourably compete with alternate energy systems. Therefore, the economics and design of the system will depend on how well all the relevant parameters are optimized to meet the user criteria.

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APPENDIX

APPENDIX TABLES

TABLE A-1. Thermal Data of Pure Fatty Acids

SAMPLE	RUN NUMBER	DATA SCANNING RATE (SECONDS POINT)	MELTING			CRYSTALLIZATION <sup>1</sup>			SAMPLE WEIGHT	FILE NUMBER
			MELTING POINT (°C)	TEMPERATURE FOR MAX. ENDOOTHERMIC HEAT FLOW (°C)	HEAT OF MELTING (J/g)	FREEZING POINT (°C)	TEMPERATURE FOR MAX. EXOTHERMIC HEAT FLOW (°C)	HEAT OF CRYSTALLIZATION (J/g)		
Palmitic	1	0.2	59.6	61.4	177			2.113	Fuks 02/E3	
Palmitic	2	0.2	59.6	61.4	178			2.113	Fuks 03/E3	
Palmitic	3	0.4	59.6	61.4	176			2.113	Fuks 04/E3	
Palmitic	4	0.4	59.6	61.4	175			2.113	Fuks 05/E3	
Palmitic	5	0.6	59.7	61.4	176			2.113	Fuks 06/E3	
Palmitic	6	0.6	59.7	61.4	176			2.113	Fuks 07/E3	
Palmitic	7	0.4	59.1	61.5	175	58.3	58.2	4.114	Fuks 08/E3	
Palmitic	8	0.4	58.8	61.5	175	58.0	57.7	5.618	Fuks 27/1	
Palmitic	9	0.4	58.3	61.5	174	58.0	56.8	11.414	Fuks 02/3	
Avg. for Palmitic	-	-	59.0	61.5	175	56.1	57.6	173	-	
Stearic	1	0.4	64.5	67.8	185	64.3	63.4	180	Fuks 06/3	
Stearic	2	0.4	64.6	67.9	181	64.6	63.3	183	Fuks 07/3	
Avg. for Stearic	-	-	64.6	67.9	183	64.5	63.4	182	-	
Lauric	1	0.4	42.3	44.7	167	40.7	39.8	157	Dor 08/8	
Lauric	2	0.4	42.1	44.1	161	40.6	39.7	167	Fuks 03/3	
Avg. for Lauric	-	-	42.2	44.4	164	40.7	39.6	162	-	
Capric	1	0.4	30.2	32.4	152	28.9	28.5	151	Dor 10/8	
Capric	2	0.4	29.7	31.2	161	28.5	28.6	161	Fuks 01/5	
Avg. for Capric	-	-	30.0	31.8	157	28.7	28.6	156	-	

TABLE A-2. Thermal Data of Binary Mixtures of Palmitic and Stearic Acids

SAMPLE	MELTING			CRYSTALLIZATION			SAMPLE WEIGHT (mg)	FILE NUMBER
	MELTING POINT (°C)	TEMPERATURE FOR MAX. ENDOOTHERMIC HEAT FLOW (°C)	HEAT OF MELTING (J/g)	FREZZING POINT (°C)	TEMPERATURE FOR MAX. EXOTHERMIC HEAT FLOW (°C)	HEAT OF CRYSTALLIZATION (J/g)		
10P/90S	61.2	66.5	174	61.9	60.1	180	14.974	Fuks 16/4
20P/80S	57.4	63.8	174	-59.7	58.7	176	8.552	Fuks 15/4
30P/70S	50.9	60.8	171	56.7	55.8	174	9.974	Fuks 14/4
40P/60S	52.7	55.5	163	54.2	52.4	168	7.239	Fuks 13/4
50P/50S	52.9	56.5	163	52.6	50.5	158	14.04	Fuks 12/4
60P/40S	51.9	55.5	163	51.9	50.5	166	11.503	Fuks 11/4
69P/31S	51.4	53.8	160	51.0	49.8	166	3.357	Fuks 17/4
70P/30S	51.1	54.3	160	51.0	49.4	165	12.461	Fuks 10/4
71P/29S	51.0	53.7	159	51.0	50.0	161	6.278	Fuks 18/4
72.5S/27.5S	53.1	53.8	162	50.8	49.7	164	6.169	Fuks 19/4
74P/26S	51.3	54.1	161	51.1	50.1	162	6.457	Fuks 20/4
80P/20S	51.4	54.8	160	51.3	49.9	162	7.543	Fuks 09/4
90P/10S	53.8	58.6	170	54.3	52.8	162	4.496	Fuks 08/4

TABLE A-3. Thermal Data of Binary Mixtures of Capric and Lauric Acids

SAMPLE	MELTING			CRYSTALLIZATION			SAMPLE WEIGHT (mg)	FILE NUMBER	
	MELTING POINT (°C)	TEMPERATURE FOR MAX. ENDOTHERMIC HEAT FLOW (°C)	HEAT OF MELTING (J/g)	NUMBER OF PEAKS ON THERMOGRAM	FREEZING POINT (°C)	TEMPERATURE FOR MAX. EXOTHERMIC HEAT FLOW (°C)			HEAT OF CRYSTALLIZATION (J/g)
10C/90L	35.7	40.1	199.8	2	36.5	36.3	158	3	Fuks 26/5
20C/80L	19.9	37.0	151	3	32.7	31.9	153	3	Fuks 23/5
30C/70L	20.4	22.3	141	3	29.3	28.3	143	2	Fuks 23/5
40C/60L	21.2	24.9	134	4	22.0	20.5	133	2	Dor 07/7
50C/50L	21.8	24.0	133	2	21.0	19.9	133	2	Fuks 05/1
55C/45L	21.7	24.0	128	2	20.0	18.8	131	3	Fuks 13/1
60C/40L	19.0	23.4	132	2	19.7	18.5	125	2	Dor 11/8 (C) Dor 12/8 (A)
65C/35L	19.3	22.3	127	1	19.0	18.1	129	2	Fuks 14/1
70C/30L	18.4	21.2	126	1	18.2	17.4	118	1	Dor 05/7
71C/29L	18.6	22.0	125	1	18.0	16.4	115	1	Dor 04/6
72C/28L	18.7	22.0	126	1	17.2	16.3	116	1	Dor 13/6
72.55/27.5L	18.5	22.0	120	1	17.9	16.2	111	1	Dor 04/7
73C/27L	18.2	20.8	124	1	15.6	16.4	116	1	Dor 03/6
74C/26L	18.8	21.2	121	1	16.8	15.8	120	1	Fuks 29/4
75C/25L	18.5	21.6	122	1	17.5	16.3	114	1	Dor 08/6
80C/20L	19.5	21.7	122	1	17.8	16.7	120	1	Fuks 03/1
90C/10L	21.9	28.4	135	2	23.3	22.8	124	2	Fuks 06/1

TABLE A-4. Thermal Data of Binary Mixtures of Lauric and Palmitic Acids

SAMPLE	MELTING			CRYSTALLIZATION			FILE NUMBER		
	MELTING POINT (°C)	TEMPERATURE FOR MAX. ENDOOTHERMIC HEAT FLOW (°C)	HEAT OF MELTING (J/g)	NUMBER OF PEAKS ON THERMOGRAM	FREEZING POINT (°C)	TEMPERATURE FOR MAX. EXOTHERMIC HEAT FLOW (°C)		HEAT OF CRYSTALLIZATION (J/g)	NUMBER OF PEAKS ON THERMOGRAM
10L/90P	55.7	59.5	171	2	56.7	55.2	173	2	Fuks 40/3 (C)/Fuks 49/3 (M)
20L/80P	50.8	57.2	172	4	52.6	52.2	171 <sup>a</sup>	4	Fuks 01/2
30L/70P	35.0	54.6	170	4	49.1	48.2	169	4	Fuks 27/2 (M)/Fuks 01/3 (C)
40L/60P	36.3	38.8	166	4	45.6	45.3	166	4	Fuks 03/2
50L/50P	37.4	39.4	165	4	40.6	40.2	164	4	Fuks 04/2
60L/40P	33.7	40.6	161	3	36.0	34.2	158	2	Dor 11/9
70L/30P	33.9	36.5	156	2	33.0	30.2	157	2	Fuks 06/2
71L/29P	34.2	36.4	152	2	33.0	30.8	156	2	Fuky 07/2 (M)/Fuks 08/2 (C)
72.5L/27.5P	33.9	35.7	150	2	32.0	30.7	155	2	Fuks 13/2
75L/25P	33.8	36.3	154	2	31.4	30.2	154	1	Fuks 05/2
77.5L/22.5P	33.2	36.0	148	1	31.0	30.0	150	1	Fuks 12/2
79L/21P	32.8	35.6	151	1	30.6	29.3	149	1	Fuks 29/2 (C)/Fuks 30/2 (M)
80L/20P	32.7	35.6	150	1	30.3	29.1	151	1	Fuks 26/2
81L/19P	33.3	36.4	151	1	30.6	29.1	152	1	Fuks 21/2 (C)/Fuks 31/2 (M)
82.5L/17.5P	33.1	36.2	146	1	30.8	30.3	149	1	Fuks 23/2 (C)/Fuks 24/2 (M)
85L/15P	33.7	36.9	149	1	31.6	30.7	152	1	Fuks 11/2
90L/10P	34.2	37.1	153	2	34.0	33.7	153	2	Fuks 10/2

TABLE A-5. Thermal Data of Binary Mixtures of Lauric and Stearic Acids

SAMPLE	MELTING			CRYSTALLIZATION			SAMPLE WEIGHT (mg)	FILE NUMBER
	MELTING POINT (°C)	TEMPERATURE FOR MAX. ENDOOTHERMIC HEAT FLOW (°C)	HEAT OF MELTING (J/g)	FREEZING POINT (°C)	TEMPERATURE FOR MAX. EXOTHERMIC HEAT FLOW (°C)	HEAT OF CRYSTALLIZATION (J/g)		
10L/90S	60.7	65.9	182	61.9	61.5	180	8.08	Fuks 08/3
20C/80S	57.4	63.9	183	59.6	58.8	179	7.307	Fuks 09/3
30L/70S	53.7	61.5	182	56.5	56.5	180	8.557	Fuks 10/3
40L/60S	31.2	59.2	178	53.4	52.4	171	10.623	Dor 19/9
50L/50S	32.5	35.7	174	50.2	49.7	171	7.676	Dor 20/9
60L/40S	33.4	35.7	155	45.3	44.8	165	4.974	Fuks 01/4
70L/30S	34.2	36.1	163	37.9	32.4	163	3.499	Fuks 02/4
80L/20S	34.0	36.4	162	34.3	32.6	160	3.4	Fuks 03/4
82.5L/17.5S	34.0	37.1	157	32.9	31.4	161	8.496	Fuks 22/4
84L/16S	33.9	37.1	157	32.3	31.1	161	7.999	Fuks 23/4
85L/15S	34.0	36.7	156	32.0	31.3	158	5.038	Fuks 06/4
86L/14S	34.2	37.5	152	31.3	30.8	157	3.973	Fuks 24/4
87.5L/12.5S	34.2	37.3	151	32.2	31.8	155	8.152	Fuks 25/4
90L/10S	34.5	37.7	155	33.3	32.1	155	8.404	Fuks 04/4

30 MOLE % CAPRIC: 70 MOLE % LAURIC

Sample: 30C/70L  
Size: 0.850 MG  
Rate: 1C/MIN  
Program: Interactive DSC V2.8  
Date: 8-Jun-83 Time: 14:52:15  
Files: FLKS.28  
Operator:  
Plotted: 8-Jun-83 15:48:33

### DSC

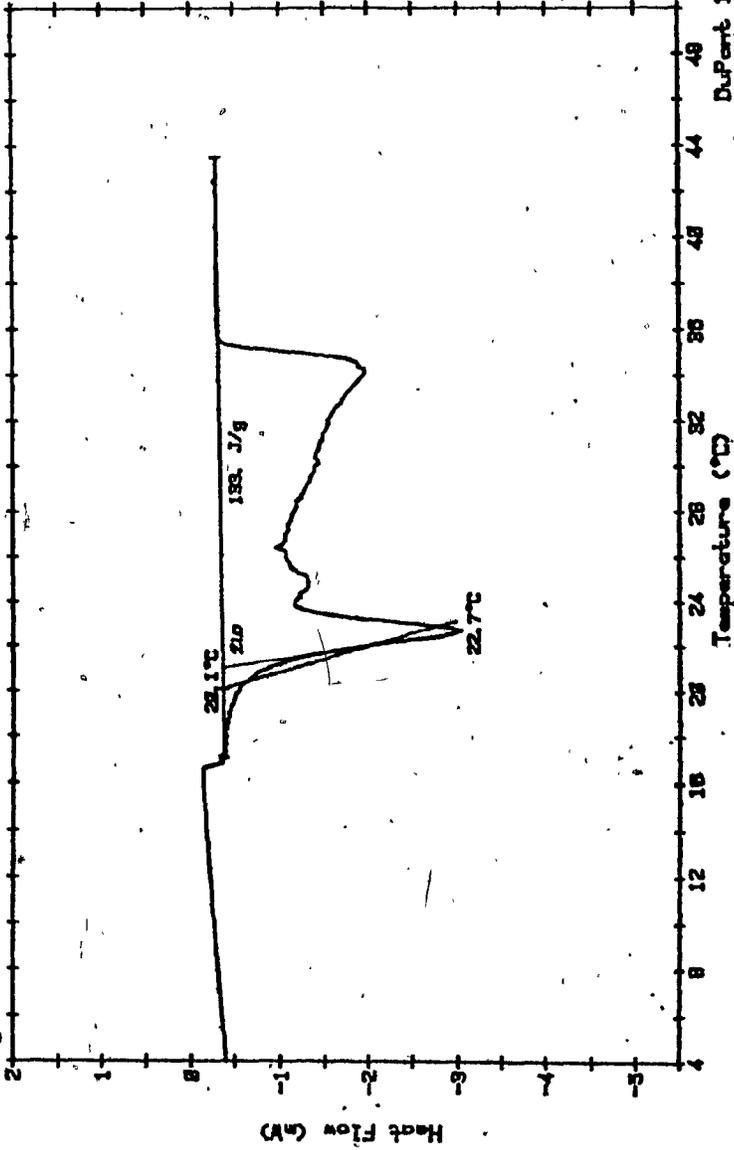


Fig. T-1. Thermogram of 30 mole % capric:70 mole % lauric  
( 1°C/min heating rate )  
DuPont 1990

Date: 8-Jun-83 Time: 14:16:12

File: FLKS.25

Operator:

Plotted: 8-Jun-83 14:48:24

### DSC

Sample: 30C/70L

Size: 8.850 MG

Rate: 2C/MIN

Program: Interactive DSC V2.8

START: 8.2  
STOP: 48.4

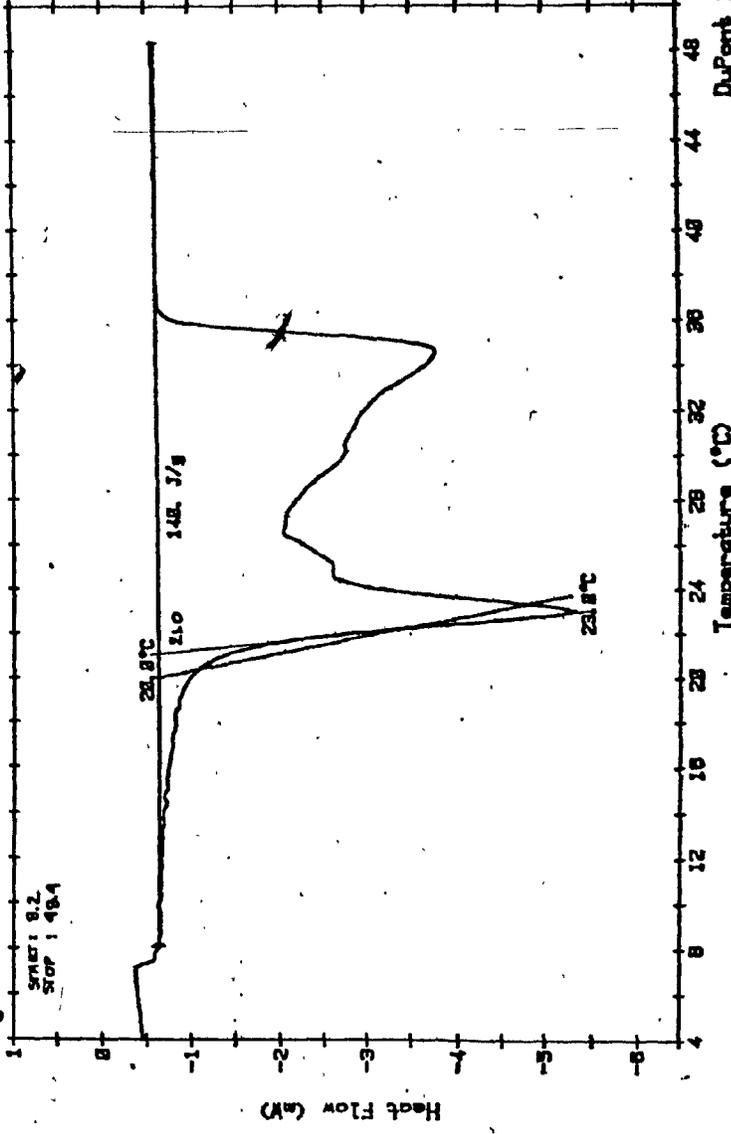


Fig. T-2. Thermogram of 70 mole % capric : 30 mole % lauric  
( 2C/min heating rate )

Samples 30C/70L  
Size: 8.850 MG  
Rate: 5C/MIN  
Program: Interactive DSC V2.8  
Date: 8-Jun-83  
Time: 13:21:38  
Files: FUMS.22  
Operator:  
Plotted: 8-Jun-83 13:48:38

# DSC

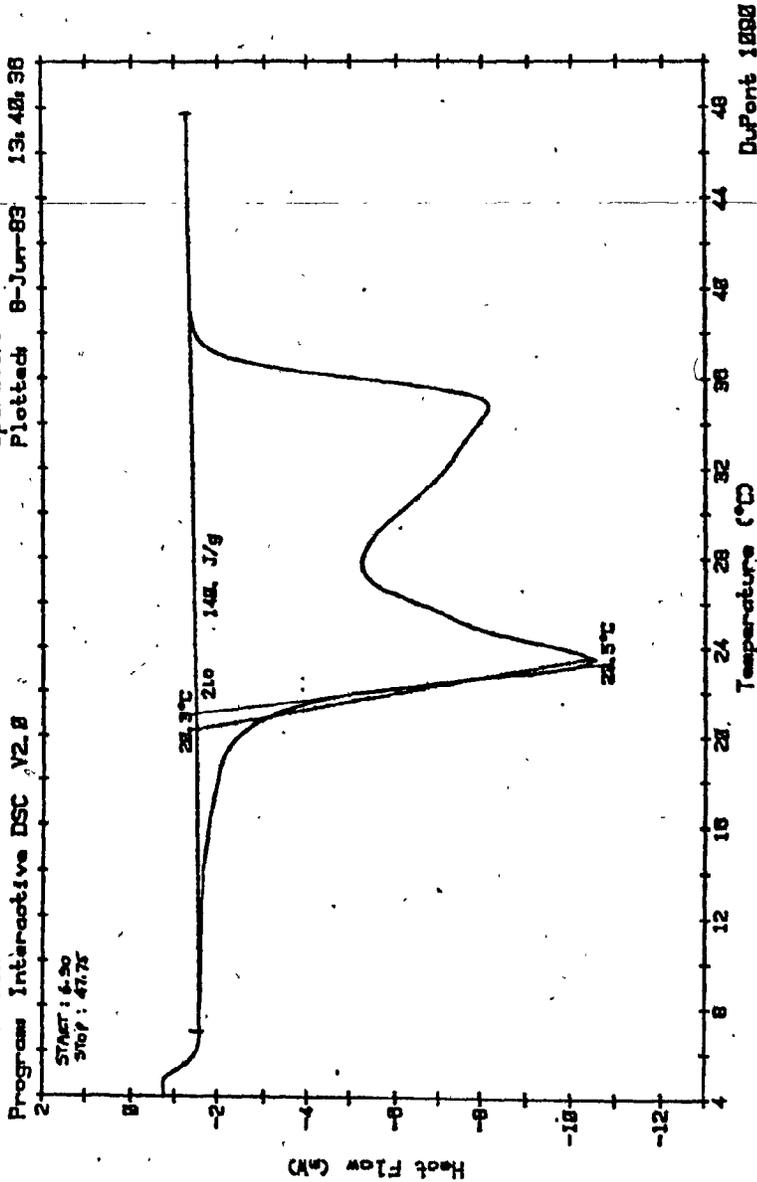


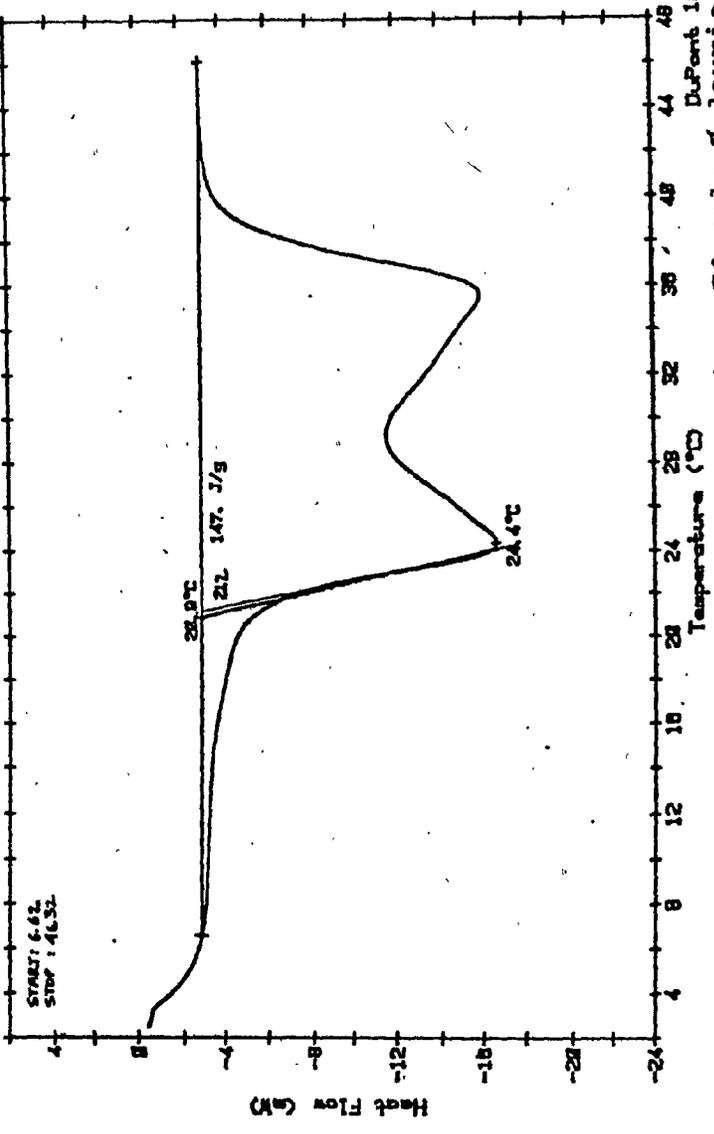
Fig. T-3. Thermogram of 30 mole % capric : 70 mole % lauric  
( 5 °C/min heating rate )

Date: 8-Jun-83 Time: 13:47:05  
File: FUKS.29  
Operator:  
Plotted: 8-Jun-83 13:58:23

### DSC

Sample: 30C/70L  
Size: 0.858 MG  
Rate: 10C/MIN  
Program: Interactive DSC V2.0

START: 6.62  
STOP: 46.52



DuPont 1090

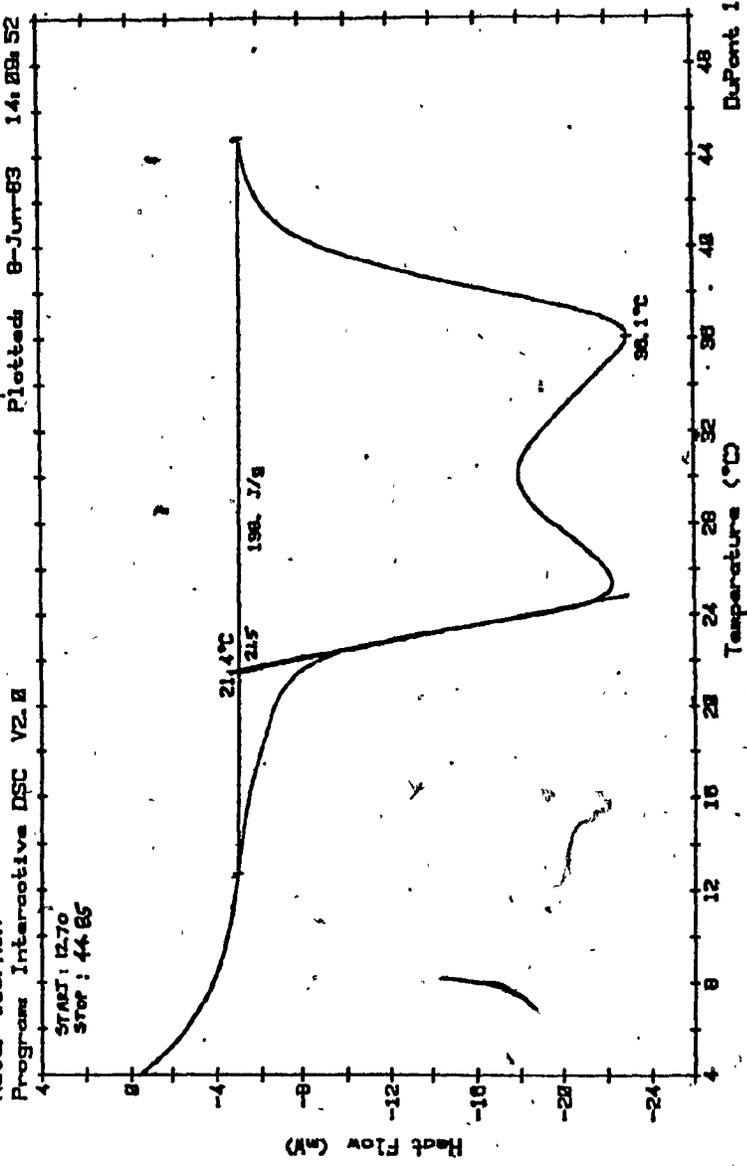
Fig. T-4. Thermogram of 30 mole % caprylic : 70 mole % lauric

( 10°C/min heating rate )

Samples 30C/70L  
Size: 8.858  
Rate: 15C/MIN  
Program: Interactive DSC V2.0  
START: 12.70  
STOP: 44.85

# DSC

Date: 8-Jun-83 Time: 14:02:28  
File: FUMS.24  
Operator:  
Plotted: 8-Jun-83 14:08:52



DuPont 1888

Fig. T-5. Thermogram of 30 mole % capric : 70 mole % lauric  
( 15°C/min heating rate )

PURE FATTY ACIDS

Sampler CA  
Size 11.54MG  
Rate 2C/MIN  
Program Interactive DSC V2.8

**DSC**

Date 25-May-83 Time 15:48:30  
File DORINA.18 DISK EIGHT  
Operator  
Plotted 11-Sep-83 14:18:38

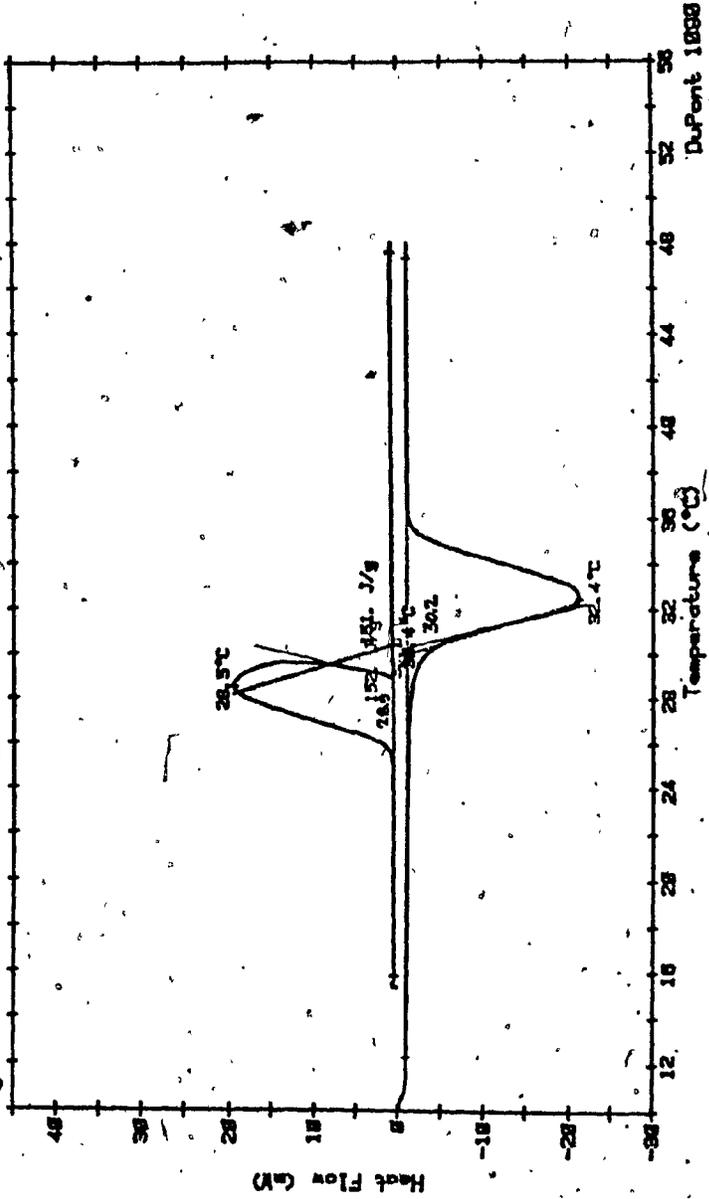


Fig. T-6. Thermogram of capric acid ( 11.54 mg )

Date: 15-Jul-83 Time: 13:43:14  
File: FUKS.01 DISC FIVE  
Operator:  
Plotted: 15-Jul-83 14:25:58

Sampler: 100 CA  
Size: 3.114 MG  
Rate: 2 C/MIN  
Program: Interactive DSC V2.0

### DSC

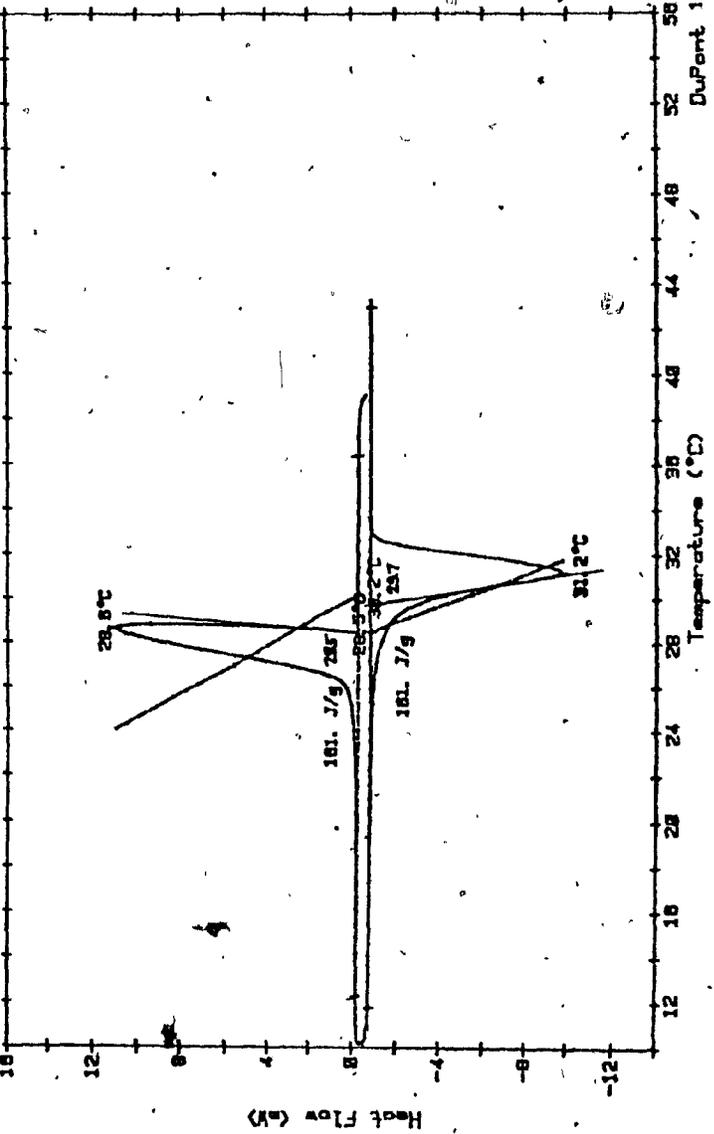


Fig. T-7. Thermogram of capric acid ( 3.114 mg )

Sample: L.A. BACKER  
Size: 8.24MG  
Rate: 2C/MIN  
Program: Interactive DSC V2.8  
Date: 25-May-83 Time: 13:18:00  
File: DORINA.08 DISK EIGHT  
Operator:  
Plotted: 11-Sep-83 13:38:45

DSC

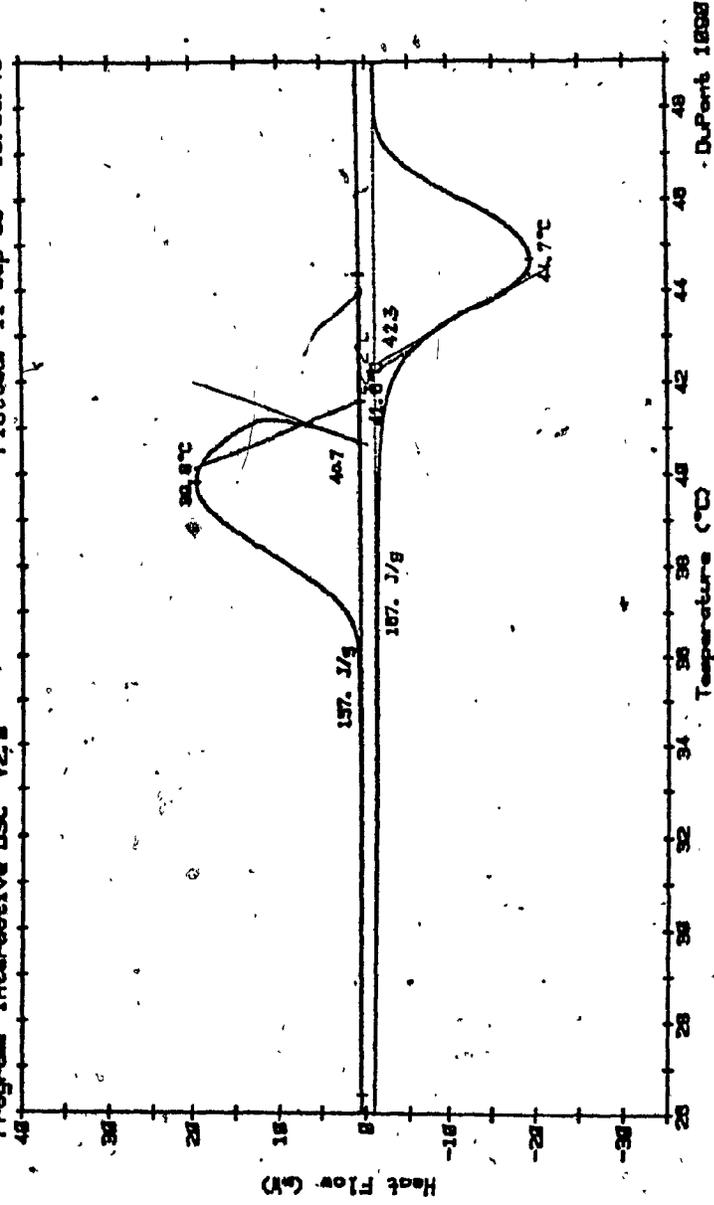


Fig. T-8. Thermogram of lauric acid ( 9.24 mg )

Sample: 108 LAU  
Size: 7.838 MG  
Rate: 2 C/MIN  
Program: Interactive DSC V2.8  
Date: 4-Jul-83 Time: 13:08:58  
File: FLKS.83 DISK THREE  
Operator:  
Plotted: 11-Sep-83 13:24:34

### DSC

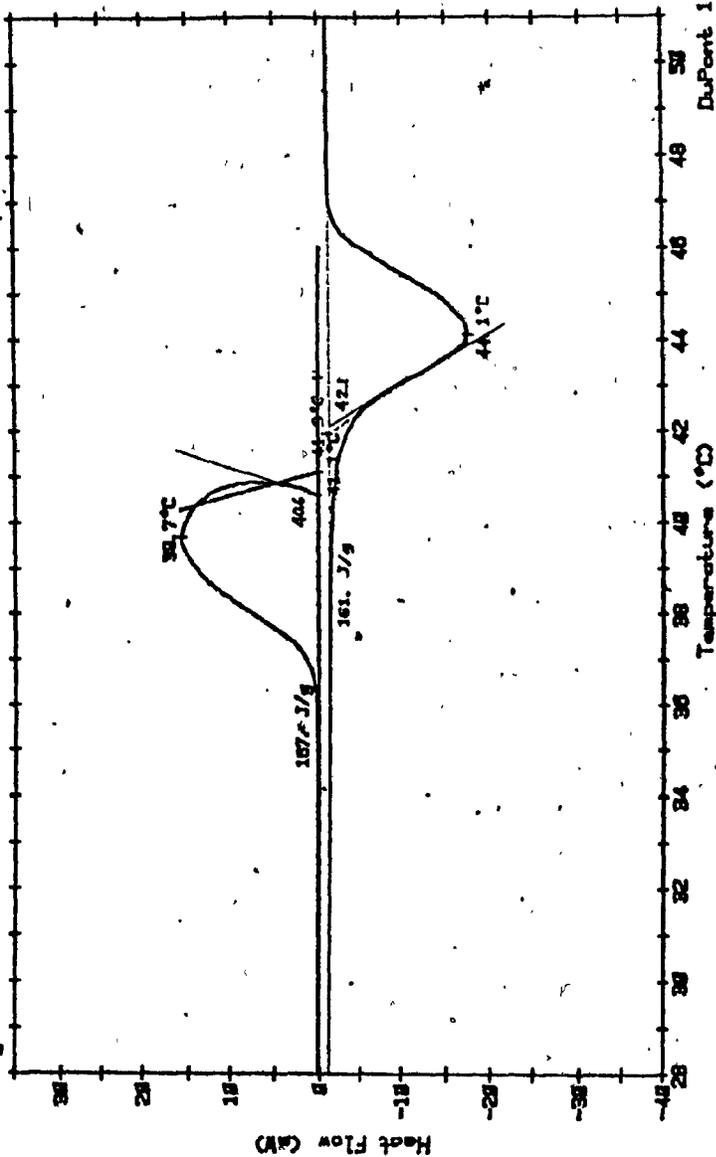


Fig. T-9. Thermogram of lauric acid ( 7.83 mg )



Date: 6-Jul-83 Time: 21:33:34  
File: FUKS.07 DISK THREE  
Operator:  
Plotted: 6-Jul-83 22:20:06

Sample: 100 ST  
Size: 7.465 MG  
Rate: 2 C/MIN  
Program: Interactive DSC V2.0

# DSC

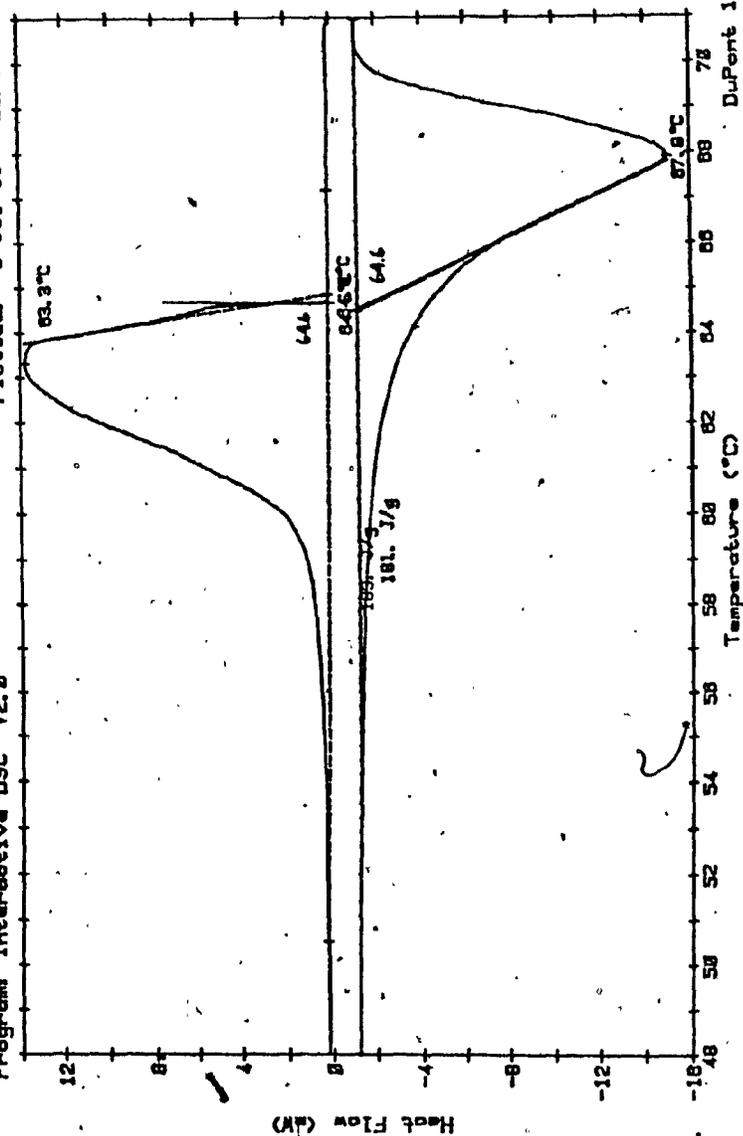


Fig. T-11. Thermogram of stearic acid (7.465 mg, run #2)

Date: 4-Jul-83 Time: 11:58:50  
File: FUKS.02 DISK THREE  
Operator:  
Plotted: 4-Jul-83 12:40:24

Samples 100 PAL  
Size: 11.414 MG  
Rate: 2 C/MIN  
Program: Interactive DSC V2.0

### DSC

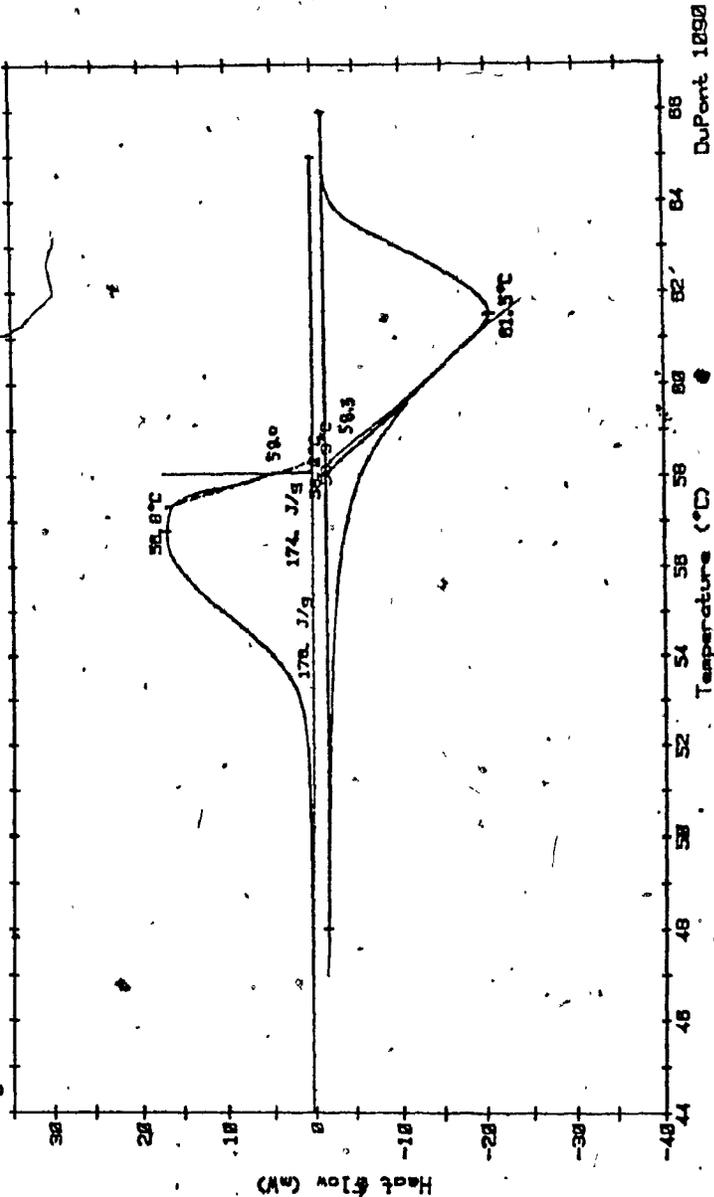


Fig. T-12. Thermogram of palmitic acid ( 11.414 mg )

Samples 100 PAL Dates 9-Jun-83 Times 11:58:41

Size 5.618 MG Files FUKS.27

Rate 20/MIN Operator  
Program Interactive DSC V2.8 Plotted 5-Sep-83 16:01:23

### DSC

Heat Flow (mW)

40

30

20

10

0

-10

-20

-30

-40

-50

-60

-70

-80

-90

-100

-110

-120

-130

-140

-150

-160

-170

-180

-190

-200

-210

-220

-230

-240

-250

-260

-270

-280

-290

-300

-310

-320

-330

-340

-350

-360

-370

-380

-390

-400

-410

-420

-430

-440

-450

-460

-470

-480

-490

-500

-510

-520

-530

-540

-550

-560

-570

-580

-590

-600

-610

-620

-630

-640

-650

-660

-670

-680

-690

-700

-710

-720

-730

-740

-750

-760

-770

-780

-790

-800

-810

-820

-830

-840

-850

-860

-870

-880

-890

-900

-910

-920

-930

-940

-950

-960

-970

-980

-990

-1000

-1010

-1020

-1030

-1040

-1050

-1060

-1070

-1080

-1090

-1100

-1110

-1120

-1130

-1140

-1150

-1160

-1170

-1180

-1190

-1200

-1210

-1220

-1230

-1240

-1250

-1260

-1270

-1280

-1290

-1300

-1310

-1320

-1330

-1340

-1350

-1360

-1370

-1380

-1390

-1400

-1410

-1420

-1430

-1440

-1450

-1460

-1470

-1480

-1490

-1500

-1510

-1520

-1530

-1540

-1550

-1560

-1570

-1580

-1590

-1600

-1610

-1620

-1630

-1640

-1650

-1660

-1670

-1680

-1690

-1700

-1710

-1720

-1730

-1740

-1750

-1760

-1770

-1780

-1790

-1800

-1810

-1820

-1830

-1840

-1850

-1860

-1870

-1880

-1890

-1900

-1910

-1920

-1930

-1940

-1950

-1960

-1970

-1980

-1990

-2000

-2010

-2020

-2030

-2040

-2050

-2060

-2070

-2080

-2090

-2100

-2110

-2120

-2130

-2140

-2150

-2160

-2170

-2180

-2190

-2200

-2210

-2220

-2230

-2240

-2250

-2260

-2270

-2280

-2290

-2300

-2310

-2320

-2330

-2340

-2350

-2360

-2370

-2380

-2390

-2400

-2410

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-2460

-2470

-2480

-2490

-2500

-2510

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-2530

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-2570

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-2590

-2600

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-2680

-2690

-2700

-2710

-2720

-2730

-2740

-2750

-2760

-2770

-2780

-2790

-2800

-2810

-2820

-2830

-2840

-2850

-2860

-2870

-2880

-2890

-2900

-2910

-2920

-2930

-2940

-2950

-2960

-2970

-2980

-2990

-3000

-3010

-3020

-3030

-3040

-3050

-3060

-3070

-3080

-3090

-3100

-3110

-3120

-3130

-3140

-3150

-3160

-3170

Sample 100X PAL  
Size 4.14 MG  
Rate 2 C/MIN  
Program Interactive DSC V2.0  
Date 15-Sep-83 Time 20:22:29  
File FUKS.88 DISKTHREE  
Operator  
Plotted 15-Sep-83 21:01:04

DSC

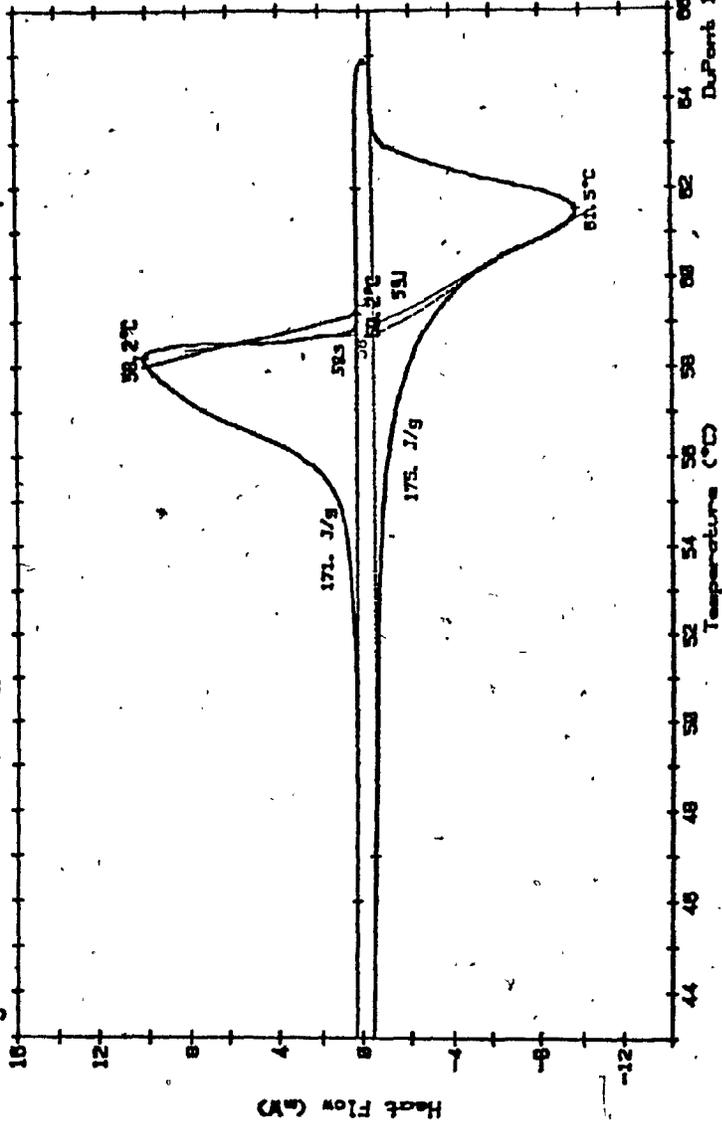


Fig. T-14. Thermogram of palmitic acid ( 4.14 mg )

Sample: 100X PAL  
Size: 2.113 MG  
Rate: 2C/MIN  
Program: Interactive DSC Y2.B

### DSC

Date: 14-Sep-83 Time: 21:18:52  
File: FUKS.02 DISKTHREE  
Operator:  
Plotted: 14-Sep-83 23:28:32

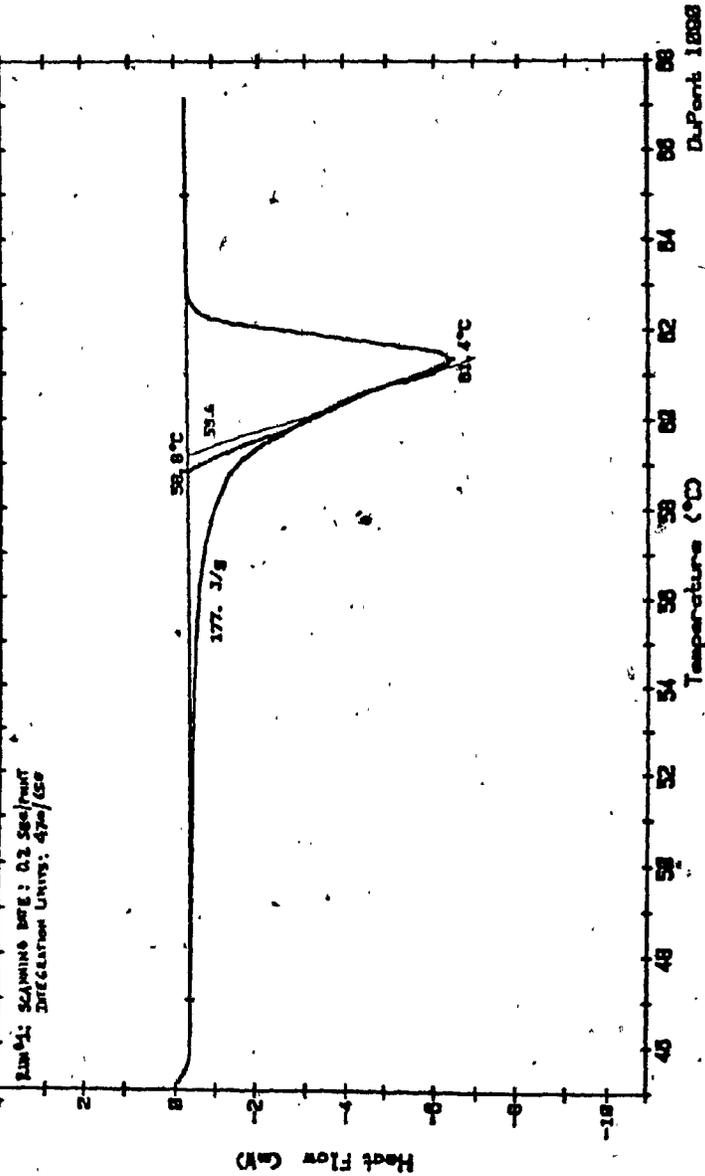


Fig. T-15. Thermogram of palmitic acid ( run #1, scanning rate: 0.2 sec/point )

Sample 188X PAL  
Size 2.113 MG  
Rate 2 C/MIN  
Program Interactive DSC V2.8  
Date 14-Sep-83 Time 21:48:07  
File FLMS.03 DISKTHREE  
Operator  
Plotted 14-Sep-83 22:03:32

### DSC

RUN #2: SCANNING RATE: 0.2 SEC/POINT  
INTEGRATION LIMITS: 470/650

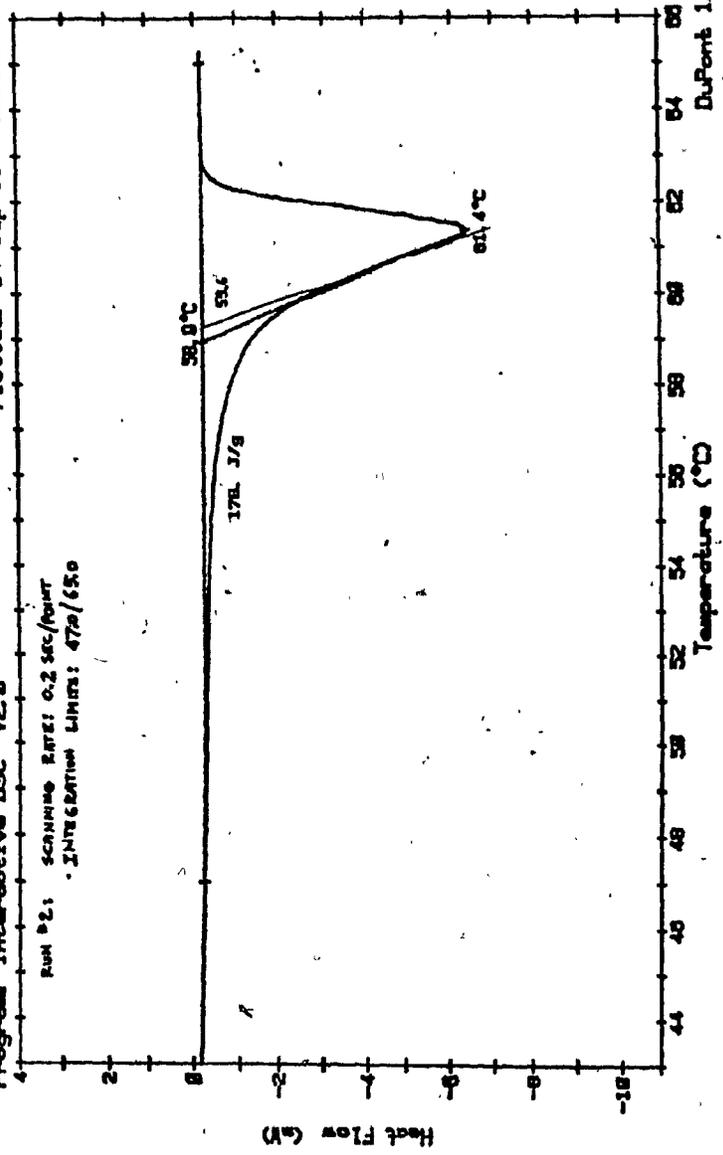
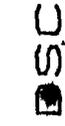


Fig. T-16. Thermogram of palmitic acid ( run #2, scanning rate: 0.2 sec/point )

Sample 100 XPAL  
Size 2.113 MG  
Rate 2 C/MIN  
Program Interactive DSC V2.8



Date 14-Sep-83 Time 22:07:48  
File FIMS.04 DISKTHREE  
Operator  
Plotted 14-Sep-83 22:24:43

RUN # : SCANNING RATE : 0.4 sec/point  
INTEGRATION LIMITS : 47.0/650

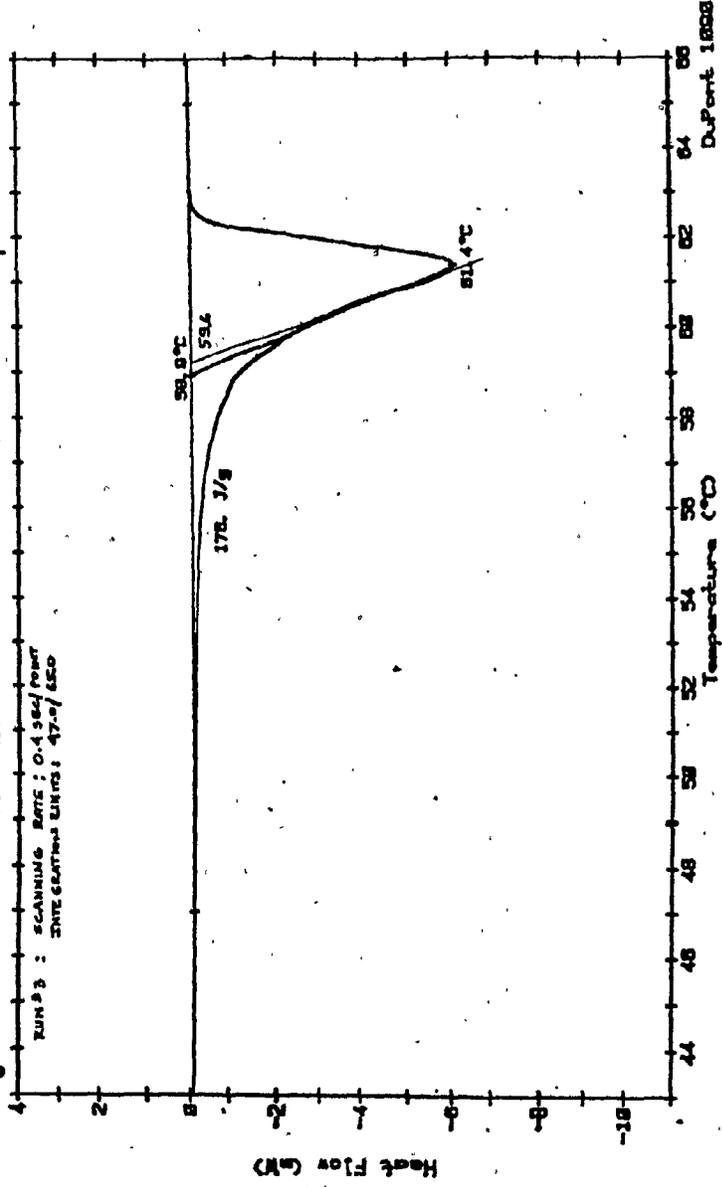


Fig. T-17. Thermogram of palmitic acid ( run #3, scanning

rate: 0.4 sec/point )

Date: 14-Sep-83 Time: 22:28:23  
File: FUKS.05 DISKTHREE  
Operator:  
Plotted: 14-Sep-83 22:44:44

# DSC

Sample: 100 PAL  
Size: 2.113 MG  
Rate: 2 C/MIN  
Program: Interactive DSC V2.8  
Run #4: SCANNING RATE: 0.4 SEC/POINT  
INTEGRATION LIMITS: 47.0/45.0

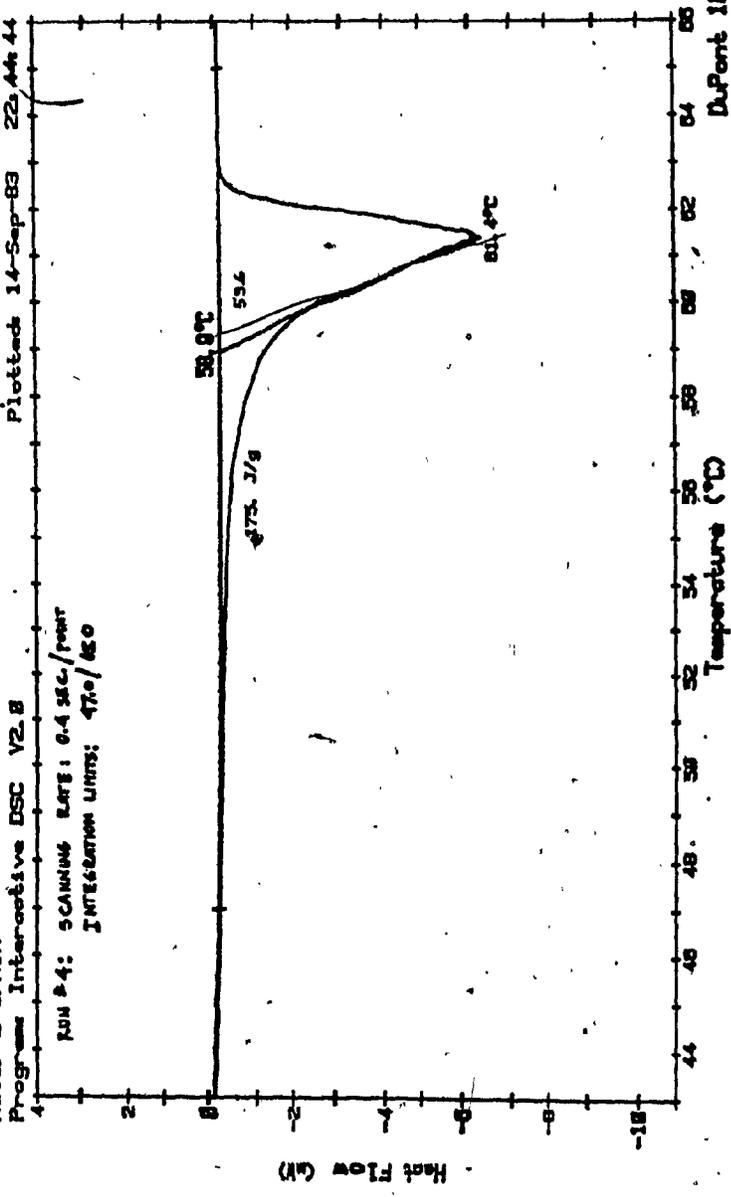


Fig. T-18. Thermogram of palmitic acid ( run #4, scanning rate: 0.4 sec/point )

Sample 100X PAL  
Size 2.113 MG  
Rate 2 C/MIN  
Program Interactive DSC V2.0  
Date 14-Sep-83 Time 22:48:03  
File FUKS.08 DISKTHREE  
Operator  
Plotted 14-Sep-83 23:04:21

### DSC

RUN 05: SCANNING RATE: 0.6 SEC/POINT  
INTEGRATION UNITS: 47.0/15.0

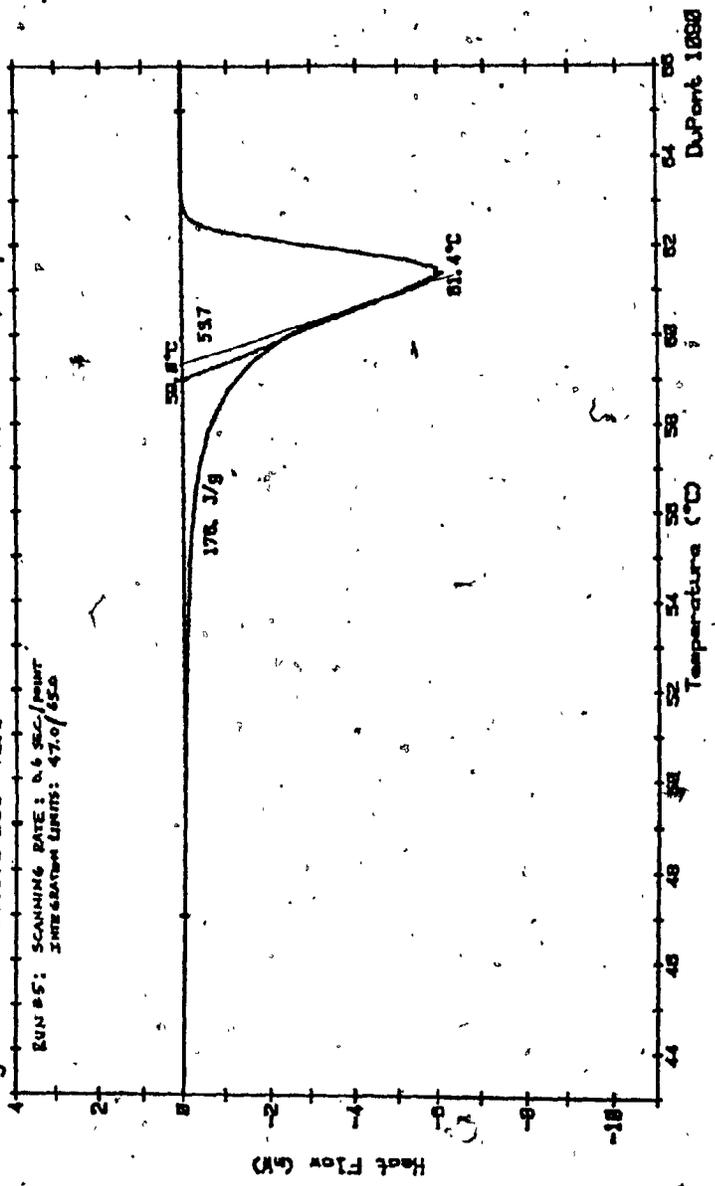


Fig. T-19. Thermogram of palmitic acid (run #5, scanning rate: 0.6 sec/point)

Date: 14-Sep-89 Time: 23:07:42  
Film: FKS:87 DISKTHREE  
Operator:  
Plotted: 14-Sep-89 23:23:48

### DSC

Sample: 100X PAL  
Size: 2.113 MG  
Rate: 2 C/MIN  
Program: Interactive DSC V2.0

Scan Rate: 0.6 sec/point  
Integrator Gain: 47.5/10

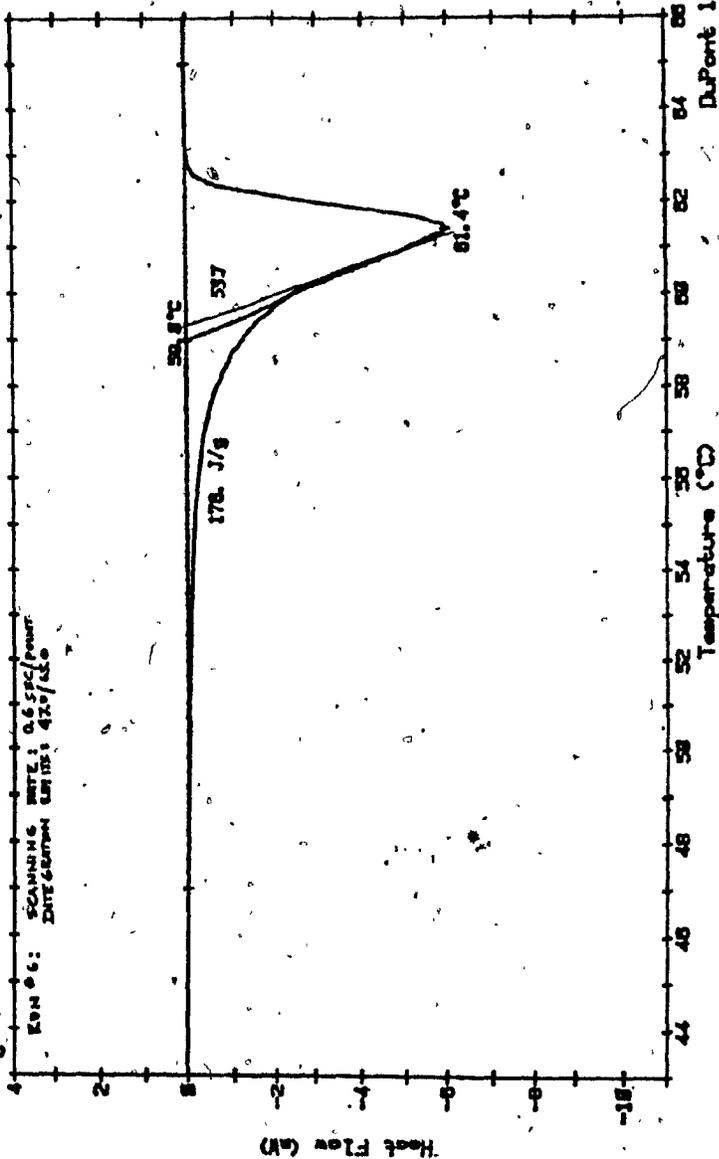


Fig. T-20. Thermogram of palmitic acid ( run #6, scanning rate: 0.6 sec/point )

BINARY MIXTURES OF CAPRIC-LAURIC ACIDS

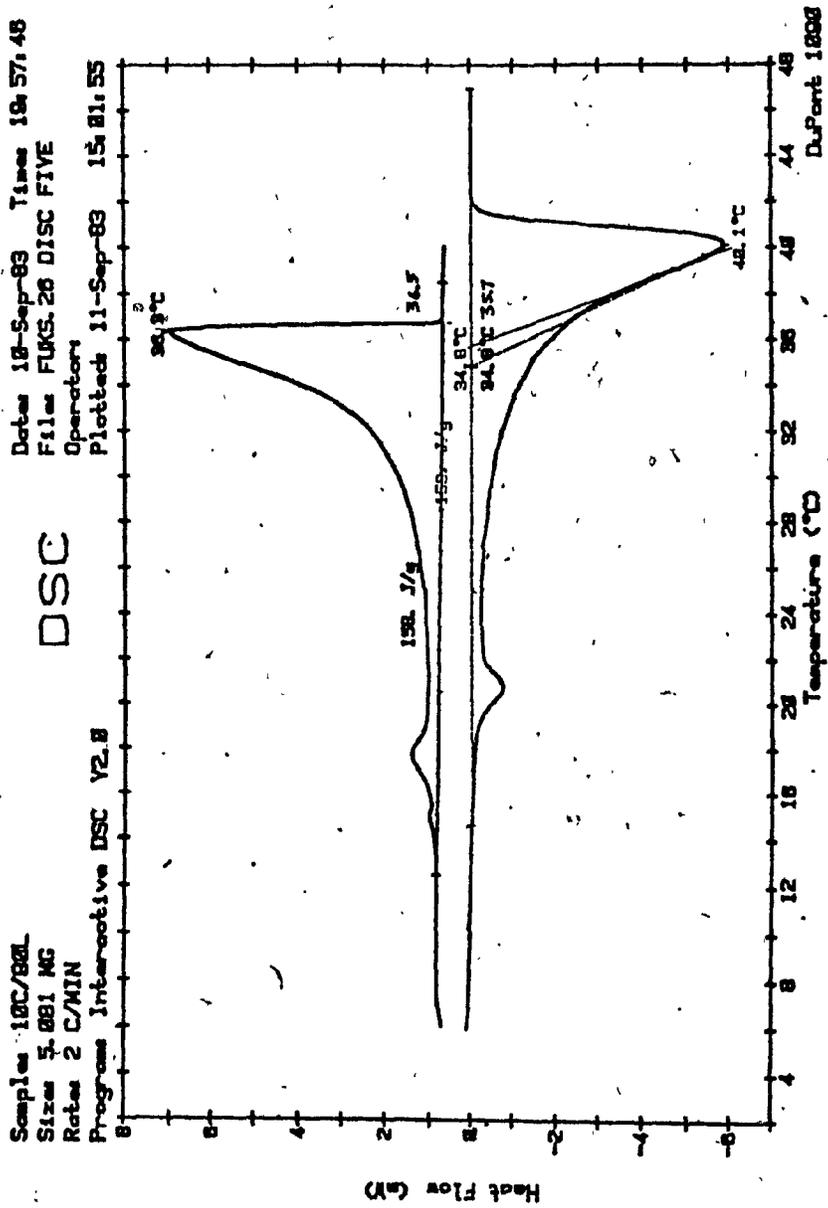


Fig. T-21. Thermogram of 10 mole % capric : 90 mole % lauric

7

Date: 10-Sep-83 Time: 16:46:27  
File: FLKS.23 DISC FIVE  
Operator:  
Plotted: 10-Sep-83 17:58:37

### DSC

Sample: 20C/80L  
Size: 5.176 MG  
Rate: 2 C/MIN  
Program: Interactive DSC V2.8

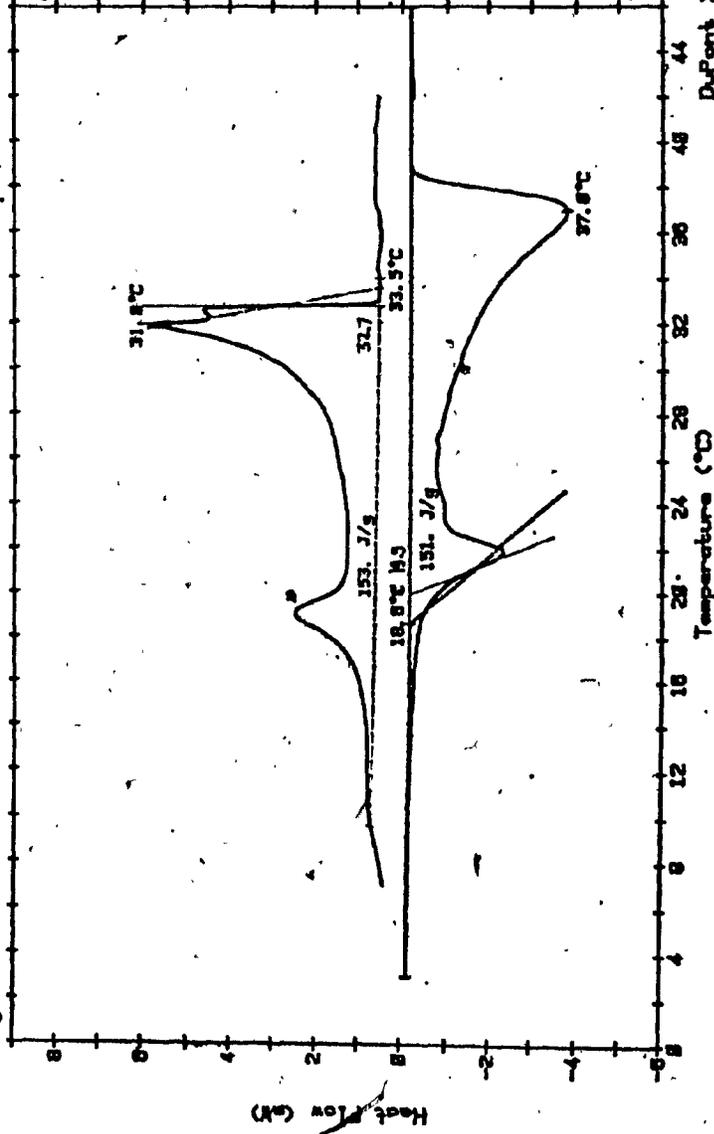


Fig. T-22. Thermogram of 20 mole % capric : 80 mole % lauric

Sample 30C/7BL  
Size 4.478 MG  
Rate 2C/MIN  
Program Interactive DSC V2.0

# DSC

Date 18-Sep-83 Time 18:23:55  
File FLKS.24 DISC FIVE  
Operator  
Plotted 18-Sep-83 18:31:01

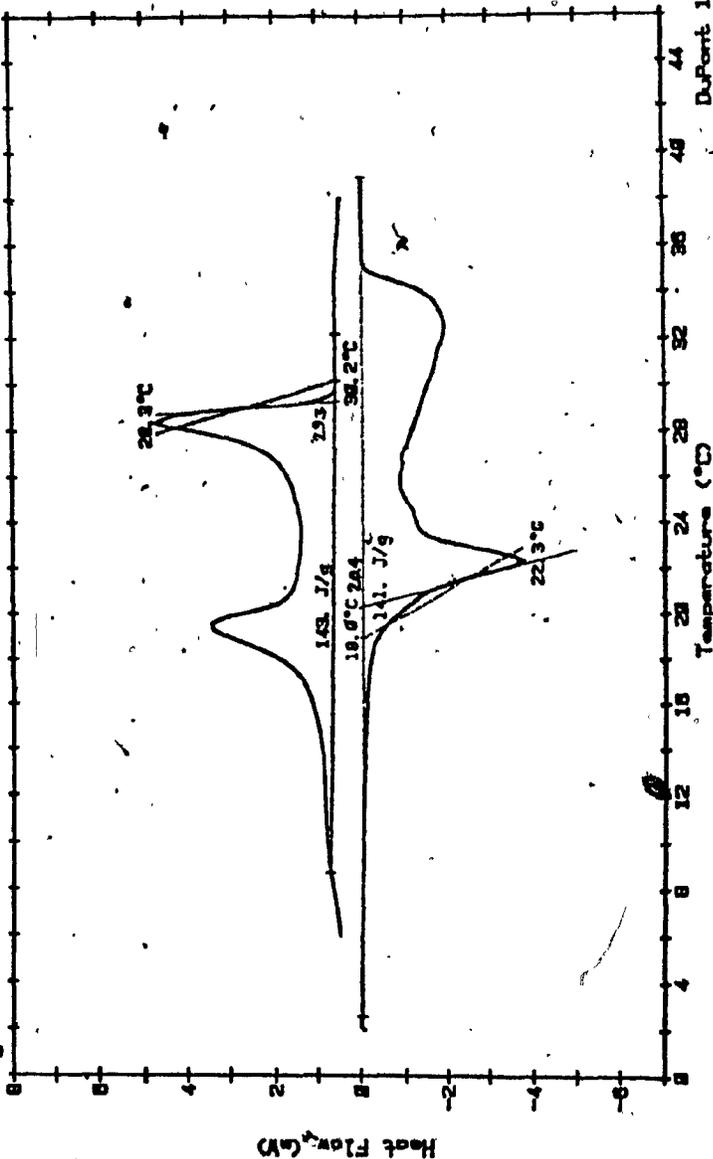


Fig. T-23. Thermogram of 30 mole % capric : 70 mole % lauric

Sample 48XCA+80XLA  
Size 18.58  
Rate 2C/MIN  
Program Interactive DSC V2.8  
Date 5-May-83 Times 14:16:18  
File DORINA.87 DISK SEVEN  
Operator  
Plotted 11-Sep-83 15:28:14

DSC

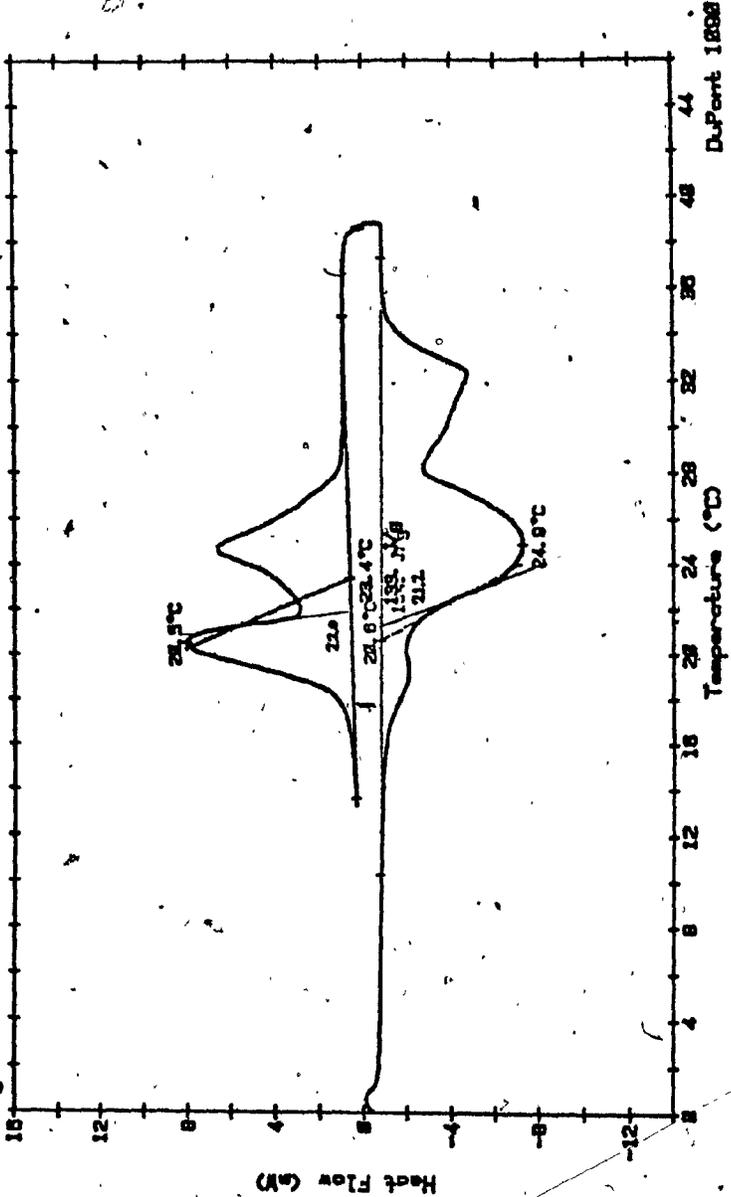


Fig. T-24. Thermogram of 40 mole % capric : 60 mole % lauric. DuPont 1888

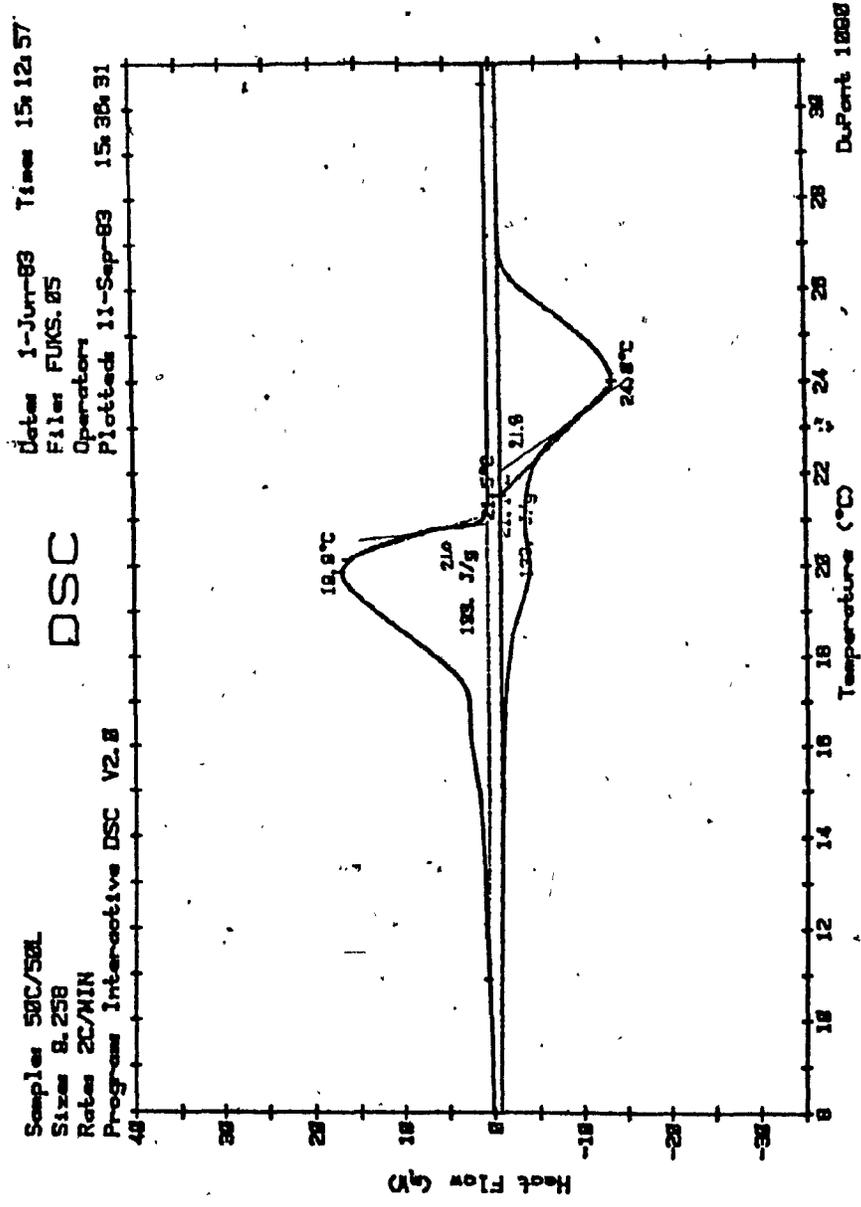


Fig. T-25. Thermogram of 50 mole % capric : 50 mole % lauric.

Sampler 55C/45L  
Size: 8.145 MG  
Rate: 2C/MIN  
Program: Interceptive DSC V2.0

Date: 8-Jun-83 Time: 12:18:35  
File: FUKS.13  
Operator:  
Plotted: 8-Sep-83 22:42:23

# DSC

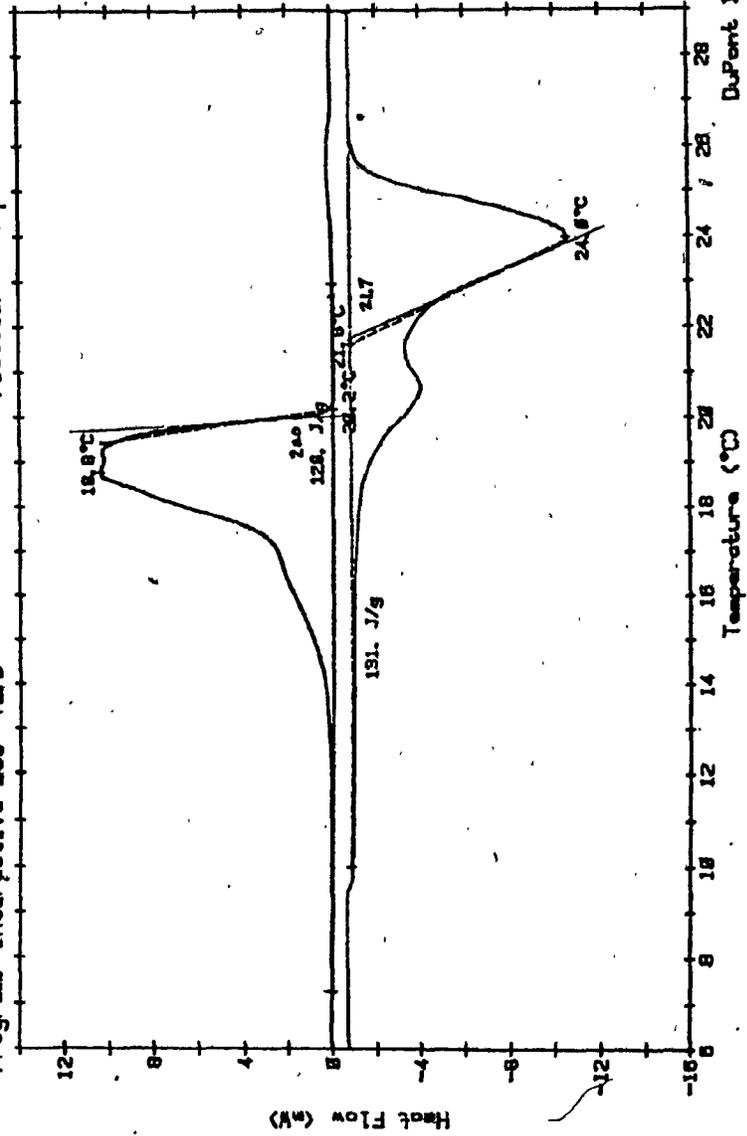


Fig. T-26. Thermogram of 55 mole % capric : 45 mole % lauric.

Date: 28-May-69 Time: 8:28  
File: DORINA,11 DISK EIGHT  
Operator:  
Plotted: 8-Sep-69 23:22:51

Sample: 60XCA+40XLA  
Size: 16.83MG  
Rate: 2C/MIN  
Program: Interactive DSC V2.8

### DSC

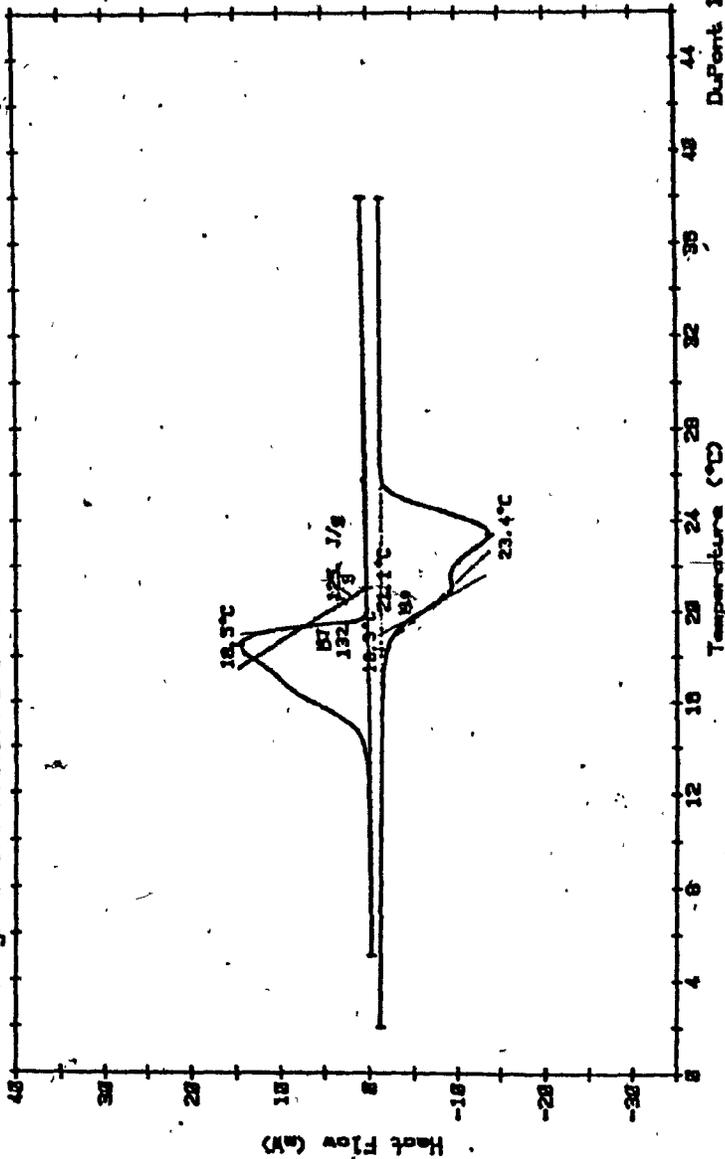


Fig. T-26. Thermogram of 60 mole % capric ; 40 mole % lauric. DuPont 1000

Sample 85C/35L  
Size 11.788 Mg  
Rate 2 c/MIN  
Program Interactive DSC V2.8

# DSC

Date 8-Jun-83 Time 13:53:08  
File FLKS.14  
Operator  
Plotted 8-Sep-83 23:37:48

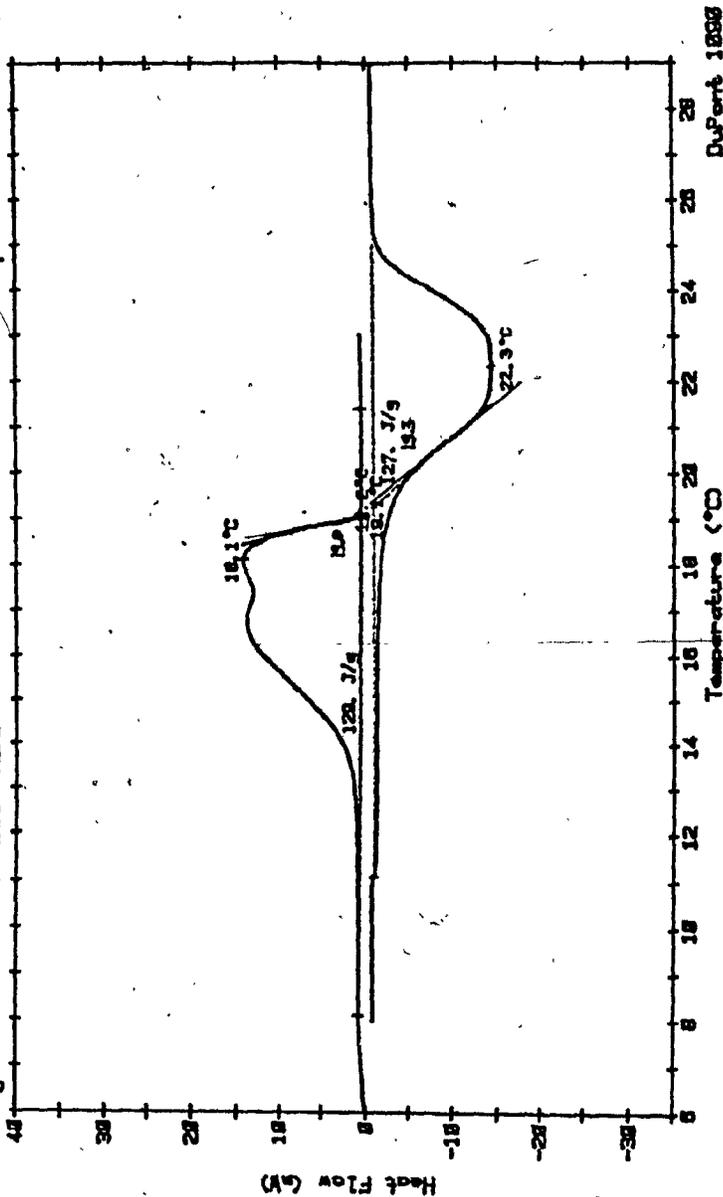


Fig. T-28. Thermogram of 65 mole % capric : 35 mole % lauric.

Sampler 782CA+382LA  
Size 0.73 Mc  
Rate 2 C/MIN  
Program Interactive DSC V2.8

**DSC**

Date 5-May-83 Time 12:39:11  
File DORINA.85 DISK SEVEN  
Operator  
Plotted 18-Jul-83 14:58:02

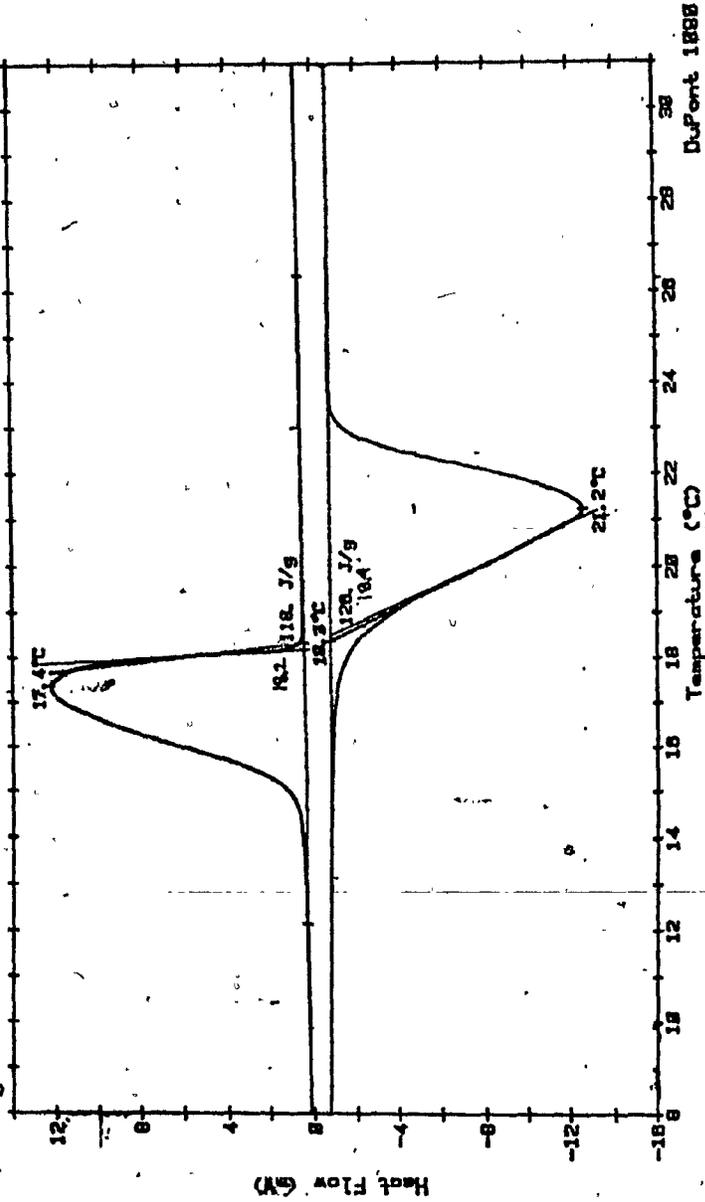


Fig. T-29. Thermogram of 70 mole % capric : 30 mole % lauric.

Samples 71XCA+28XLA  
Size 21.18MG  
Rate 2C/MIN  
Program Interactive DSC V2.8

**DSC**

Dates 9-May-83 Times 14:15:58  
Files DORINA.84 DORINA.DISK SIX  
Operator  
Plotted 18-Jul-83 15:06:42

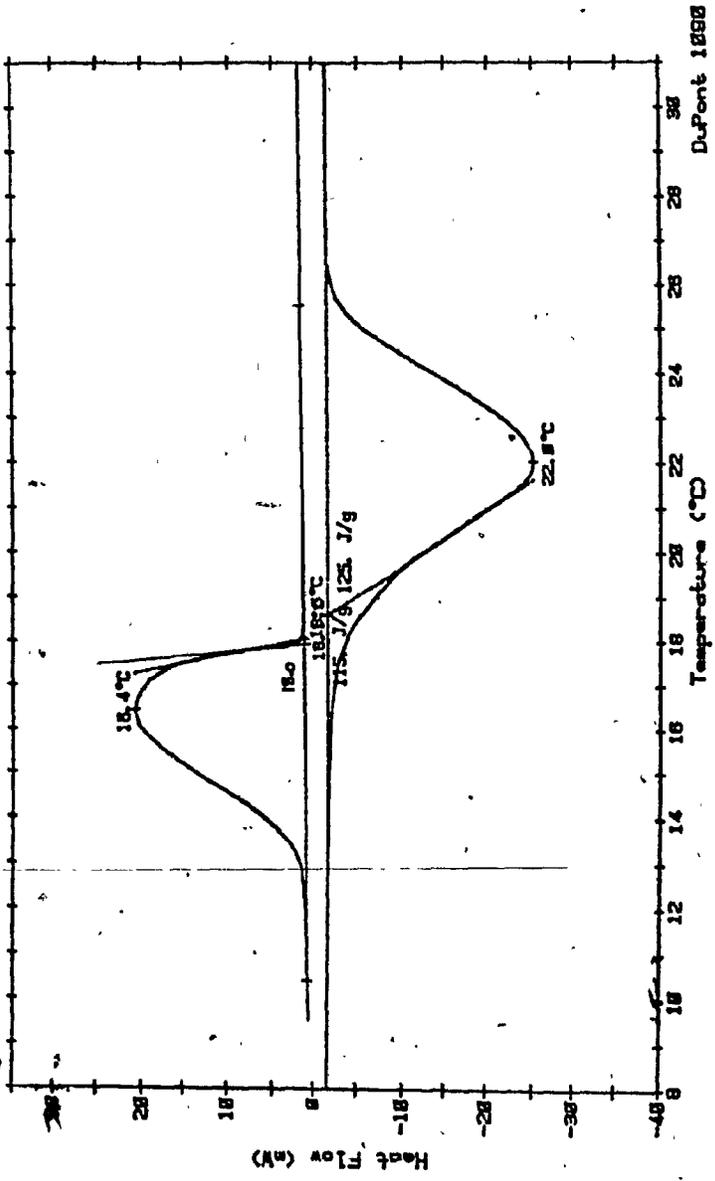


Fig. T-30. Thermogram of 71 mole % capric : 29 mole % lauric.

Sampler 723CA+28XLA  
Size 18.48MG  
Rate 2MG/MIN  
Program Interactive DSC V2.8

# DSC

Date: 8-May-83 Times 15:03:18  
File: DORINA.13 DORINA.DISK SIX  
Operator:  
Plotted: 18-Jul-83 15:22:58

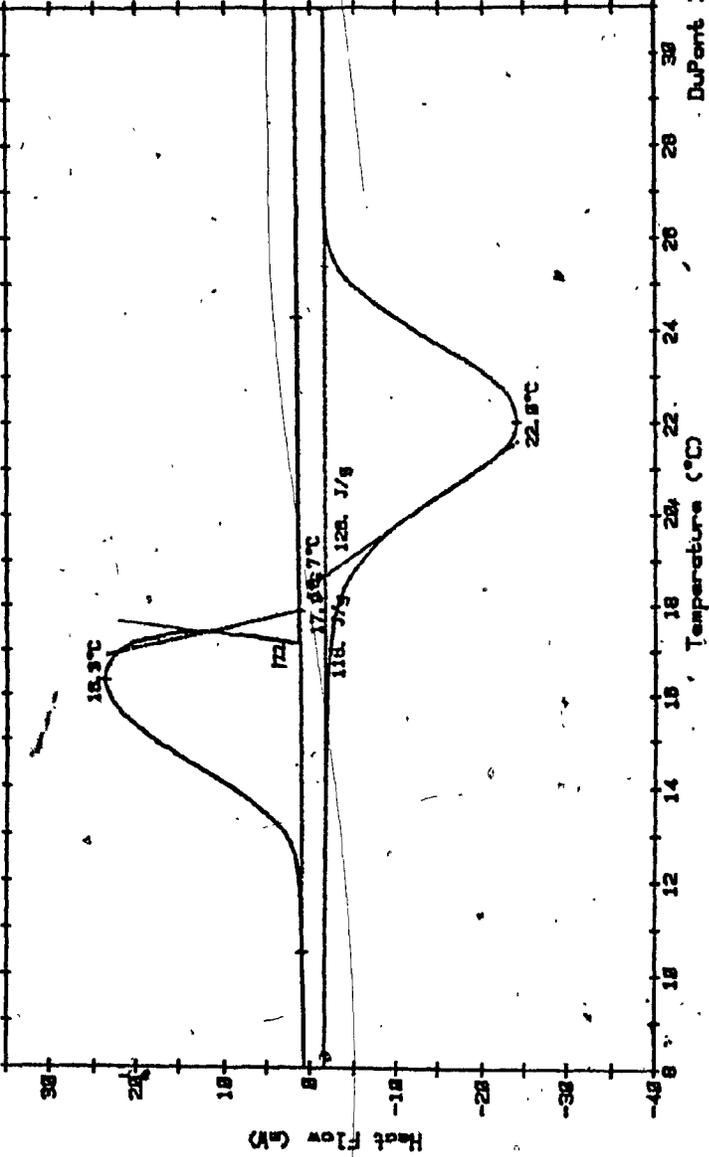


Fig. T-31. Thermogram of 72 mole % capric ; 28 mole % lauric.

Date: 5-May-83 Time: 11:55:25  
File: DORINA\_04 DISK SEVEN  
Operator:  
Plotted: 18-Jul-83 15:48:37

Sample: 72.5ICA+27.5LA  
Size: 18.48MG  
Rate: 2C/MIN  
Program: Interactive DSC V2.0

### DSC

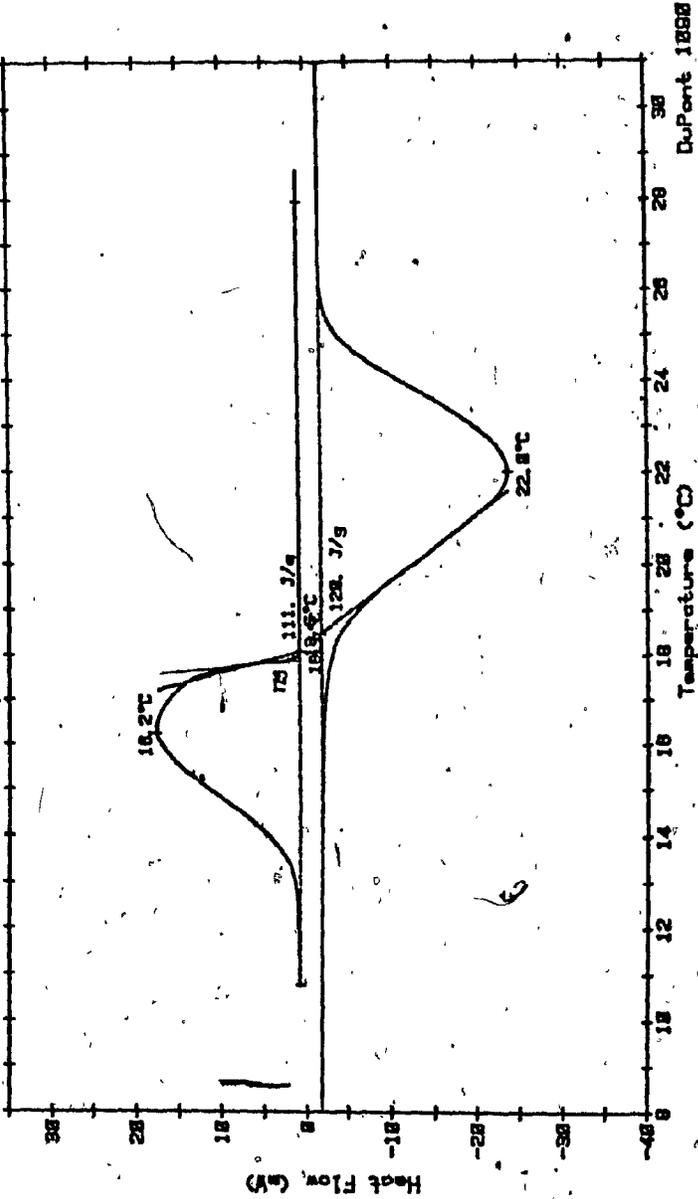


Fig. T-32. Thermogram of 72.5 mole % capric ; 27.5 mole % lauric.

3

Samples 73CA+27LA  
Size 5.75MG  
Rate 2C/MIN  
Programs Interactive DSC V2.8

DSC

Date 11-May-83 Times 18:18:22  
File DORINA.83 DORINA.DISK SIX  
Operator  
Plotted 18-Jul-83 18:04:48

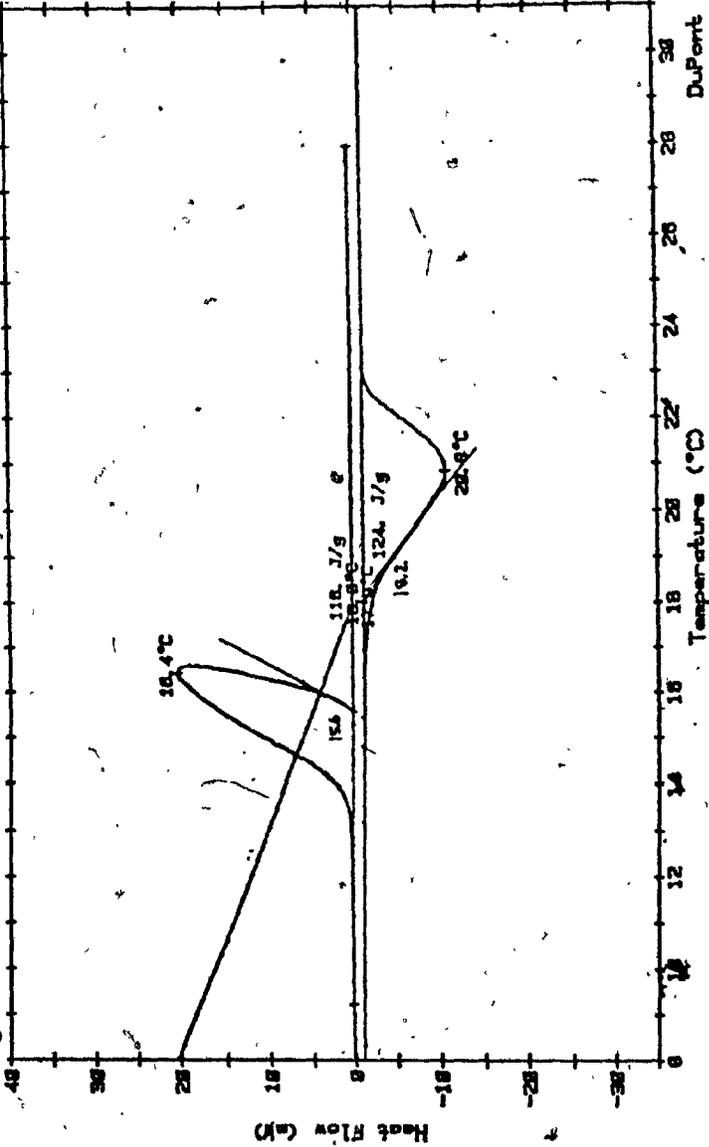


Fig. T-33. Thermogram of 27 mole % capric ; 27 mole % lauric.

Sample: 74C/26L  
Size: 5.745 MG  
Rate: 2 C/MIN  
Program: Interactive DSC V2.8

Date: 15-Jul-83 Time: 12:37:42  
File: FUKS.29 DISC FOUR  
Operator: Pfottach 18-Sep-83 22:32:48

### DSC

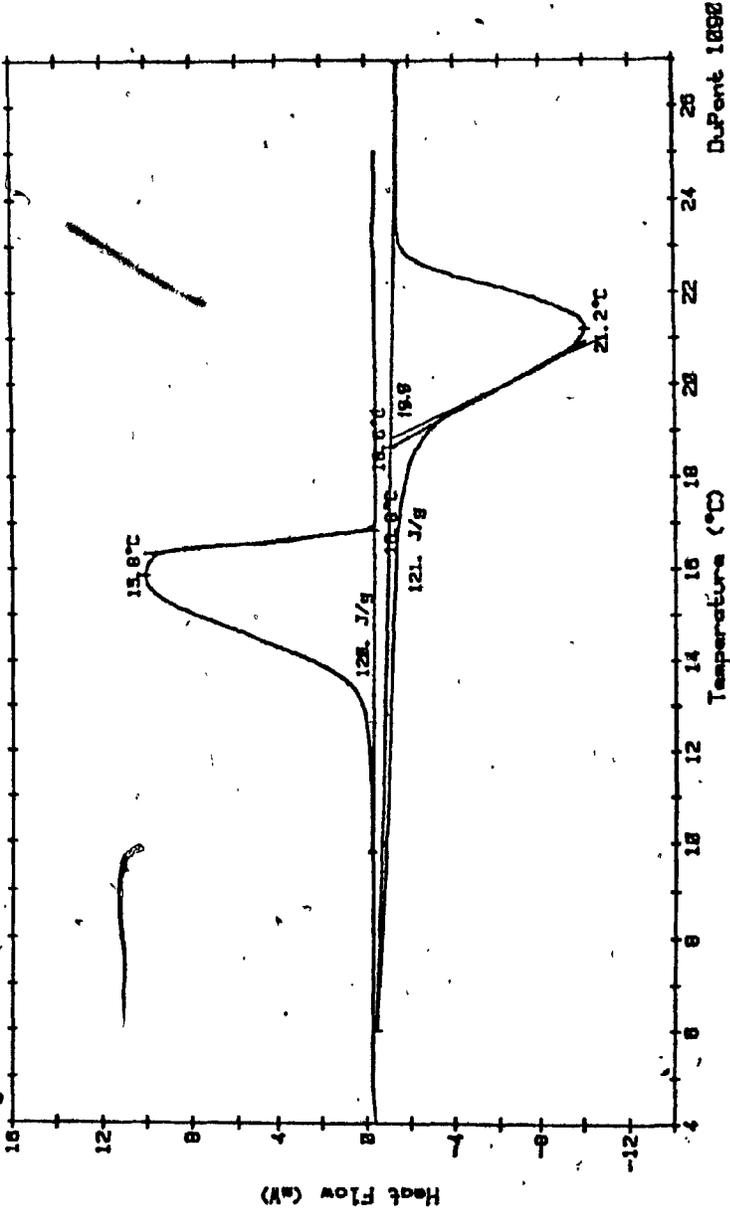


Fig. T-34. Thermogram of 74 mole % capric : 26 mole % lauric.

Dates 11-May-83 Times 12:41:05  
Files DORINA.08 DORINA.DISK SIX  
Operator  
Plotted 18-Jul-83 18:54:53

Sample 75CA+25LA  
Size 9.78MG  
Rate 2C/MIN  
Program Interactive DSC V2.0

### DSC

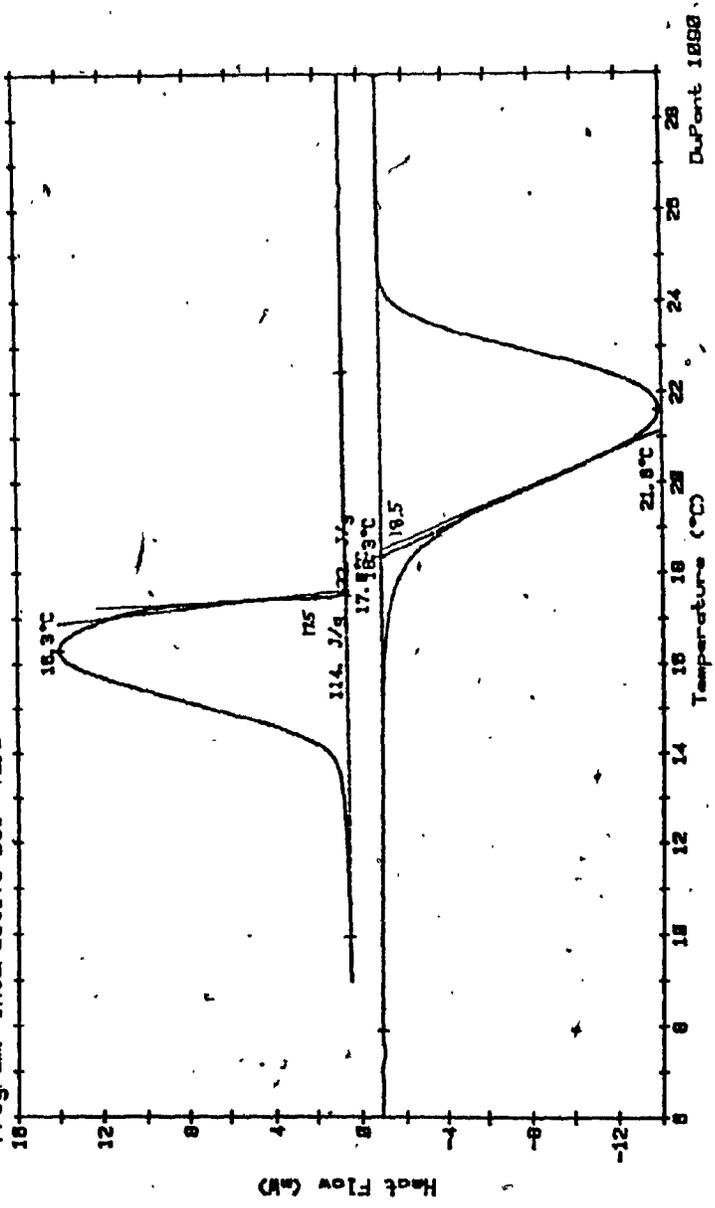


Fig. T-35. Thermogram of 75 mole % capric & 25 mole % lauric.

Sample: 88C/28L  
Size: 5.888 MG  
Rate: 2C/MIN  
Program: Interactive DSC V2.8  
Date: 1-Jun-83 Time: 11:44:24  
File: FLKS.83  
Operator:  
Plotted: 11-Sep-83 18:11:17

### DSC

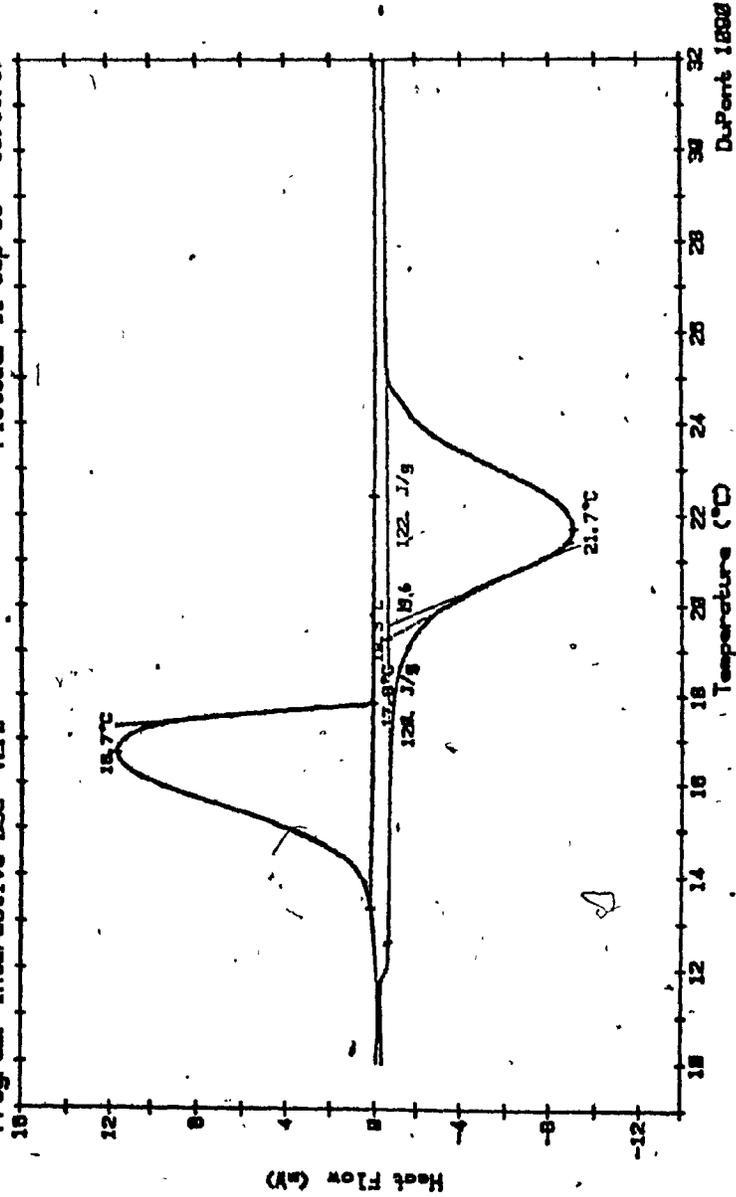


Fig. T-36. Thermogram of 80 mole % capric ; 20 mole % lauric.

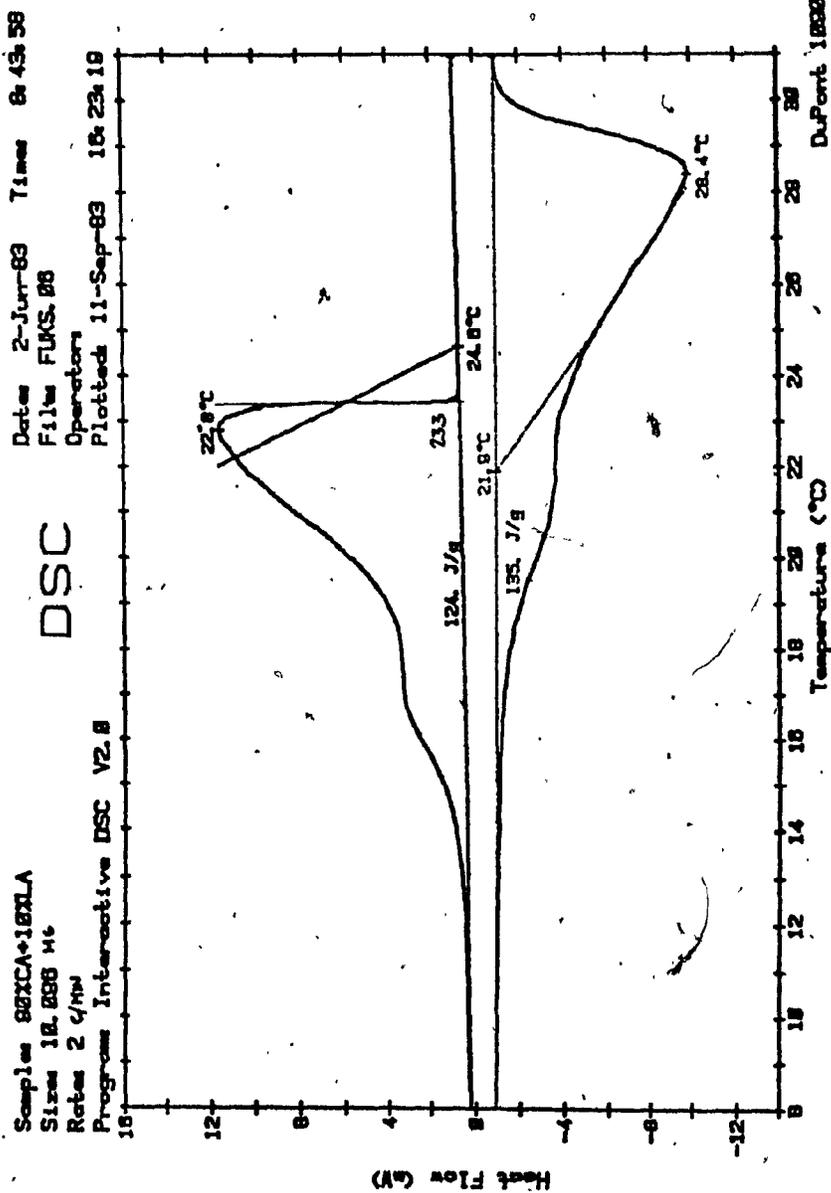


Fig. T-37. Thermogram of 90 mole % capric : 10 mole % lauric.

BINARY MIXTURES OF PALMITIC-STEARIC ACIDS

Sample 10P/90S  
Size 14.878MG  
Rate 2C/MIN  
Program Interactive DSC Y2.8

### DSC

Date 11-Jul-83 Time 14:38:18  
File FUKS:18 DISC FOUR  
Operator  
Plotted 5-Sep-83 15:28:19

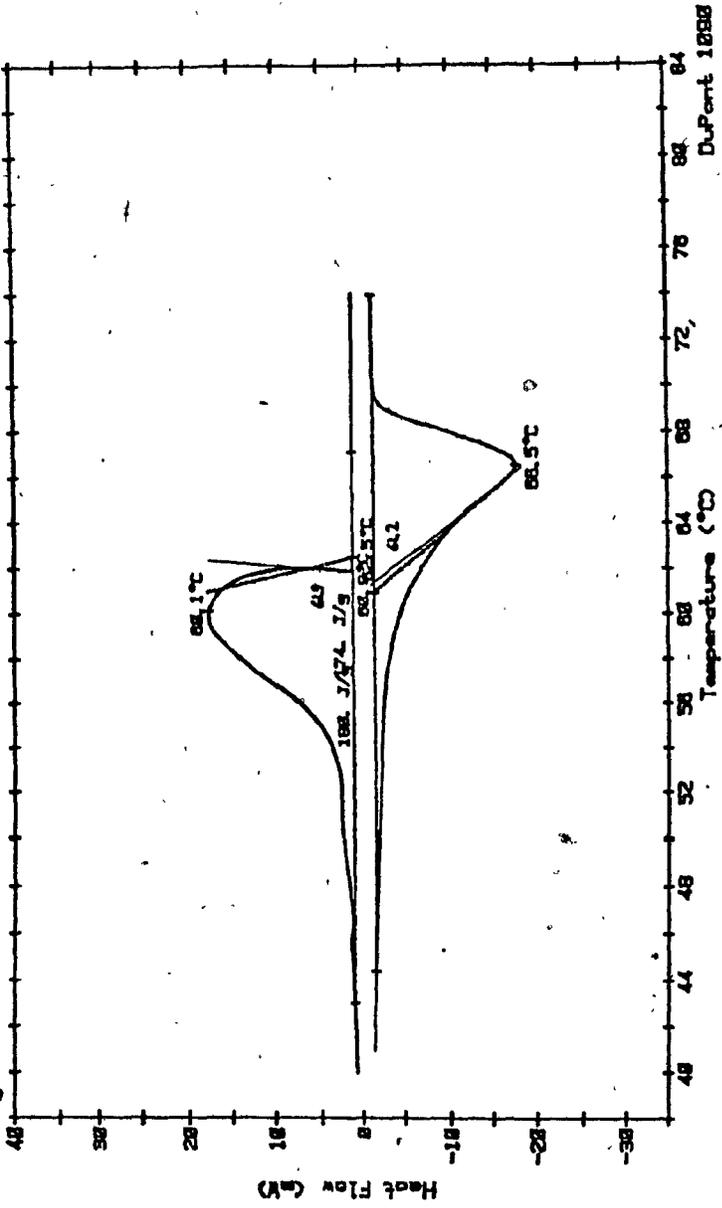


Fig. T-38. Thermogram of 10 mole % palmitic : 90 mole % stearic.

Dates 11-Jul-83 Times 13:31:25  
Files FUKS.15 DISC FOUR  
Operator  
Plotted 5-Sep-83 15:18:31

# DSC

Sample 20P/80S  
Size 8.552 MG  
Rate 2C/MIN  
Program Interactive DSC V2.8

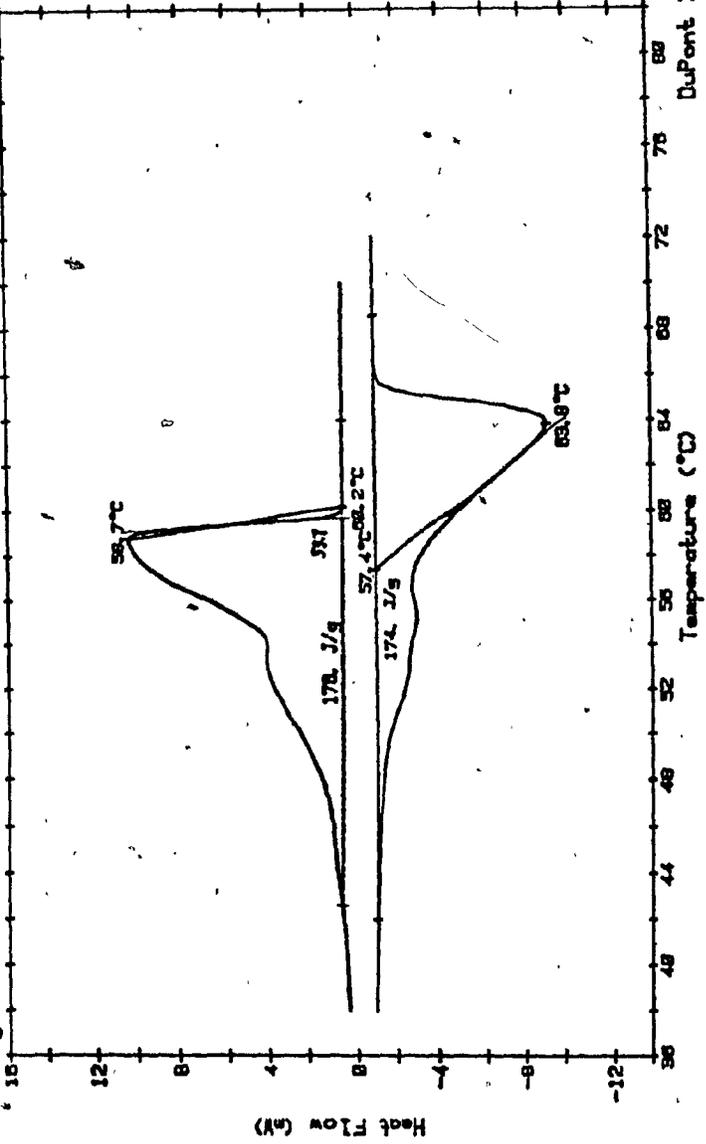


Fig. T-39. Thermogram of 20 mole % palmitic : 80 mole % stearic.

Samples 30P/70S  
Size: 9.974 MG  
Rate: 2C/MIN  
Program Interactive DSC V2.8  
Date: 11-Jul-83 Time: 12:35:25  
File: FUKS.14 DISC FOUR  
Operator  
Plotted: 11-Jul-83 15:54:14

### DSC

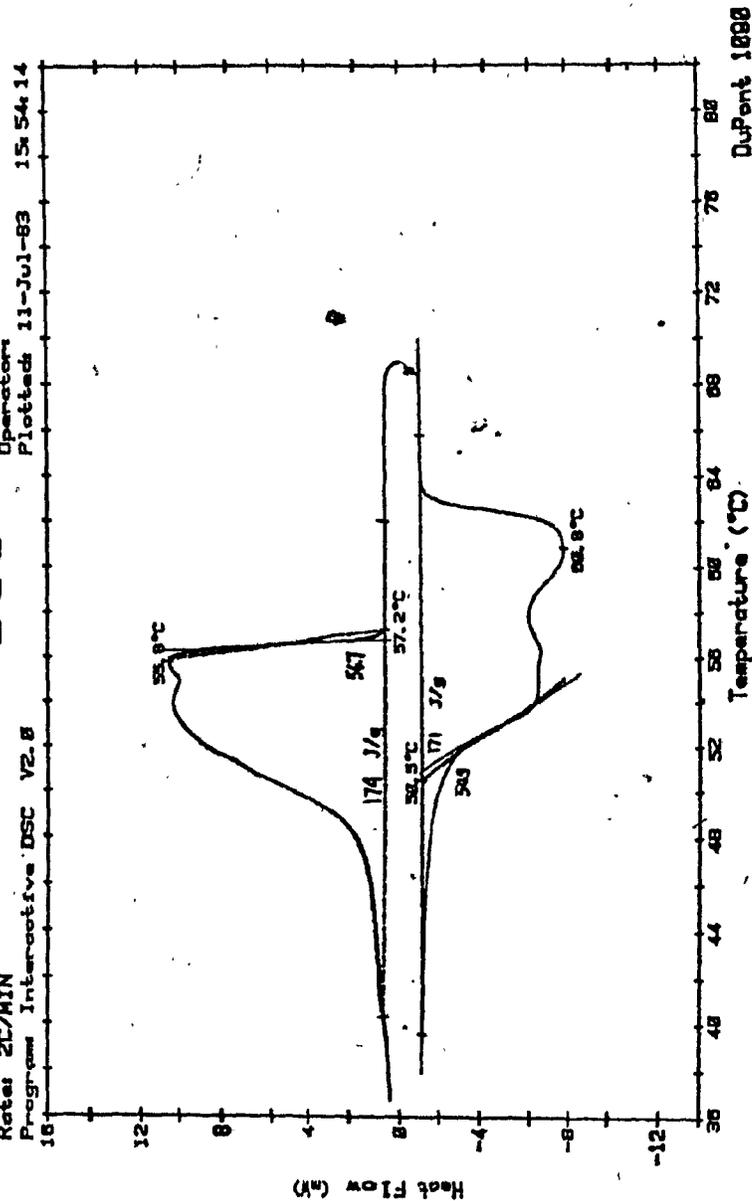


Fig. T-40. Thermogram of 30 mole % palmitic : 70 mole % stearic.

Sample: 4BP/50S  
Size: 7.239 MG  
Rate: 2C/MIN  
Program: Interactive DSC V2.8

### DSC

Date: 11-Jul-83 Time: 11:43:20  
File: FMS.13 DISC FOUR  
Operator:  
Plotted: 11-Jul-83 16:07:59

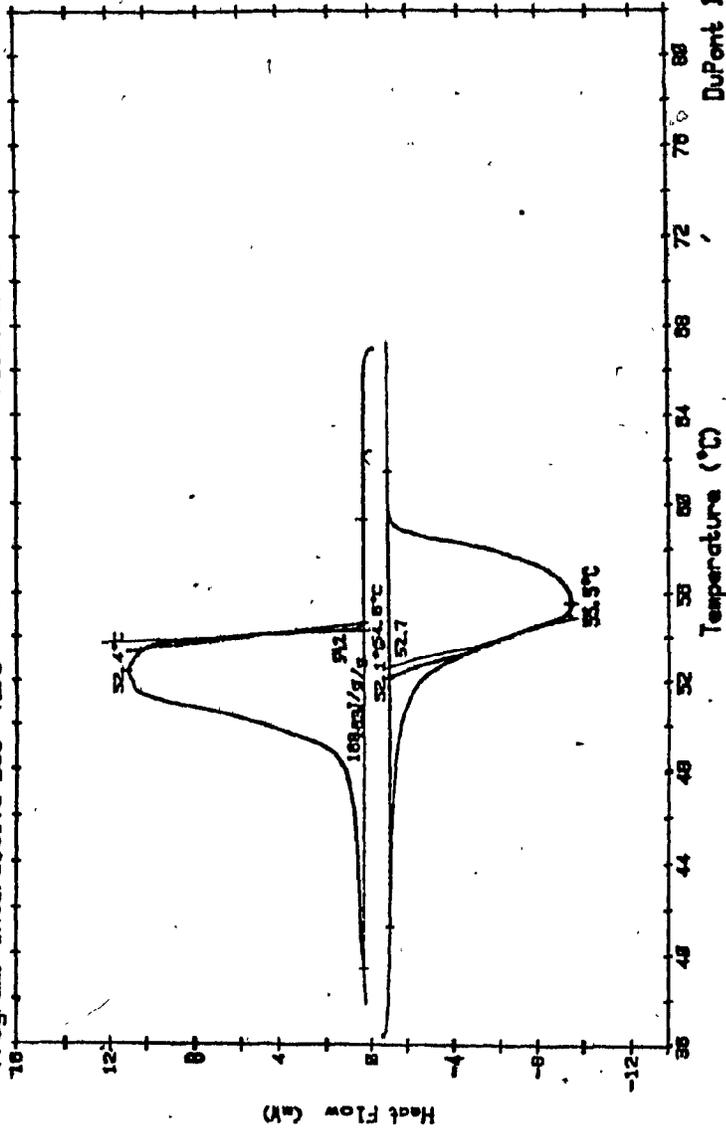


Fig. T-41. Thermogram of 40 mole % palmitic ; 60 mole % stearic.

Date: 11-Jul-63 Time: 18:41:48  
File: FUMS.12 PISC FOUR  
Operator:  
Plotted: 5-Sep-63 15:53:41

# DSC

Sample: 5BP/5BS  
Size: 14.848MG  
Rate: 2C/MIN  
Program: Interactive DSC V2.8

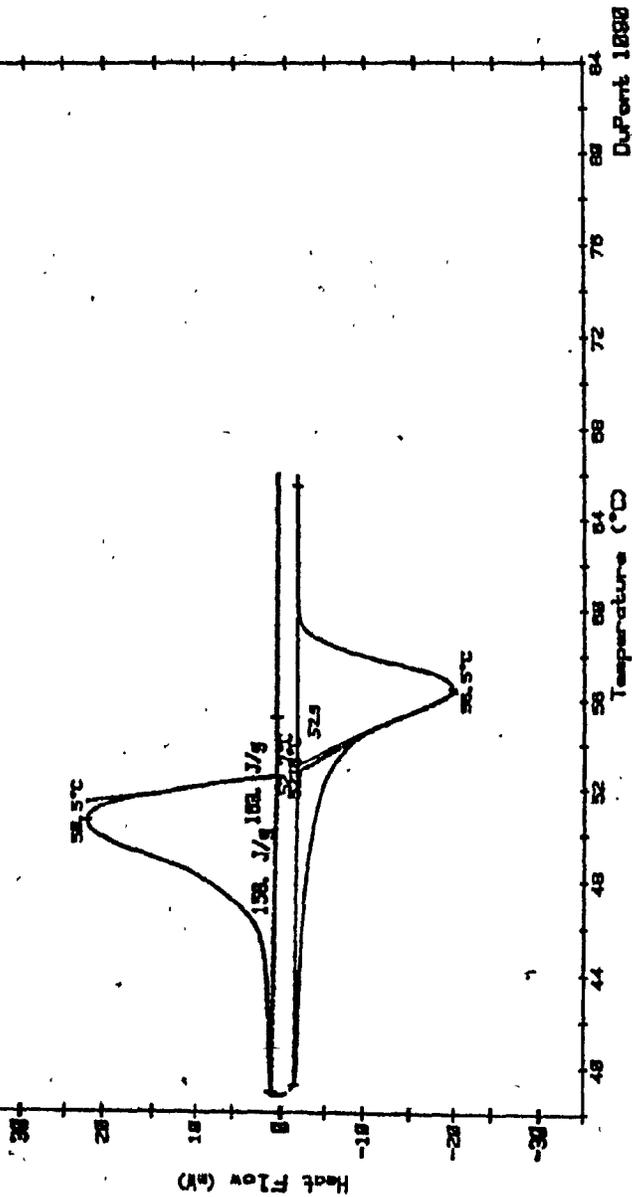


Fig. T-42. Thermogram of 50 mole % palmitic : 50 mole % stearic.

Date: 11-Jul-83 Time: 9:37:38  
File: FUKS.11 DISC FOUR  
Operator:  
Plottack: 5-Sep-83 14:32:18

Sampler: 80P/40S  
Size: 11.583MG  
Rate: 2C/MIN  
Program: Interactive DSC V2.B

### DSC

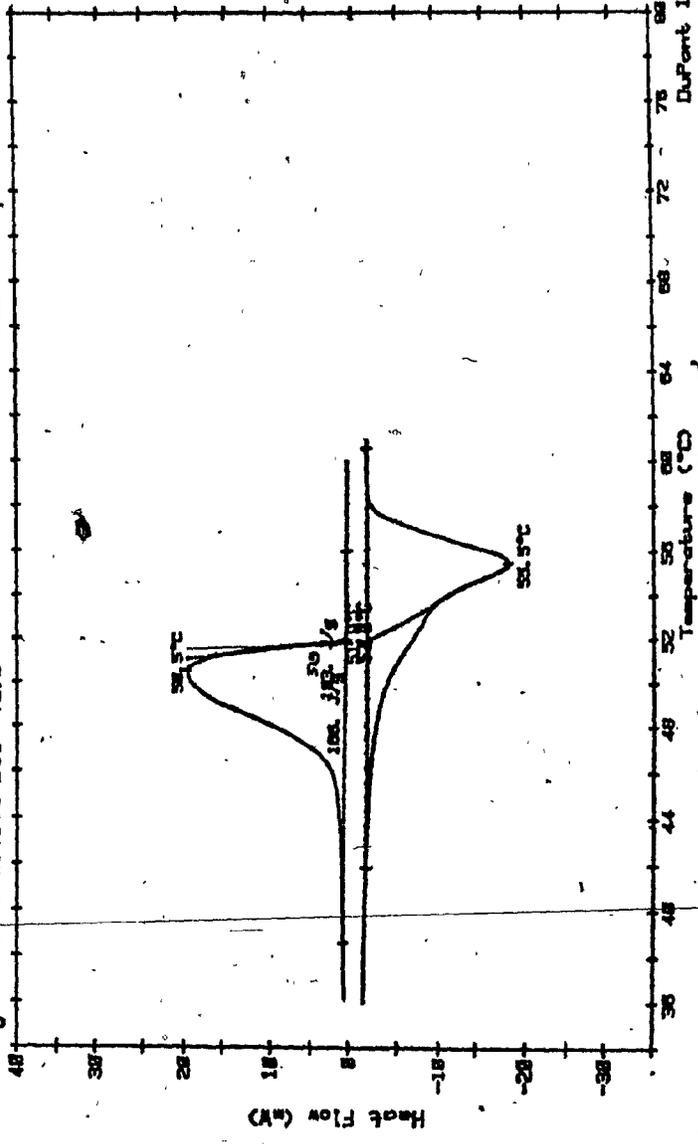


Fig. T-43. Thermogram of 60 mole % palmitic ; 40 mole % stearic.

Sample: 88P/319  
Size: 3.357 MG  
Rate: 2 C/MIN  
Program: Interactive DSC V2.8

# DSC

Date: 11-Jul-83 Time: 20:01:43  
File: FUKS.17 DISC FOUR  
Operator:  
Plotted: 2-Aug-83 23:01:54

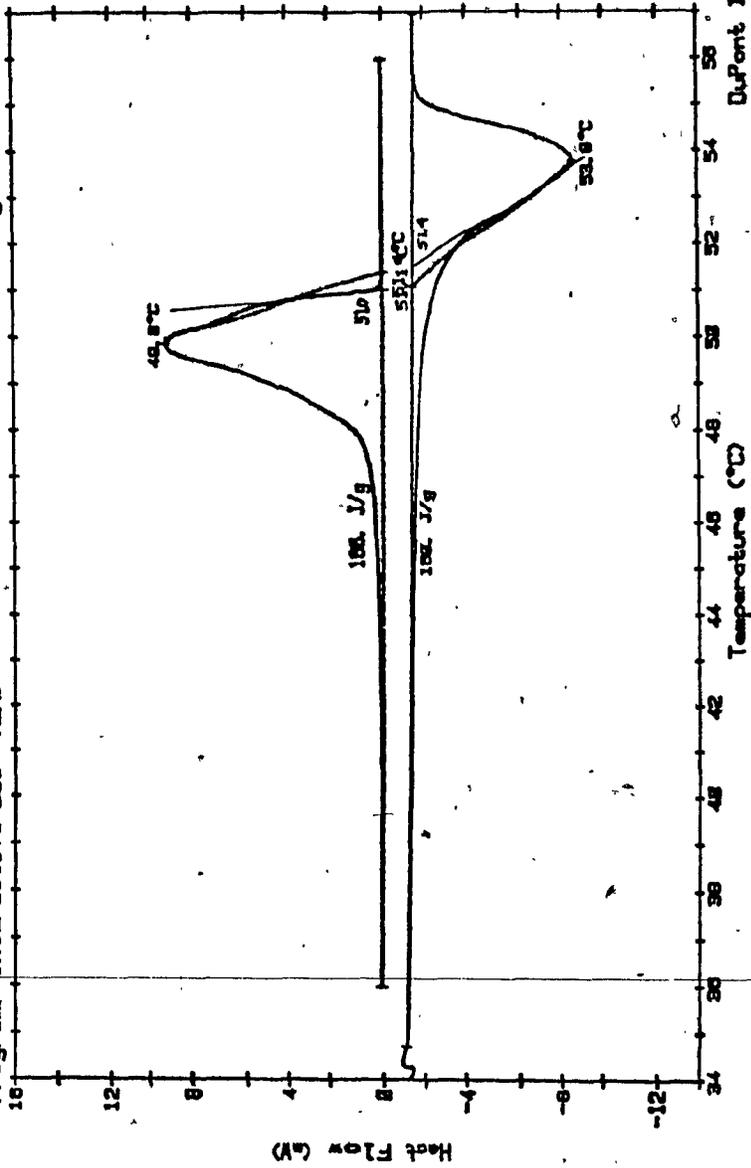


Fig. T-44. Thermogram of 69 mole % palmitic : 31 mole % stearic.

Sample: 70P/30S  
Size: 12.461 MG  
Rate: 2 C/MIN  
Program: Interactive DSC V2.0

# DSC

Date: 18-Jul-83 Time: 17:11:43  
File: FUKS.10 DISC FOUR  
Operator:  
Plotted: 5-Sep-83 14:02:19

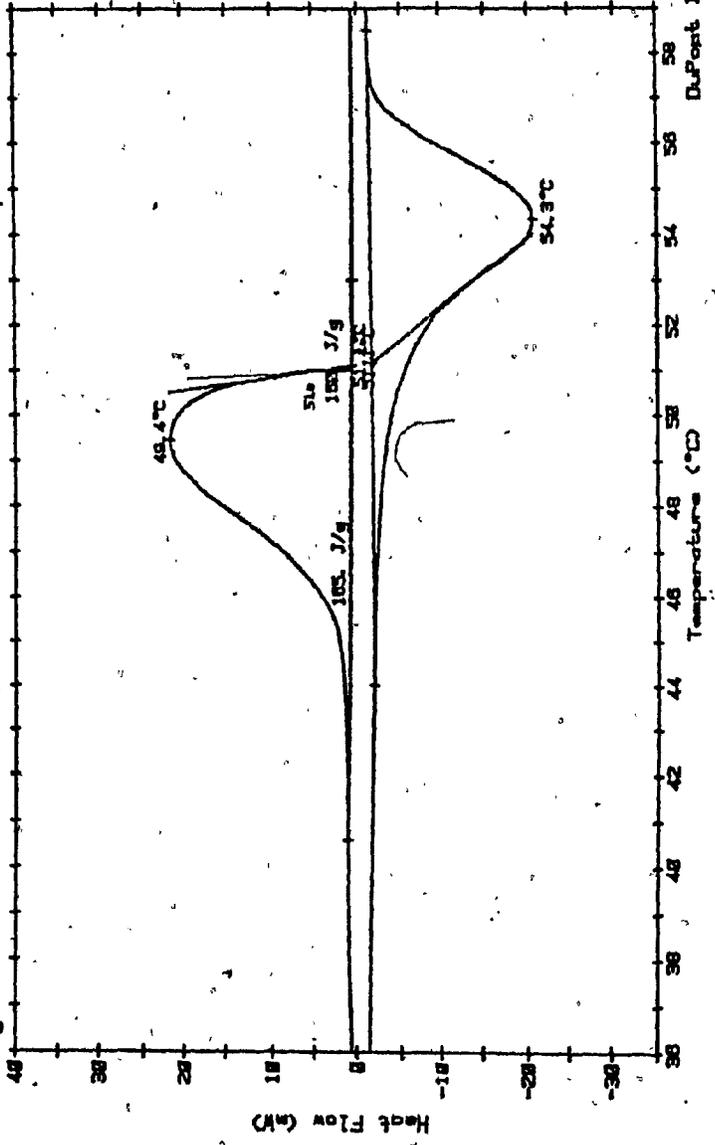


Fig. T-45. Thermogram of 70 mole % palmitic ; 30 mole % stearic.

Sample: 71P/29S  
Size: 0.218 MG  
Rate: 2 C/MIN  
Program: Interactive DSC V2.8  
Date: 11-Jul-83 Time: 20:56:58  
File: FUKS.18 DISC FOUR  
Operator:  
Plotted: 5-Sep-83 13:44:51

### DSC

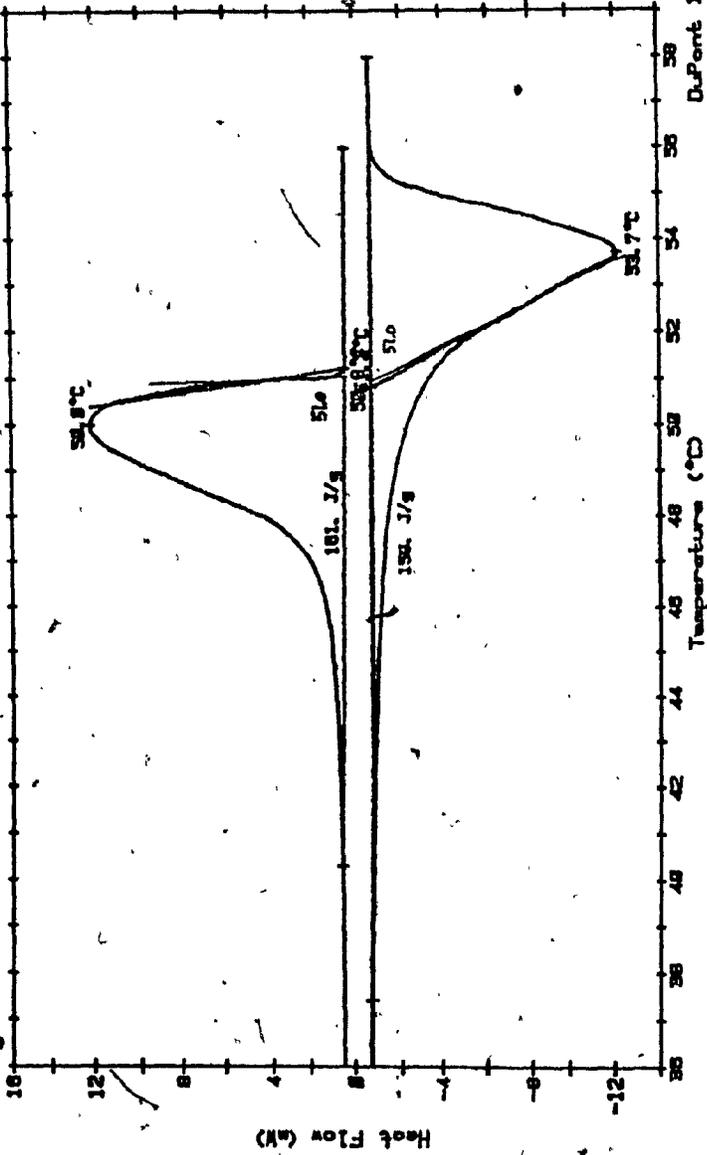


Fig. T-46. Thermogram of 71 mole % palmitic : 29 mole % stearic.

Sample 72.5P/27.5S  
Size 8.169 MG  
Rate 2 C/MIN  
Program Interactive DSC V2.0

DSC

Date 11-Jul-83 Time 2h 44:52  
File FUKS.19 DISC FOUR  
Operator  
Plotted 5-Sep-83 13:28:03

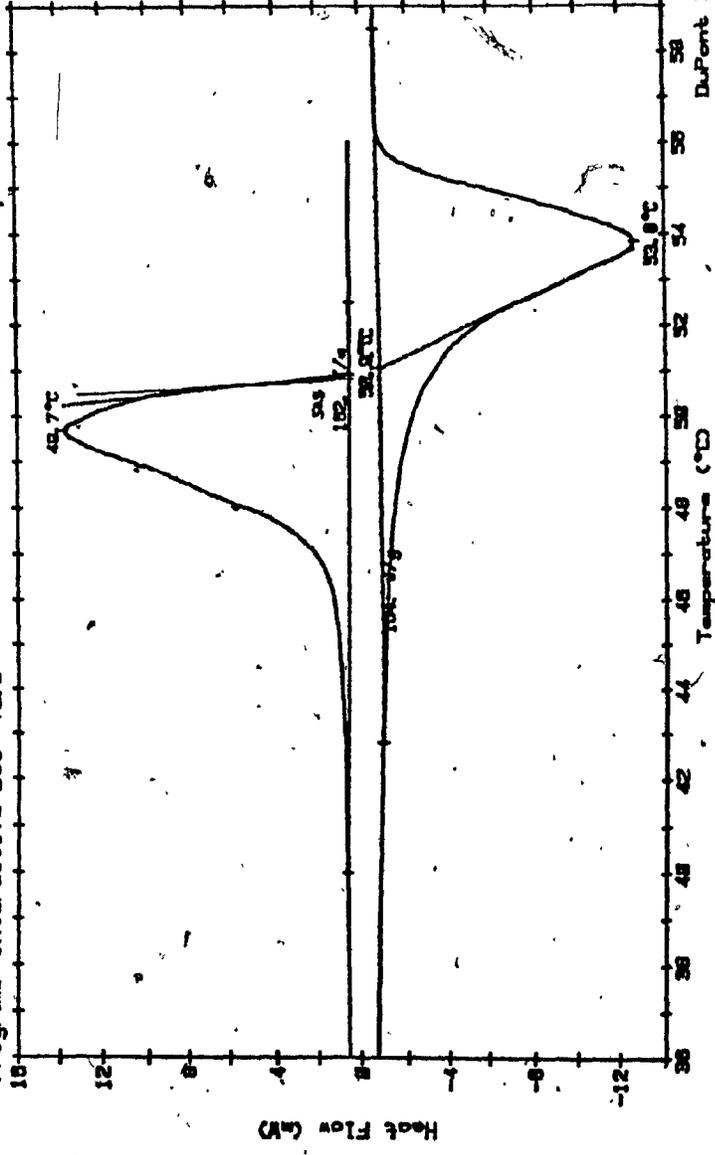


Fig. T-47. Thermogram of 72.5 mole % palmitic ; 27.5 mole % stearic.

Sample 74P/268  
Size 8.457 MG  
Rate 2 C/MIN  
Program Interactive DSC V2.0  
Date 11-Jul-83 Time 22:35:34  
Files FLKS.20 DISC FOUR  
Operator  
Plotted 5-Sep-83 15M3:21

### DSC

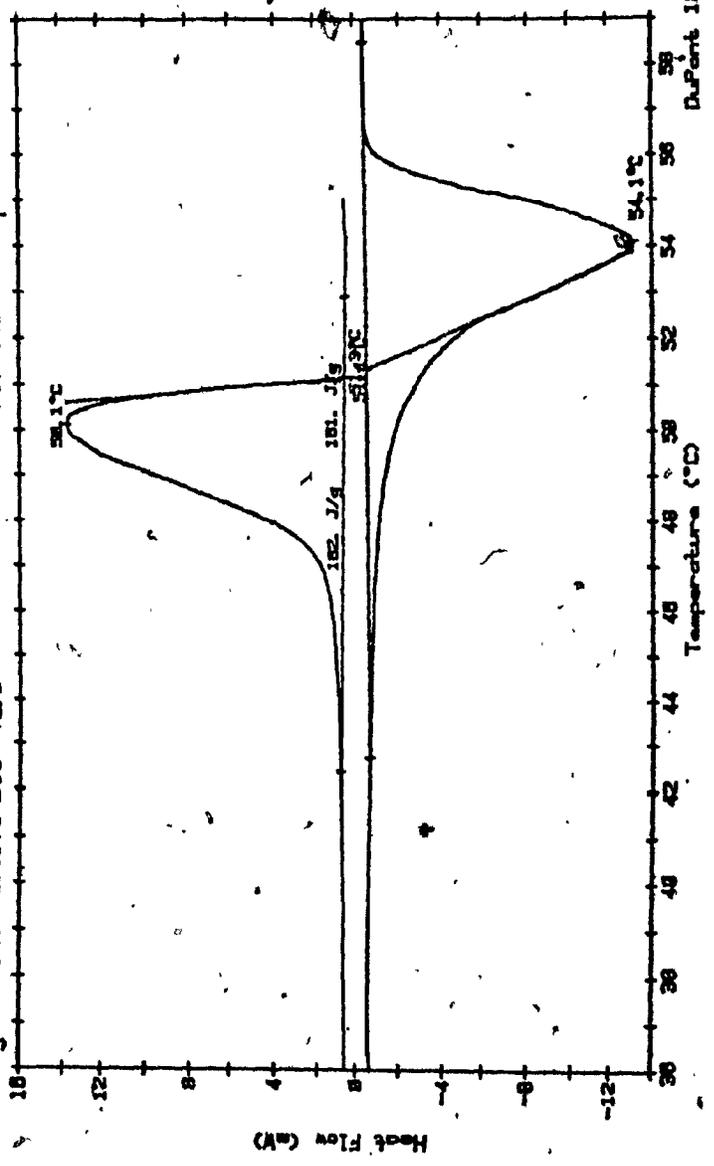


Fig. T-48. Thermogram of 74 mole % palmitic : 26 mole % stearic.

Sample 80P/20S  
Size 7.543 MG  
Rate 2 C/MIN  
Program Interactive DSC V2.8  
Date 18-Jul-83 Times 16:28:32  
Files FUKS.89 DISC FOUR  
Operator  
Plotted 18-Jul-83 17:09:18

### DSC

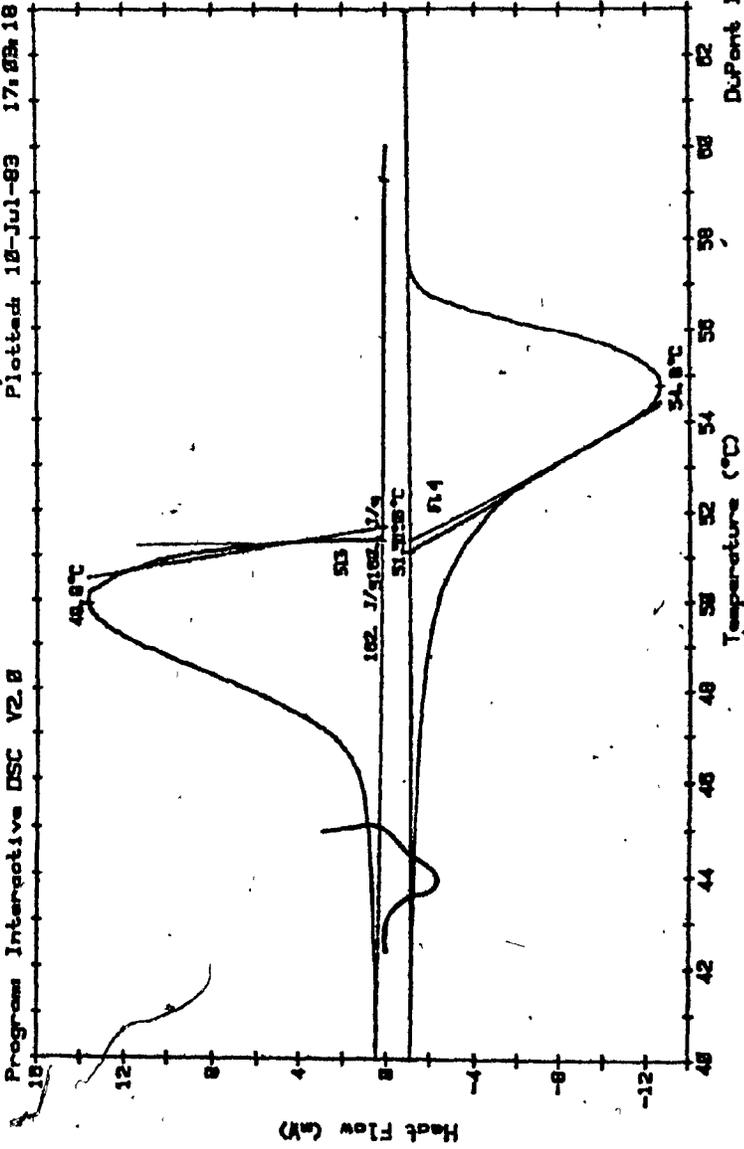


Fig. T-49. Thermogram of 80 mole % palmitic : 20 mole % stearic.

Date: 18-Jul-83 Time: 15:58:21  
File: FUKS.08 DISC FOUR  
Operator:  
Plotted: 5-Sep-83 13:05:25

Sample: 90P/10S  
Size: 4.496 MG  
Rate: 2 C/MIN  
Program: Interactive DSC V2.0

### DSC

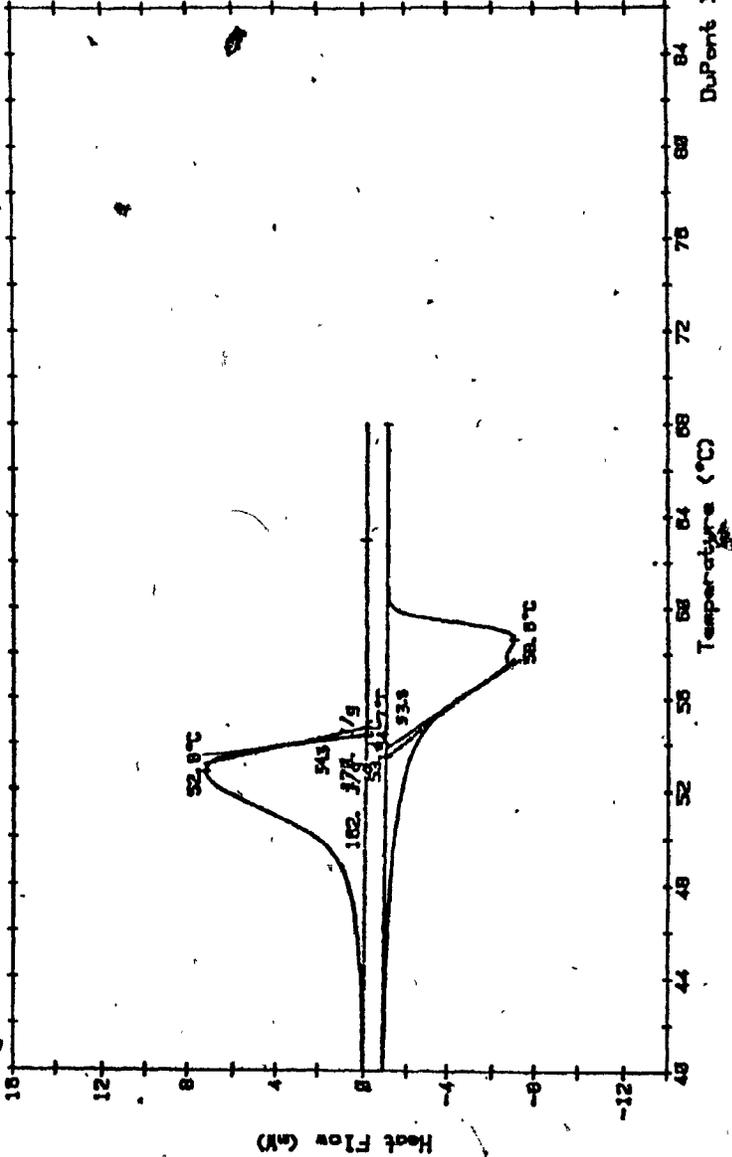


Fig. T-50. Thermogram of 90 mole % palmitic : 10 mole % stearic.

BINARY MIXTURES OF LAURIC-STEARIC ACIDS

Sample 802SA+187LA  
Size 8.88MG  
Rate 2 C/MIN  
Program Interactive DSC Y2.8

### DSC

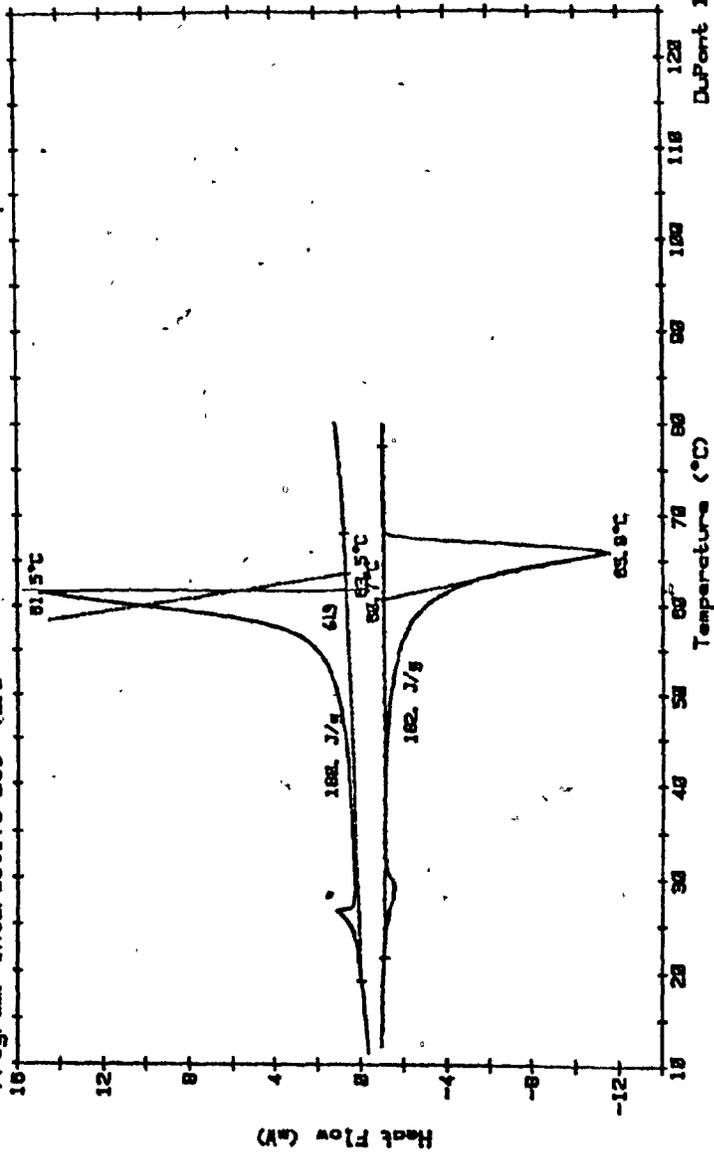


Fig. T-51. Thermogram of 10 mole % lauric : 90 mole % stearic.

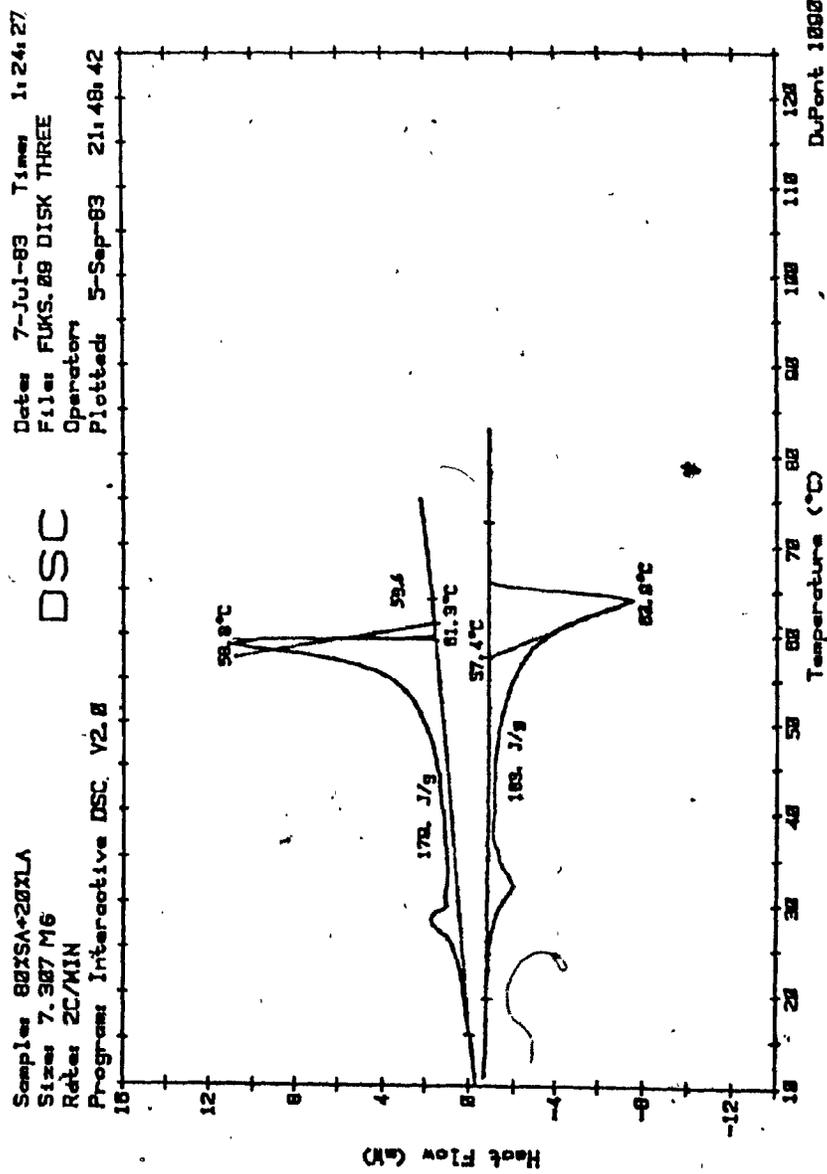


Fig. T-52. Thermogram of 20 mole % lauric : 80 mole % stearic.

Date: 8-Jul-83 Times: 9:17:38  
File: FUKS.10 DISK THREE  
Operator:  
Plotted: 8-Jul-83 10:48:39

Sample: 78XSA-30XLA  
Size: 8.557MG  
Rate: 2C/MIN  
Program: Interactive DSC V2.8

### DSC

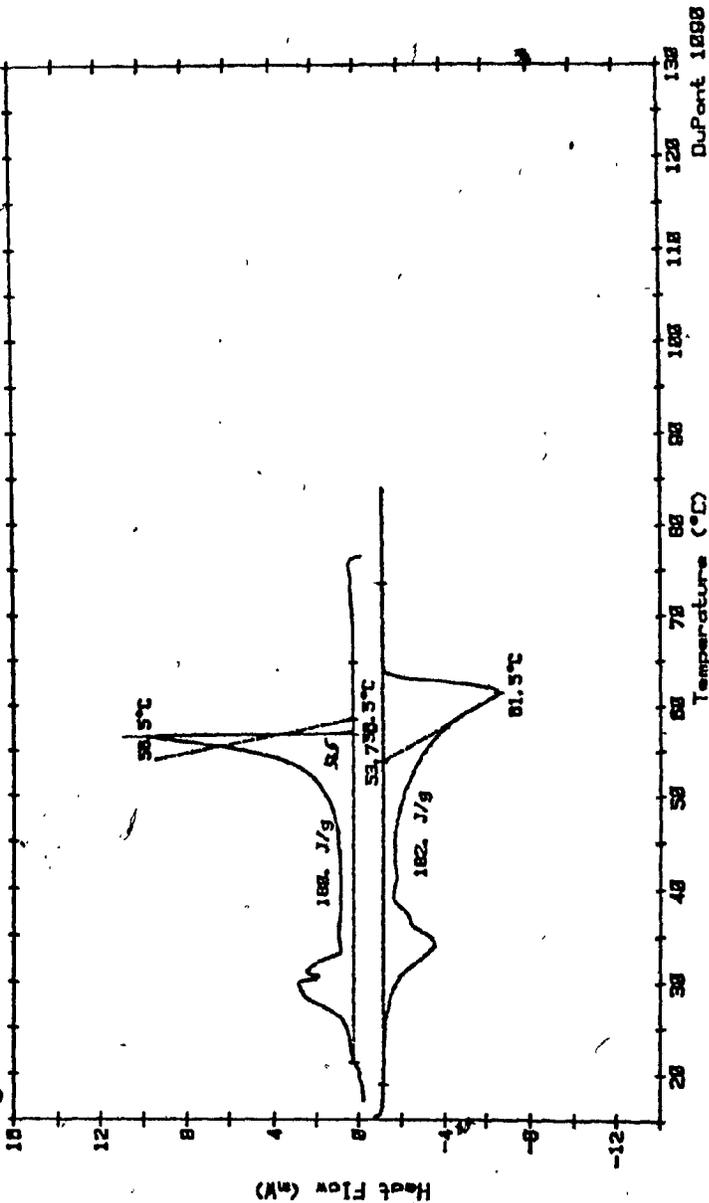


Fig. T-53. Thermogram of 30 mole % lauric : 70 mole % stearic.

Samples 8025A+482LA  
Size 10.823MG  
Rate 2C/MIN  
Program Interactive DSC V2.0

**DSC**

Dates 8-Jul-83 Times 14:22:25  
Files DORINA.19 DISK NINE  
Operator  
Plotted 2-Aug-83 21:21:18

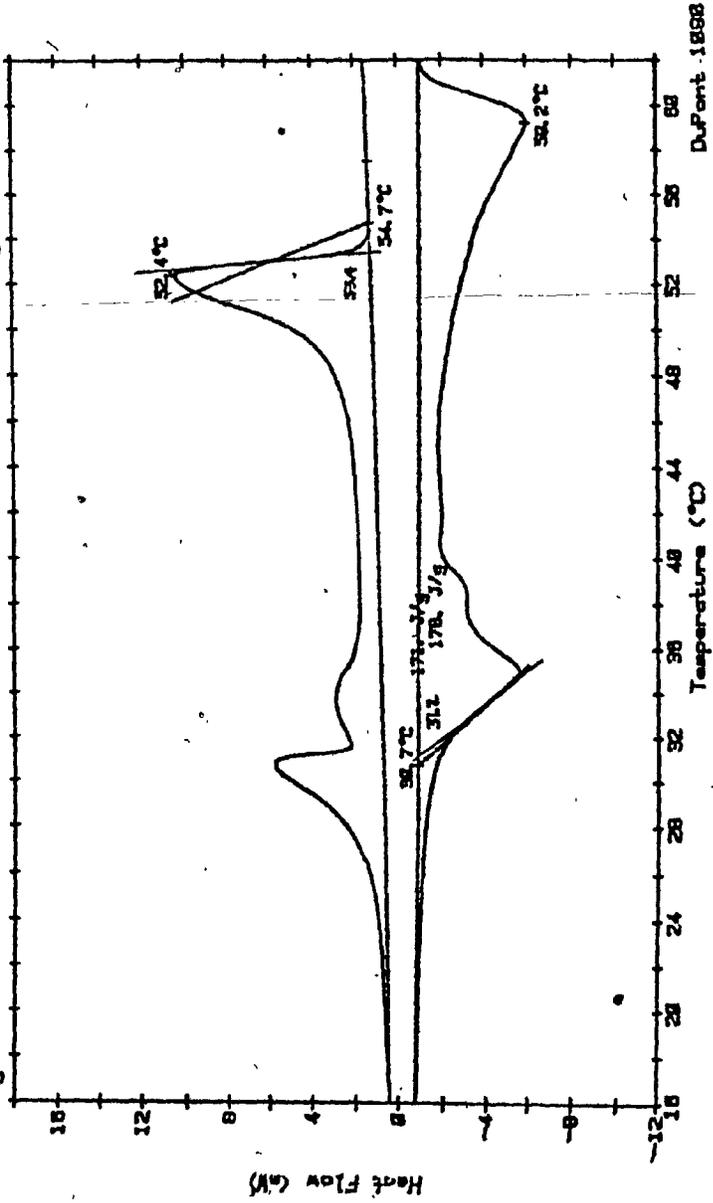


Fig. T-54. Thermogram of 40 mole % lauric : 60 mole % stearic.

Samples 50XSA+50ZLA  
Size 7.676 MG  
Rate 2C/MIN  
Program Interactive DSC V2.8

**DSC**

Date 8-Jul-83 Time 15:54:46  
File DORINA.28 DISK NINE  
Operator  
Plotted 8-Jul-83 19:38:43

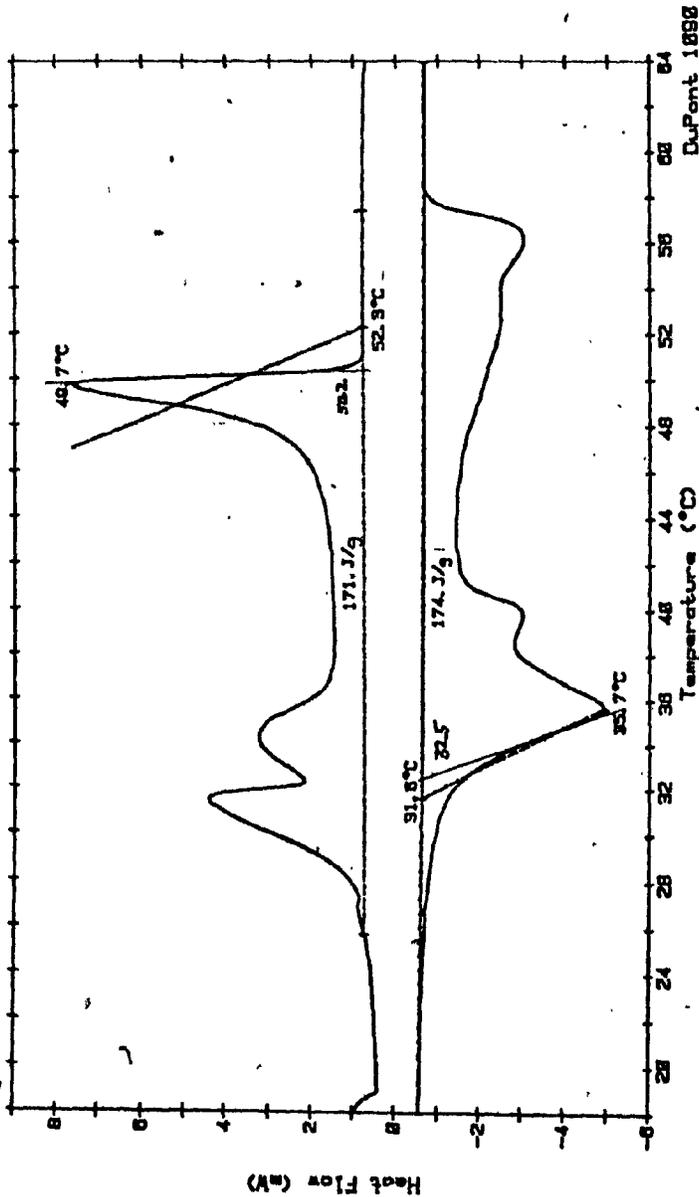


Fig. T-55. Thermogram of 50 mole % lauric / 50 mole % stearic. DuPont 1898

Sample: 80L/48S  
Size: 4.874 MG  
Rate: 2 C/MIN  
Program: Interactive DSC V2.8

# DSC

Date: 8-Jul-83 Time: 18:23:24  
File: FUKS.01 DISC FOUR  
Operator:  
Plotted: 8-Jul-83 19:27:48

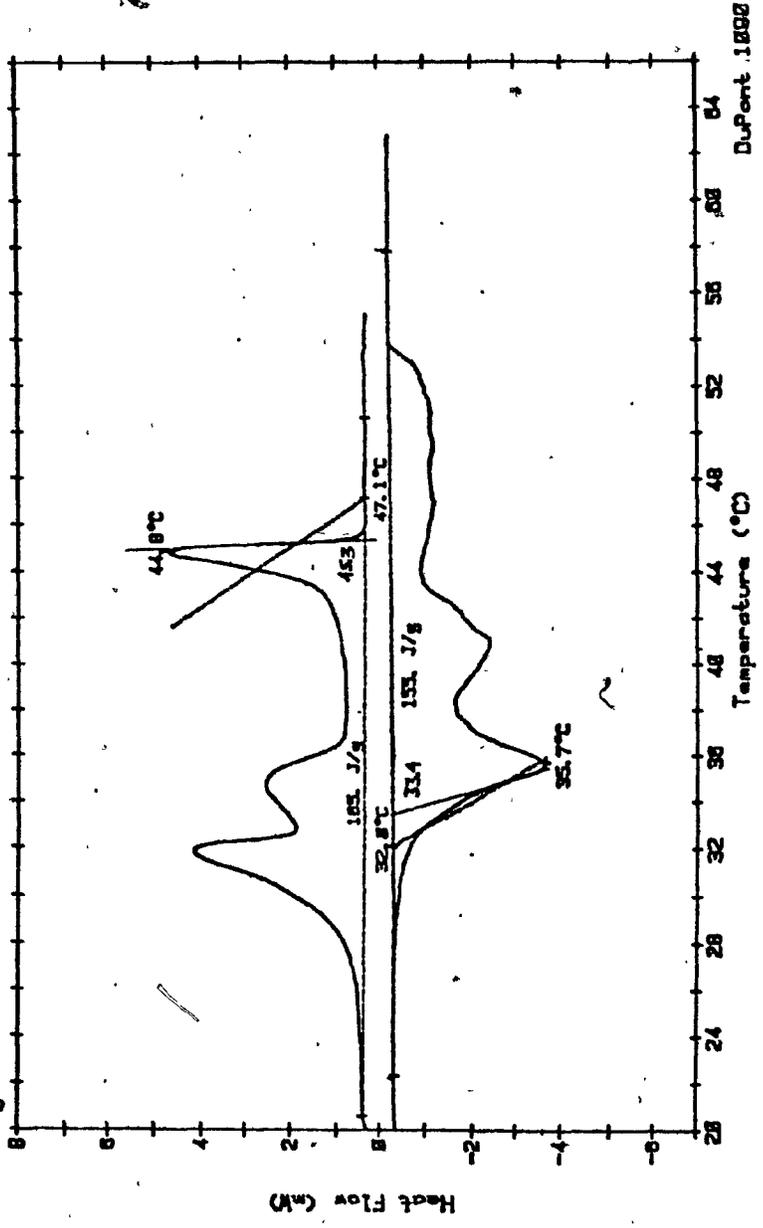


Fig. T-56. Thermogram of 60 mole % lauric : 40 mole % stearic.

Samples 70L/30S  
Size: 3.798 MG  
Rate: 2 C/MIN  
Program: Interactive DSC V2.8

# DSC

Date: 8-Jul-83 Time: 19:38:44  
File: FUKS.02 DISC.FOUR  
Operator:  
Plotted: 8-Jul-83 20:37:12

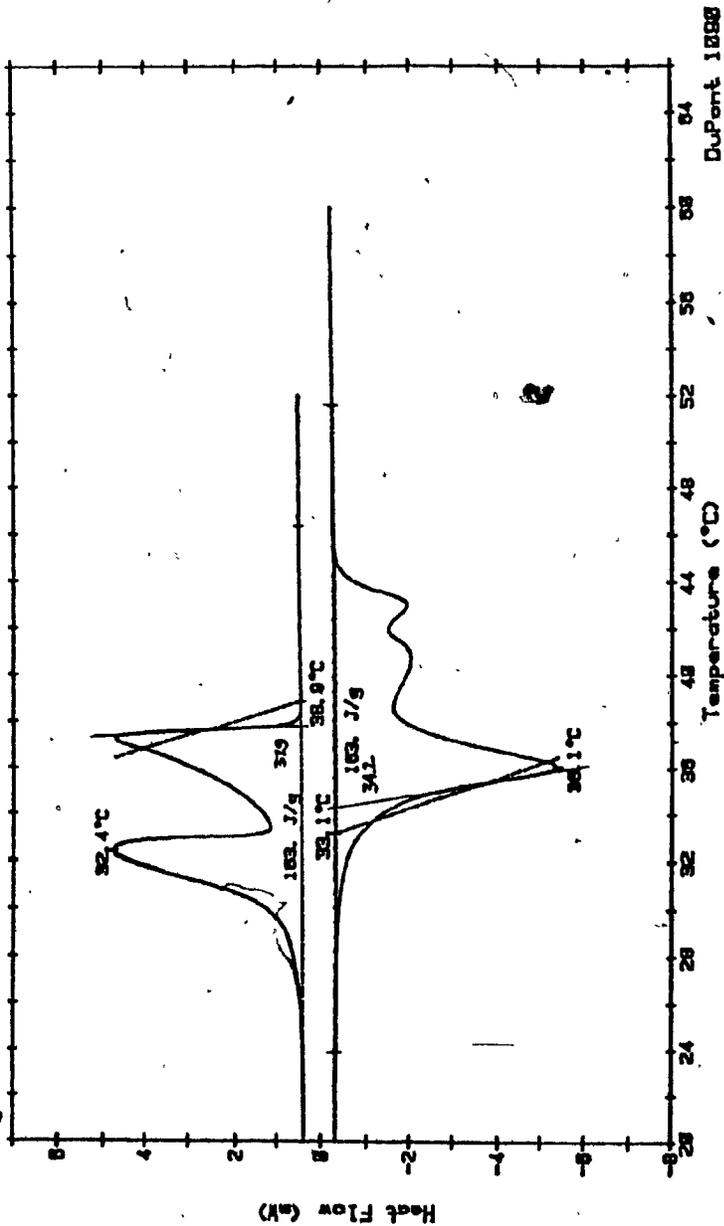


Fig. T-57. Thermogram of 70 mole % lauric : 30 mole % stearic.

Sample 88L/28S  
Size 3.4 MG  
Rate 2 C/MIN  
Program Interactive DSC V2.8

# DSC

Date 8-Jul-83 Times 20:48:03  
File FUKS.03 DISC FOUR  
Operator  
Plotted 8-Jul-83 21:26:16

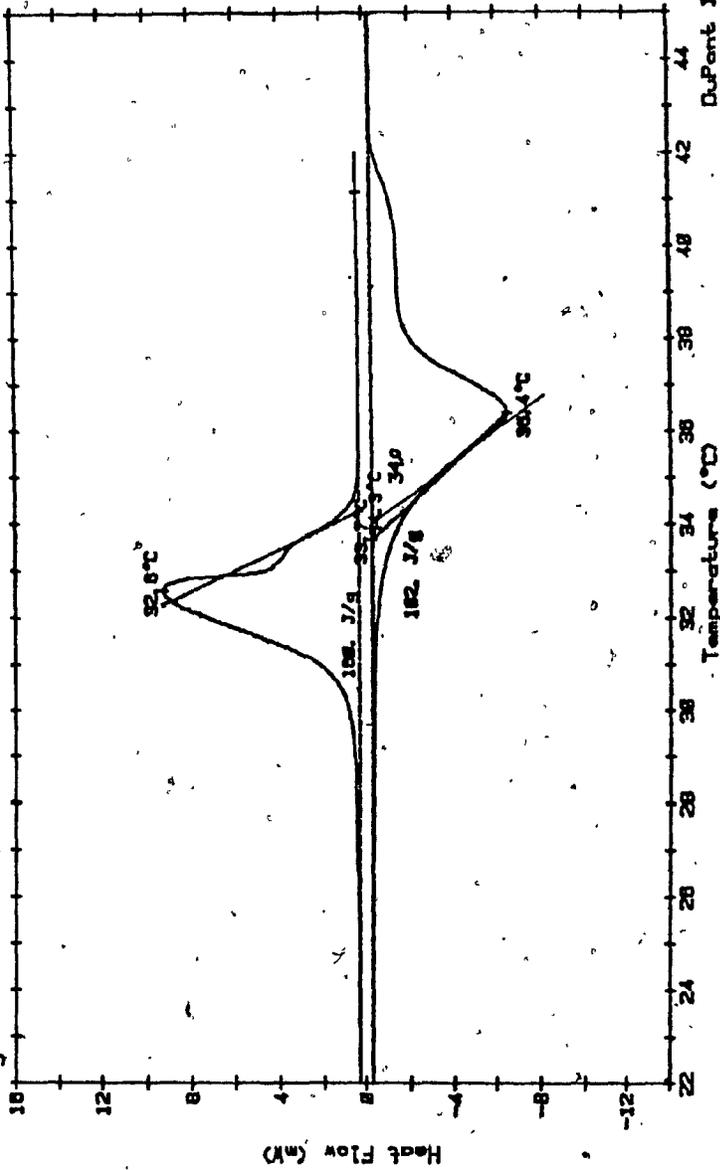


Fig. T-58. Thermogram of 80 mole % lauric : 20 mole % stearic.

Sampler 82.5L/17.5B  
Size 8.496HC  
Rate 2C/MIN  
Program Interactive DSC V2.8

DSC

Date 12-Jul-83 Time 11:13:44  
File FMS.22 DISC FOUR  
Operator  
Plotted 12-Jul-83 12:02:23

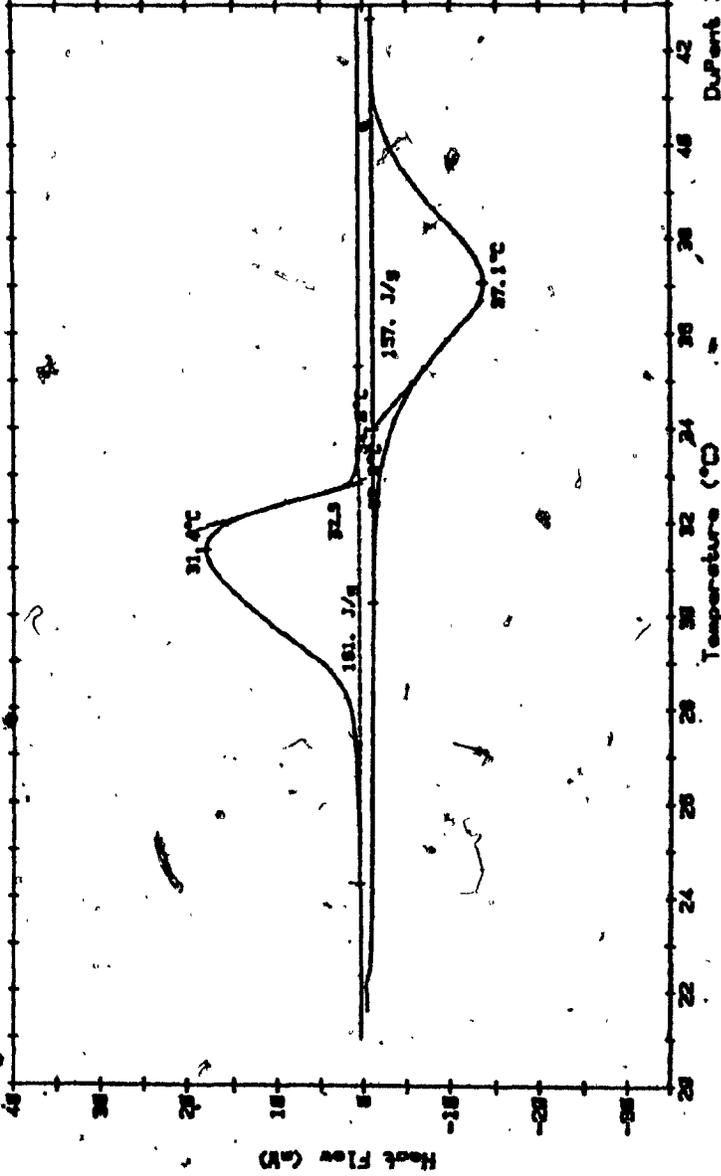


Fig. T-59. Thermogram of 82.5 mole % lauric ; 17.5 mole % stearic.

Samples: 84L/16S  
Size: 7.908 MG  
Rate: 2 C/MIN  
Program: Interactive DSC V2.8

DSC

Date: 12-Jul-83 Time: 20:12:58  
File: FUKS.23 D1SC FOUR  
Operator:  
Plotted: 12-Jul-83 20:51:32

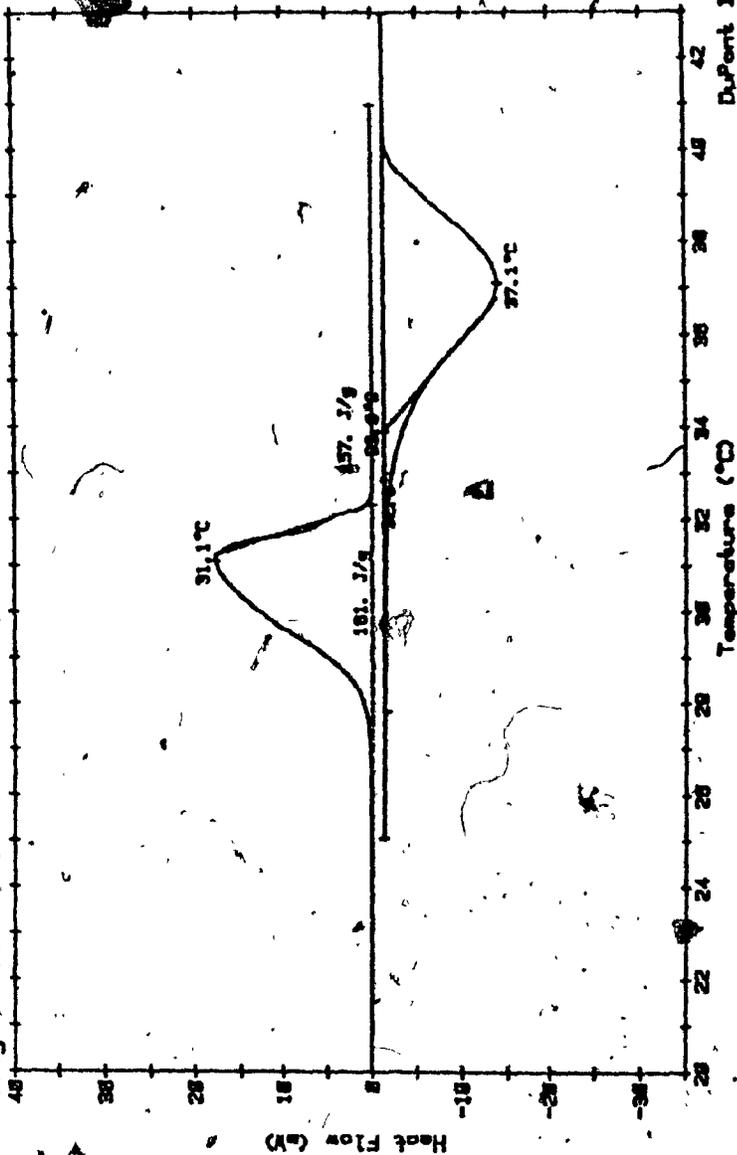


Fig. T-60. Thermogram of 84 mole % lauric, 16 mole % stearic.

Sample 65L/155  
Size 5.836 MG  
Rate 2 C/MIN  
Program Interactive DSC V2.8  
Date 8-Jul-83 Time 22:58:81  
File FLKS.88 D18C FOUR  
Operator  
Plotted 8-Jul-83 23:31:41

### DSC

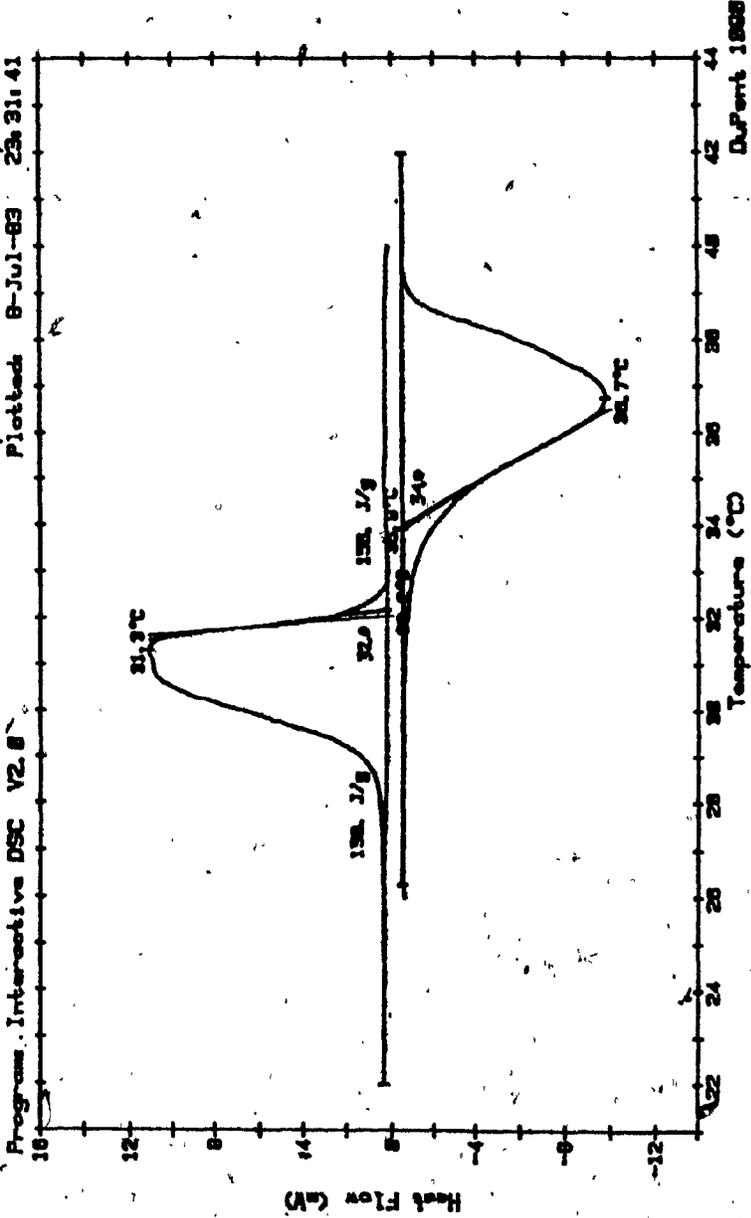


Fig. T-61. Thermogram of 85 mole % lauric : 15 mole % stearic.

Sample 86L/14P  
Size 3.973 MG  
Rate 2 C/MIN  
Program Interactive DSC V2.8

### DSC

Date 12-Jul-89 Time 20:57:48  
File FUMS.24 DISC FOUR  
Operator  
Plotted 12-Jul-89 21:31:23

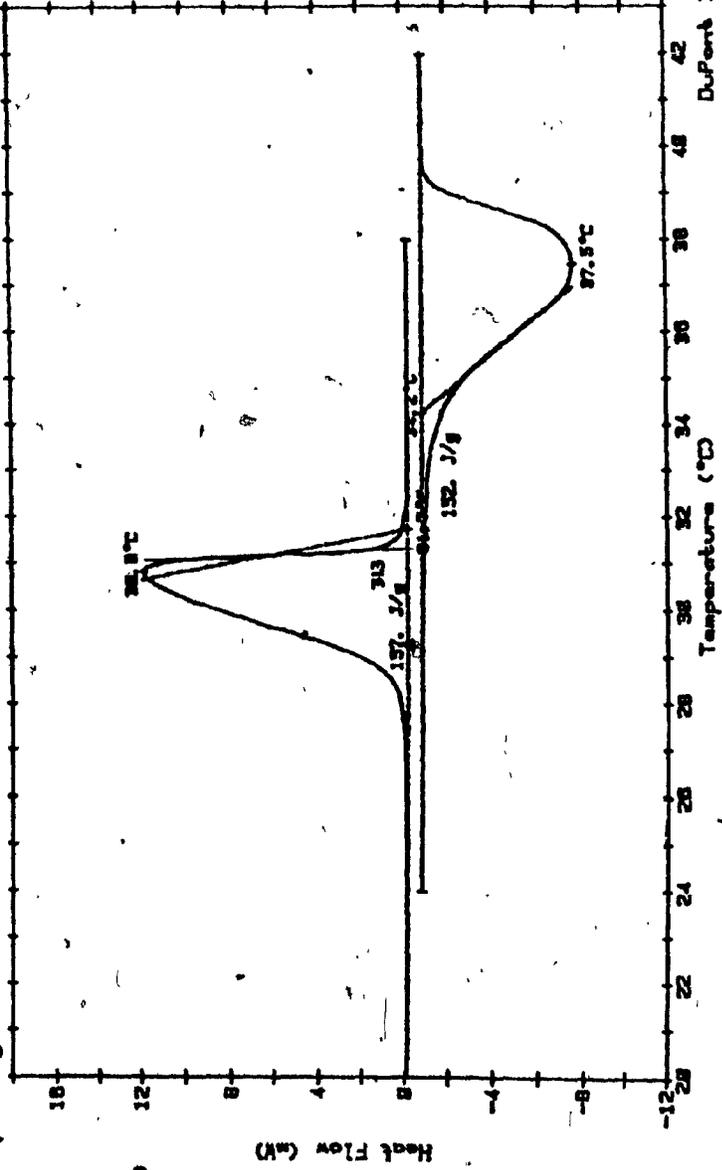


Fig. T-62. Thermogram of 86 mole % lauric ; 14 mole % stearic.

Sample 87.5/12.55  
Size 8.152 MG  
Rate 2 C/MIN  
Program Interactive DSC V2.8  
Date 12-Jul-83 Time 21:38:01  
File FUKS.25 DISC FOUR  
Operator  
Plotted 5-Sep-83 21:12:18

### DSC

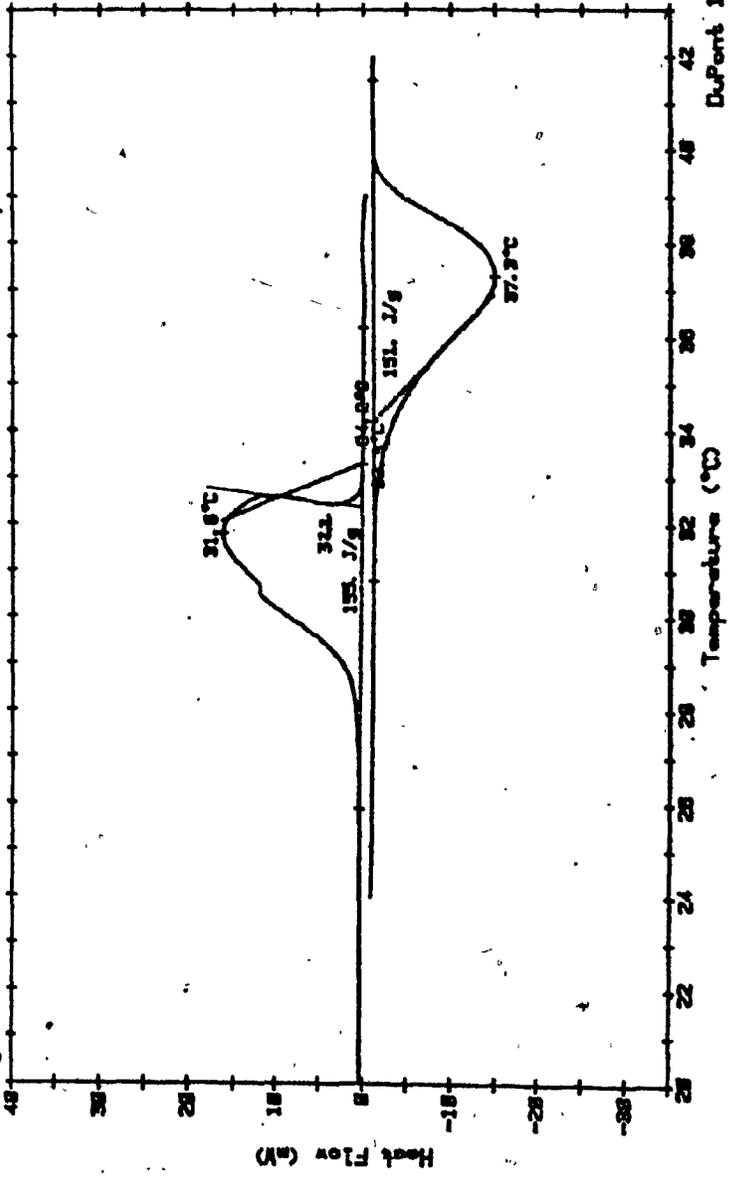


Fig. T-63. Thermogram of 87.5 mole % lauric : 12.5 mole % stearic.

Date: 8-Jul-68 Time: 21:34:05  
File: FLKS.84 DISC FOUR  
Operator:  
Plotted: 5-Sep-68 22:58:38

Sample: 80L/18S  
Size: 8.484 MG  
Rate: 2 C/MIN  
Program: Interactive DSC V2.0

# DSC

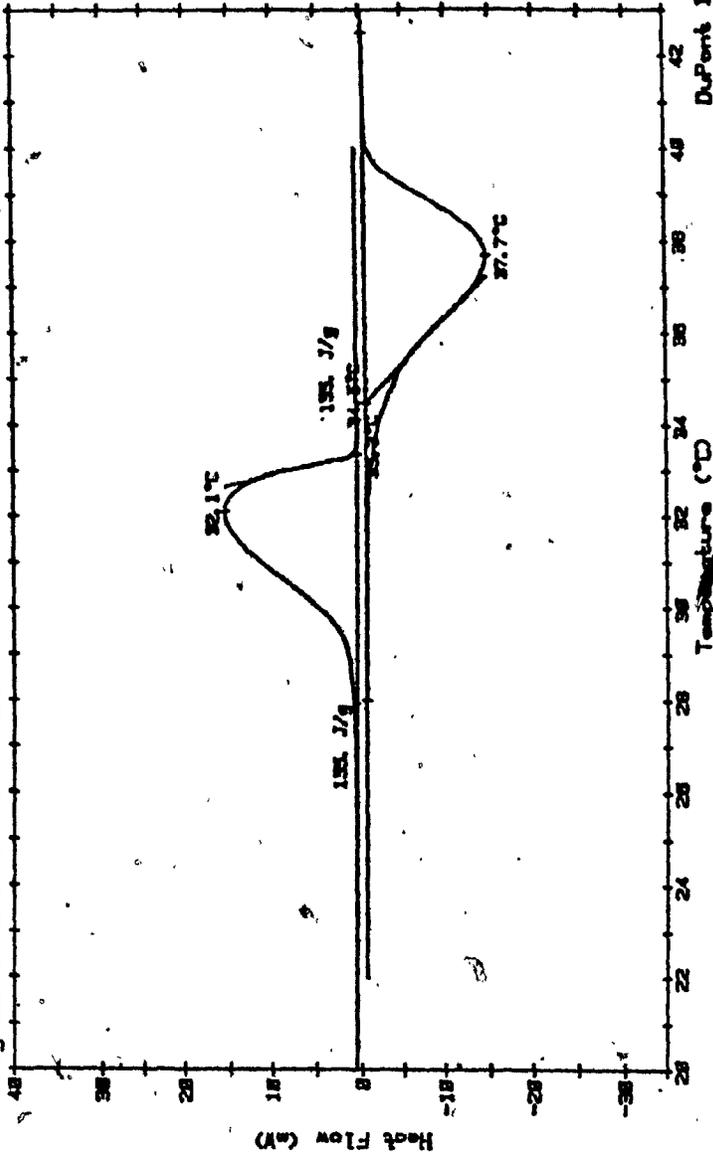


Fig. T-64. Thermogram of 90 mole % lauric ; 10 mole % stearic.

**BINARY MIXTURES OF LAURIC-PALMITIC ACIDS**

Date 17-Sep-63 Time 12:28:48  
File FLKS.43 DISK THREE  
Operator  
Plotted 17-Sep-63 13:28:28

### DSC

Sample 10L/80P  
Size 3.689 MG  
Rate 2 C/MIN  
Program Interactive DSC V2.8

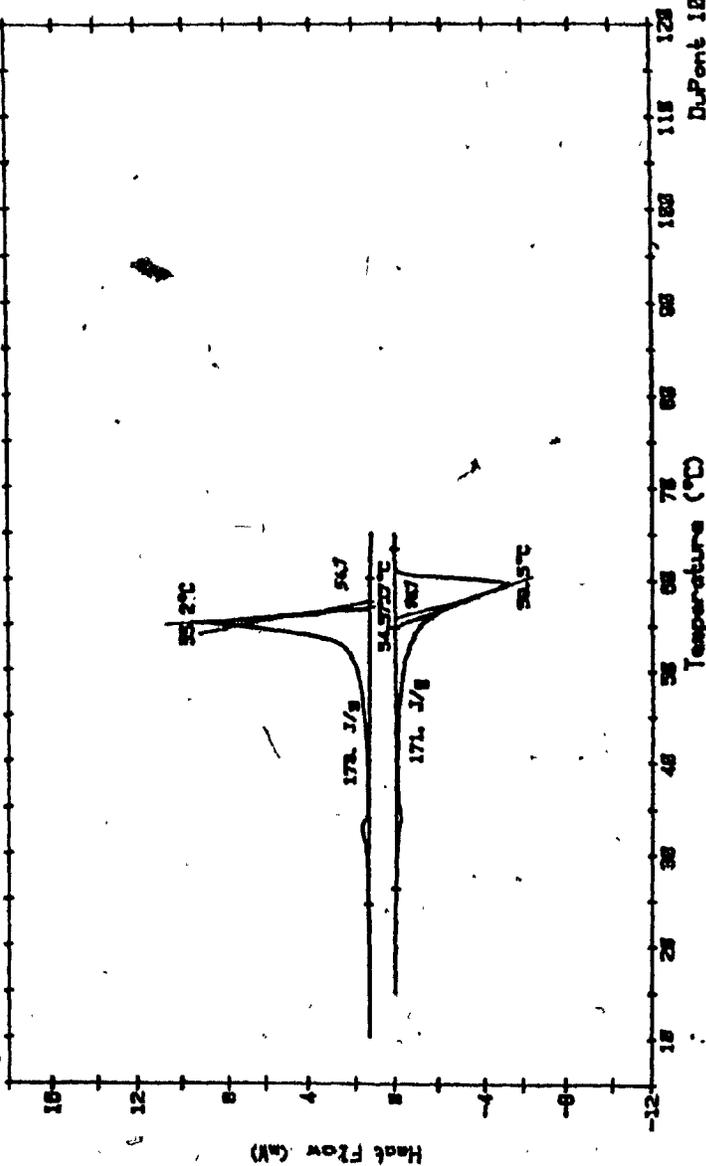


Fig. T-65. Thermogram of 10 mole % lauric : 90 mole % palmitic.

Sample 28L/80P  
Size 11.387MG  
Rate 2C/MIN  
Program Interactive DSC V2.8  
Plotted 18-Sep-83 21:03:49

### DSC

Date 9-Jun-83 Time 15:13:12  
File FUMS.B1 DISK TWO  
Operator

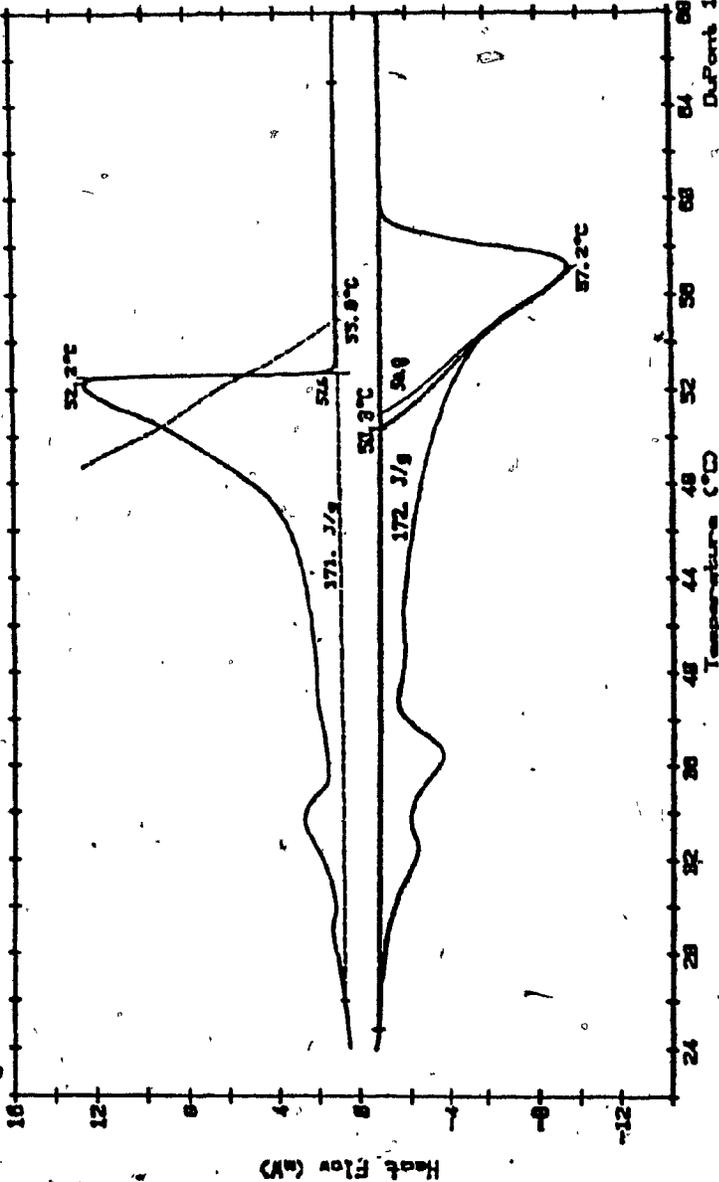


Fig. T-66. Thermogram of 20 mole % lauric ; 80 mole % palmitic. D.Pert 1802

Sample 30L/70P  
Size 18.188 MG  
Rate 2 C/MIN  
Program Interactive DSC V2.8

**DSC**

Date 1-Jul-83 Time 16:28:18  
File FWS.81 DISK THREE  
Operator  
Plotted 15-Sep-83 22:25:37

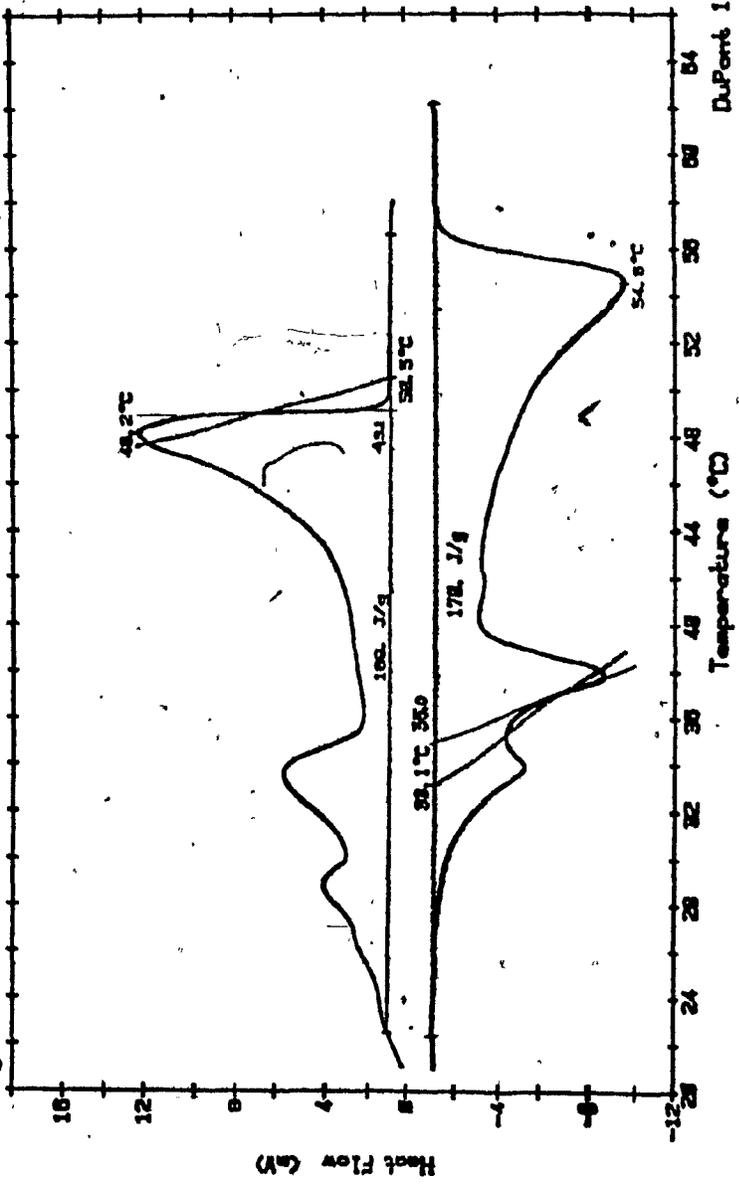


Fig. T-67. Thermogram of 30 mole % lauric ; 70 mole % palmitic. DuPont 1088

Sample 48L/DBP  
Size 8.818 MG  
Rate 20/MIN  
Program Interactive DSC V2.8  
Dates 15-Jun-83 Time 12:58:48  
Files FUKS.83 DISK TWO  
Operator  
Plotted 1-Jul-83 17:24:53

### DSC

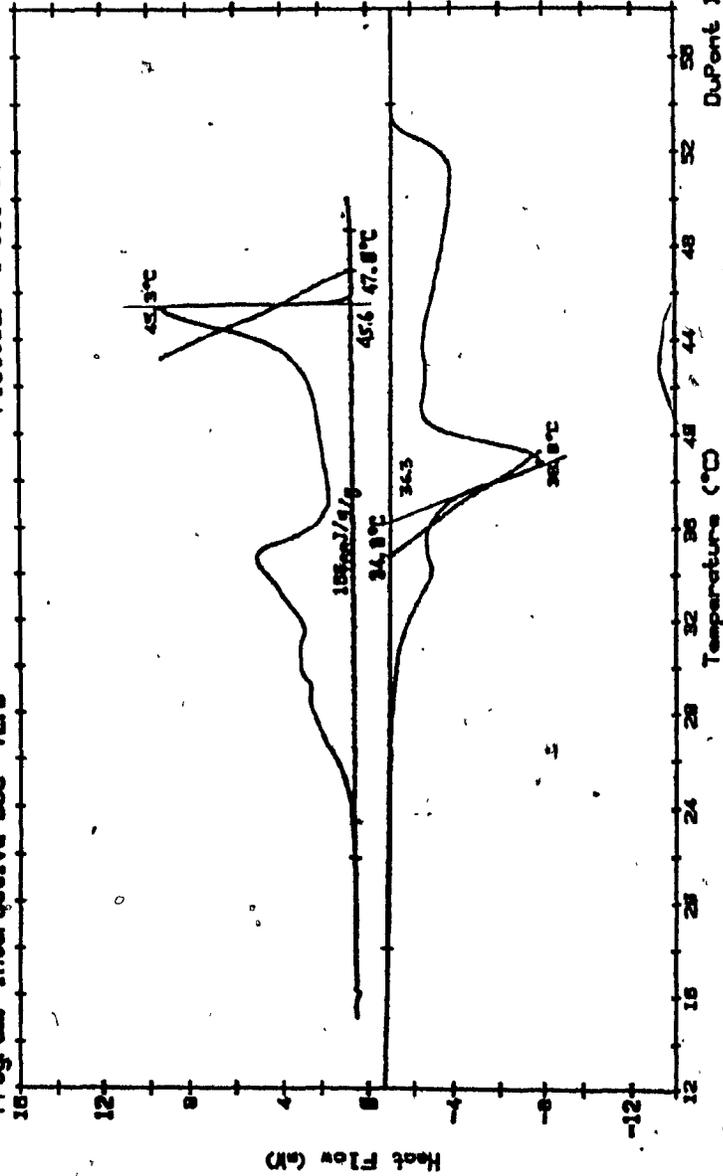


Fig. T-68. Thermogram of 40 mole % lauric : 60 mole % palmitic.  
DuPont 1898B

Sample 50L/50P

Size 5.158 MG

Rate 20/MIN

Program Interactive DSC V2.8

Date 15-Jun-83

Time 14:58:87

File FUKS.84 DISK TWO

Operator

Plotted 1-Jul-83

17:41:21

# DSC

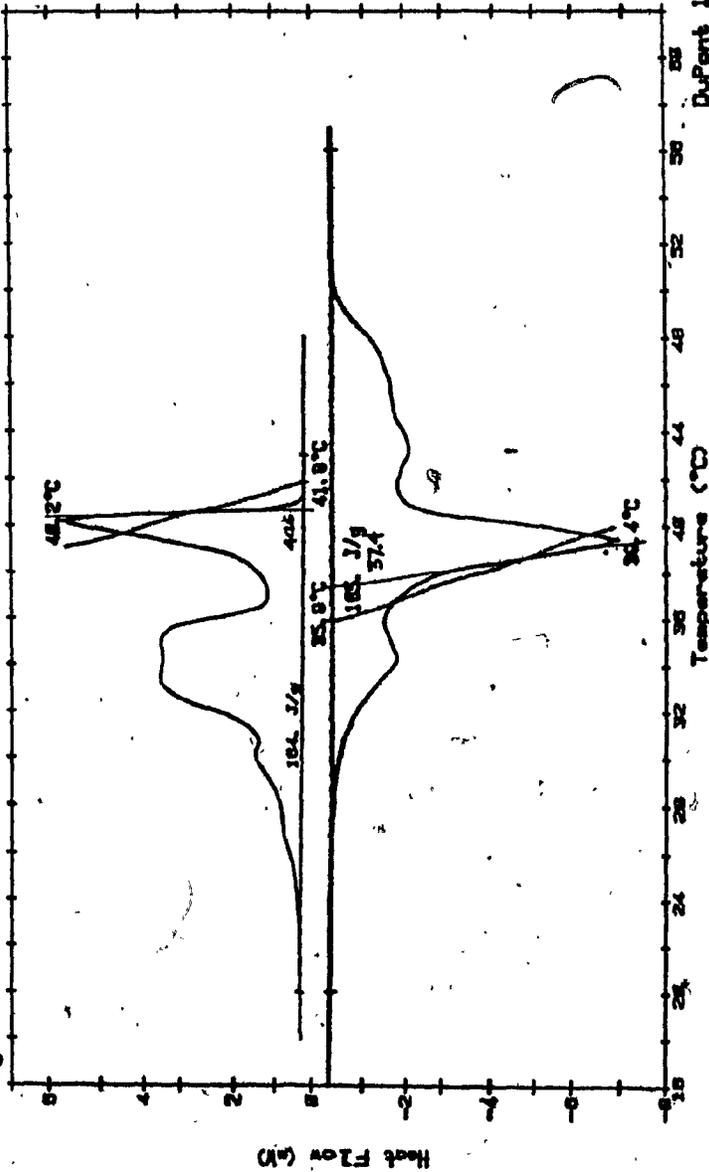


Fig. T-69. Thermogram of 50 mole % lauric : 50 mole % palmitic.

Sample 66XL 48XP  
Size 9.867MG  
Rate 20EG/MIN  
Program Interactive DSC V2.0

DSC  
Date 18-Jun-83 Time 15:44:14  
File DORINA.11 DISK NINE  
Operator  
Plotted 15-Sep-83 23:21:53

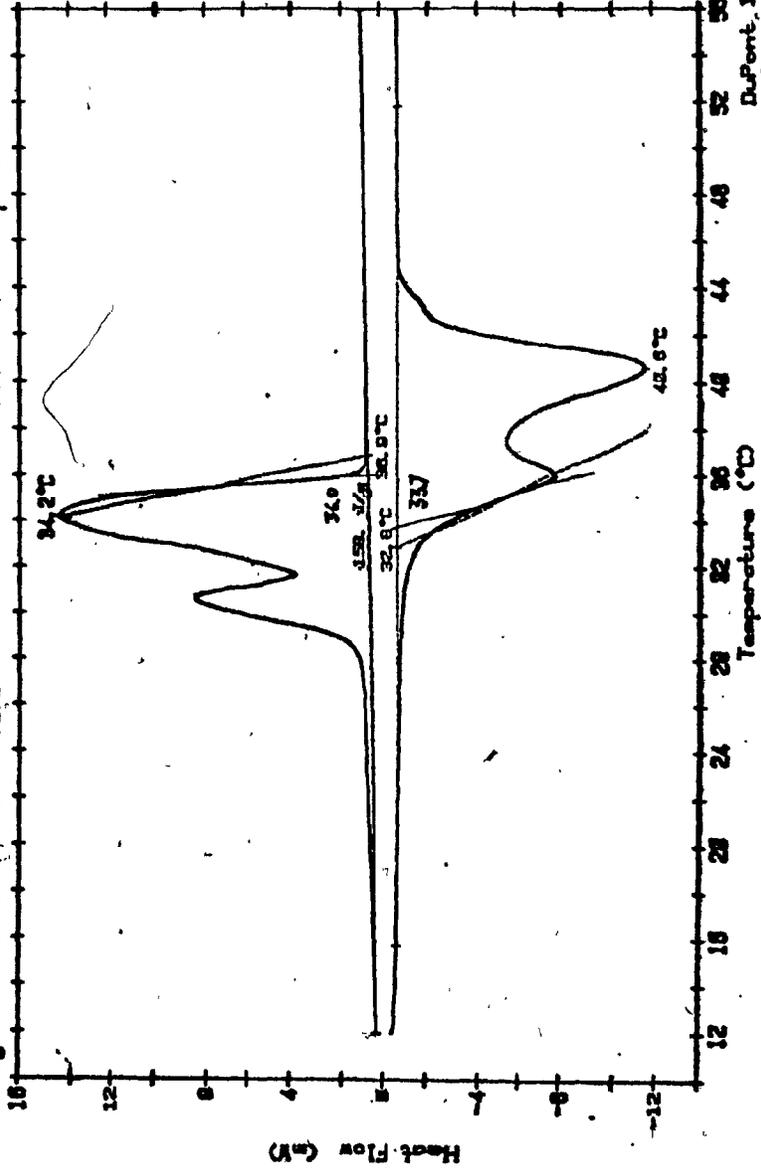


Fig. T-70. Thermogram of 60 mole % lauric : 40 mole % palmitic.

Sample: 7BL/30P  
Size: 12.377 MG  
Rate: 2C/MIN  
Program: Interactive DSC V2.0

DSC

Date: 17-Jun-83 Time: 13:42:58  
File: FUKS.08 DISK TWO  
Operator:  
Plotted: 1-Jul-83 17:58:25

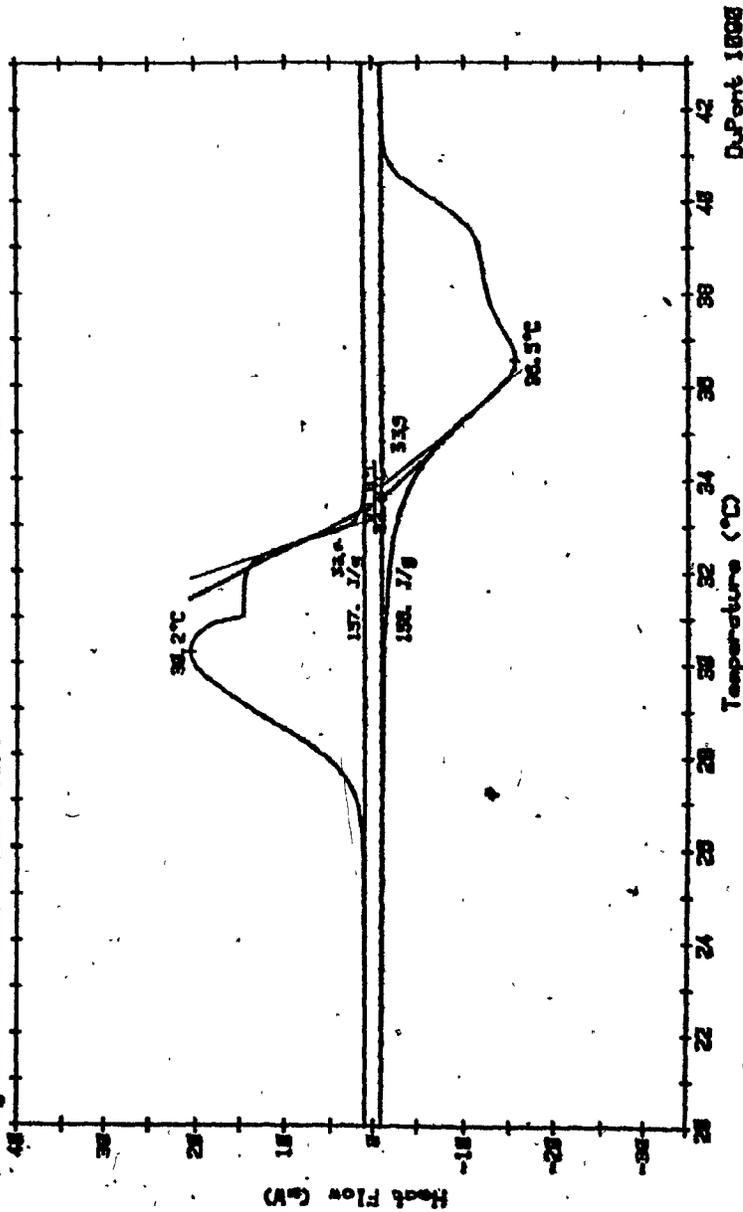


Fig. T-71. Thermogram of 70 mole % lauric ; 30 mole % palmitic.

Date 20-Jun-83 Time 18:57:82  
File FUKS.87 DISK TWO  
Operator  
Plotted 15-Sep-83 22:49:83

### DSC

Sample 71L/28P  
Size 8.134 MG  
Rate 50/MIN  
Program Interactive DSC V2.8

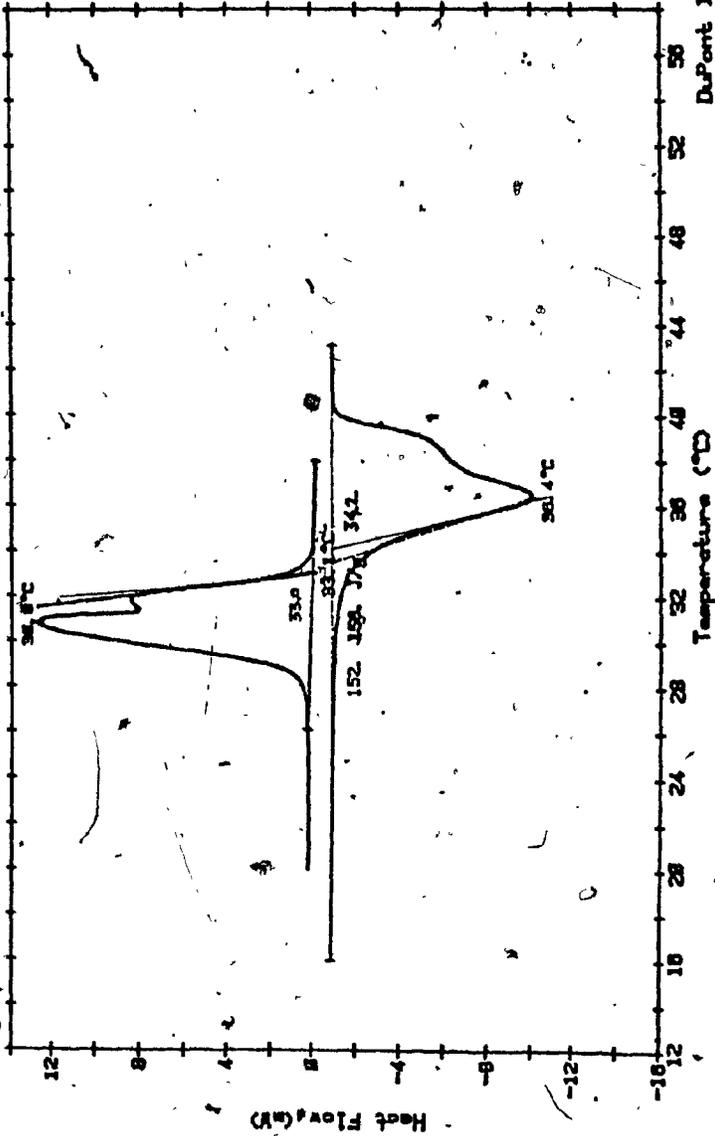


Fig. T-72. Thermogram of 71 mole % lauric : 29 mole % palmitic.

Date: 23-June-83 Time: 13:04:41  
File: FLKS.19 DISK TWO  
Operator:  
Plotted: 15-Sep-83 23:00:55

Sample: 72.5L/27.5P  
Size: 3.028MG  
Rate: 2C/MIN  
Program: Interactive DSC V2.0

### DSC

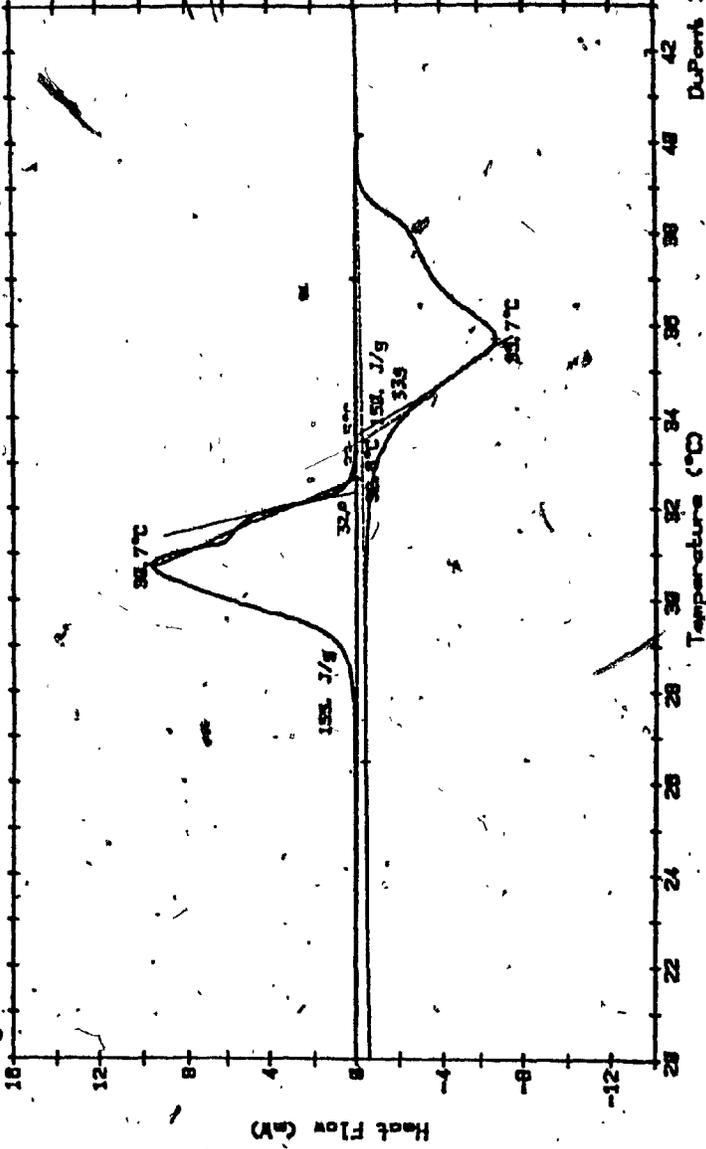


Fig. T-73. Thermogram of 72.5 mole % lauric : 27.5 mole % palmitic.

Dates 17-Jun-83 Times 12:16:84  
Files FUKS.05 DISK TWO  
Operator  
Plotted 15-Sep-83 23:32:38

Samples 75L/25P  
Size 8.573 MG  
Rate 2C/MIN  
Program Interactive DSC V2.8

### DSC

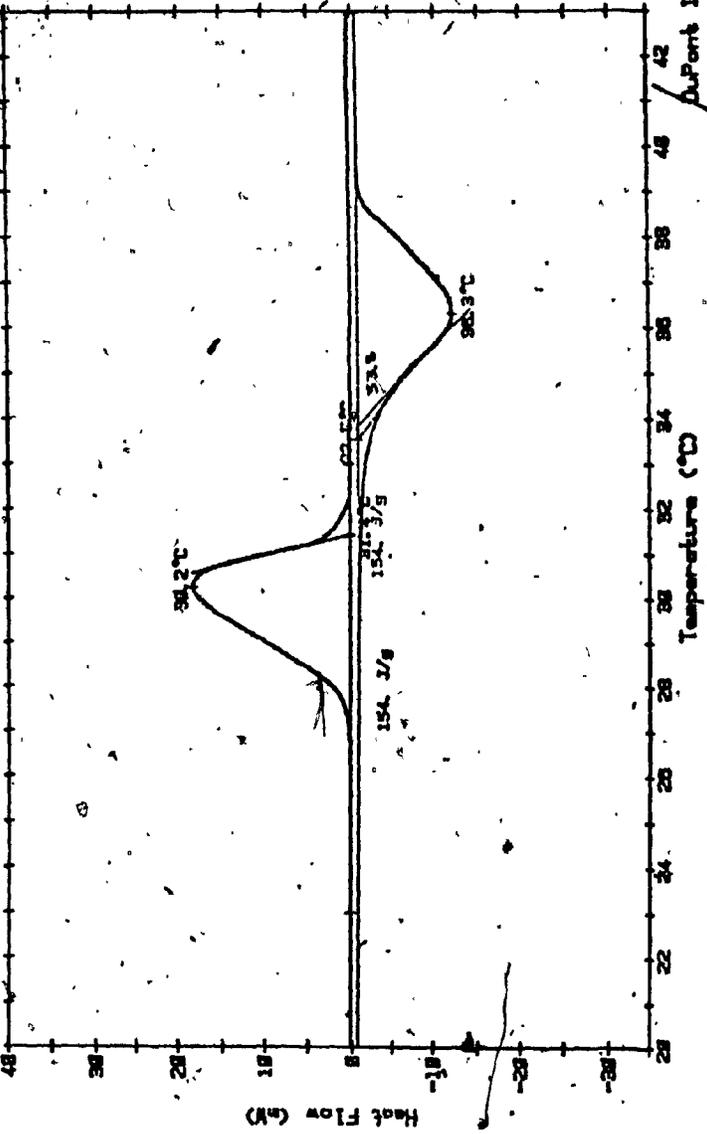


Fig. T-74. Thermogram of 75 mole % lauric + 25 mole % palmitic.

Sample: 77.5L/22.5P  
Size: 8.824 MG  
Rate: 20/MIN  
Program: Interactive DSC V2.0  
Date: 23-Jun-83 Time: 12:21:48  
File: FUNK.12 DISK TWO  
Operator:  
Plotted: 15-Sep-83 23:47:08

### DSC

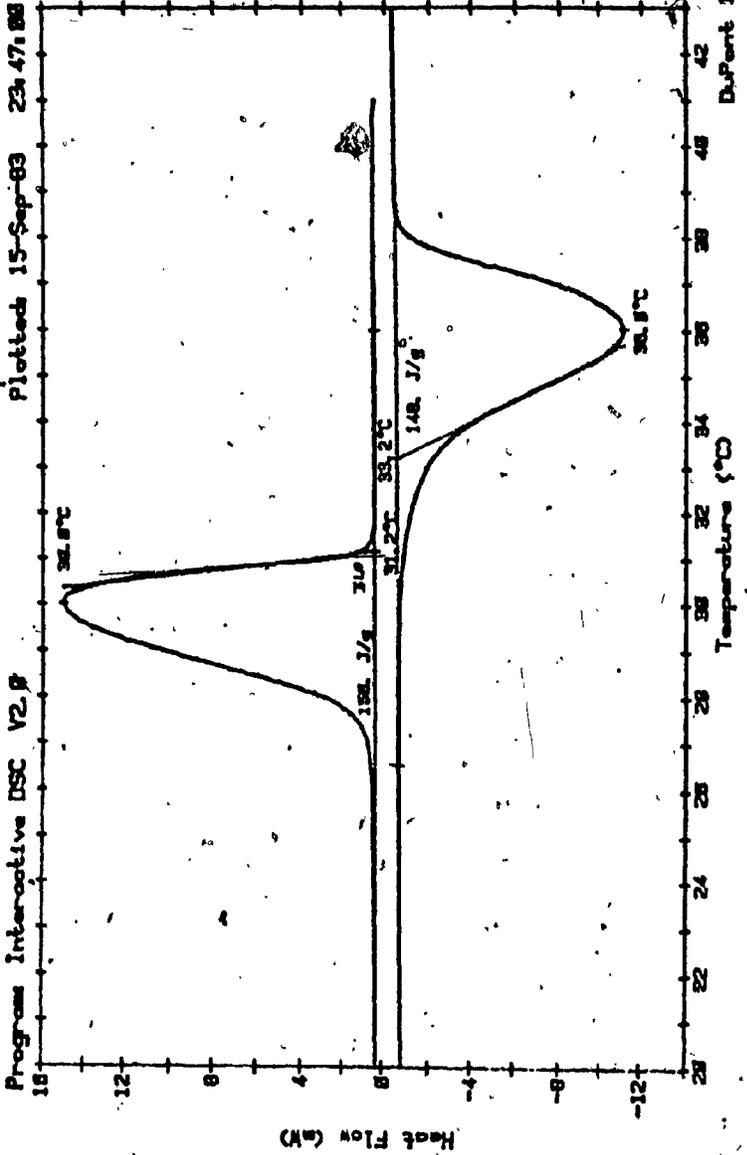


Fig. T-75. Thermogram of 77.5 mole % lauric : 22.5 mole % palmitic.

Date: 1-Jul-83 Time: 13:27:42  
File: FLKS.88 DISK TWO  
Operator:  
Plotted: 16-Sep-83 8:04:00

Sample: 78L/21P  
Size: 8.938 MG  
Rate: 2 C/MIN  
Program: Interactive DSC V2.8

### DSC

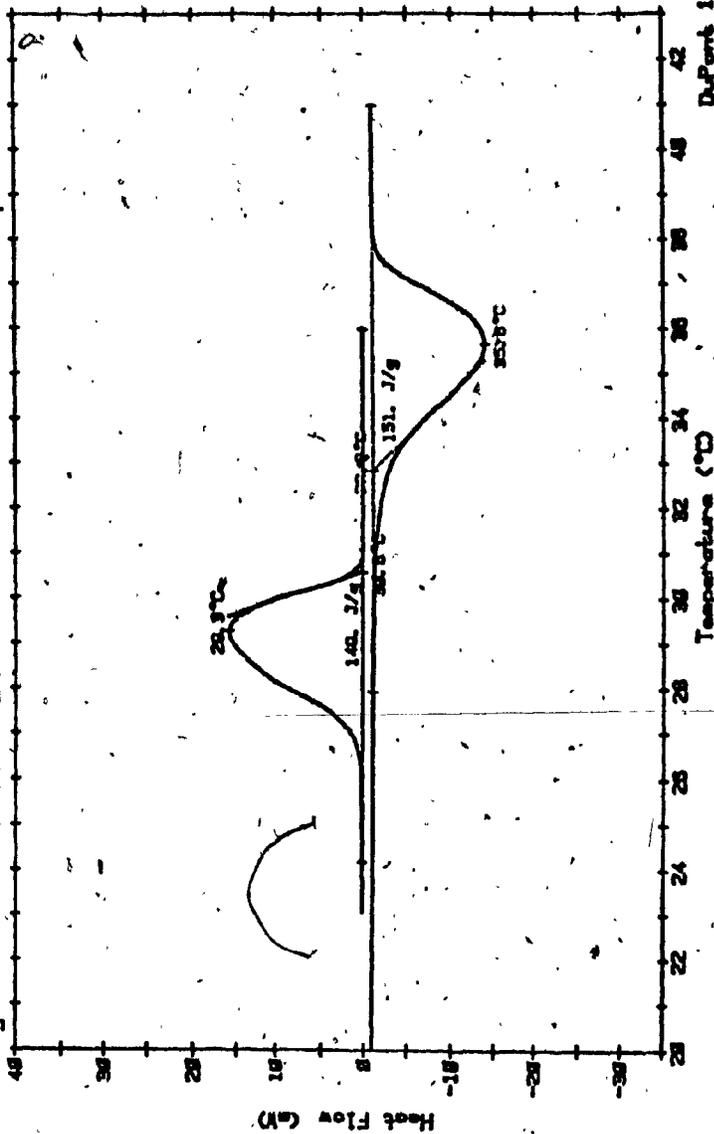


Fig. T-76. Thermogram of 79 mole % lauric ; 21 mole % palmitic.

Sample: BBL/26P  
Size: 8.558 MG  
Rate: 2 C/MIN  
Program: Interactive DSC V2.0  
Date: 1-Jul-83 Time: 11:52:23  
File: FUMS.28 DISK TWO  
Operator:  
Plotted: 18-Sep-83 8:15:00

# DSC

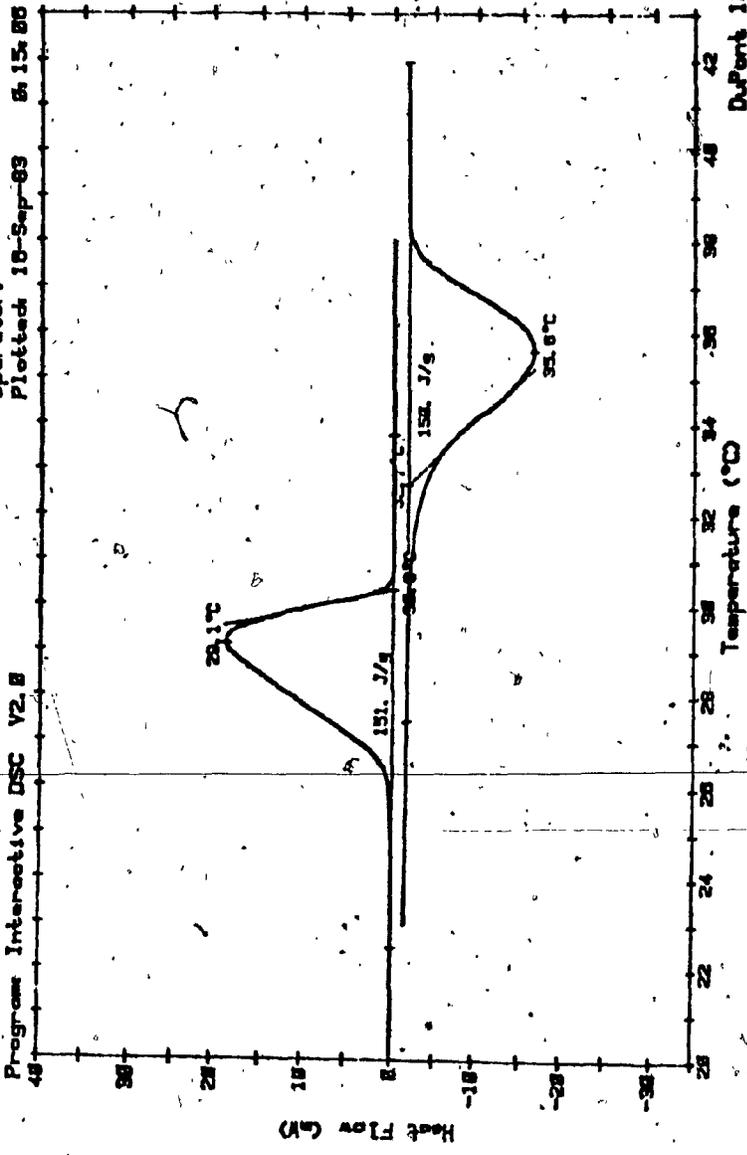


Fig. T-77. Thermogram of 80 mole % lauric : 20 mole % palmitic.

Date: 28-Jun-83 Time: 21:23:21  
File: FMS.21 DISK TWO  
Operator:  
Plotted: 16-Sep-83 0:33:00

Sample: 01L/18P  
Size: 12.383 MG  
Rate: 2 C/MIN  
Program: Interactive DSC V2.8  
DSC

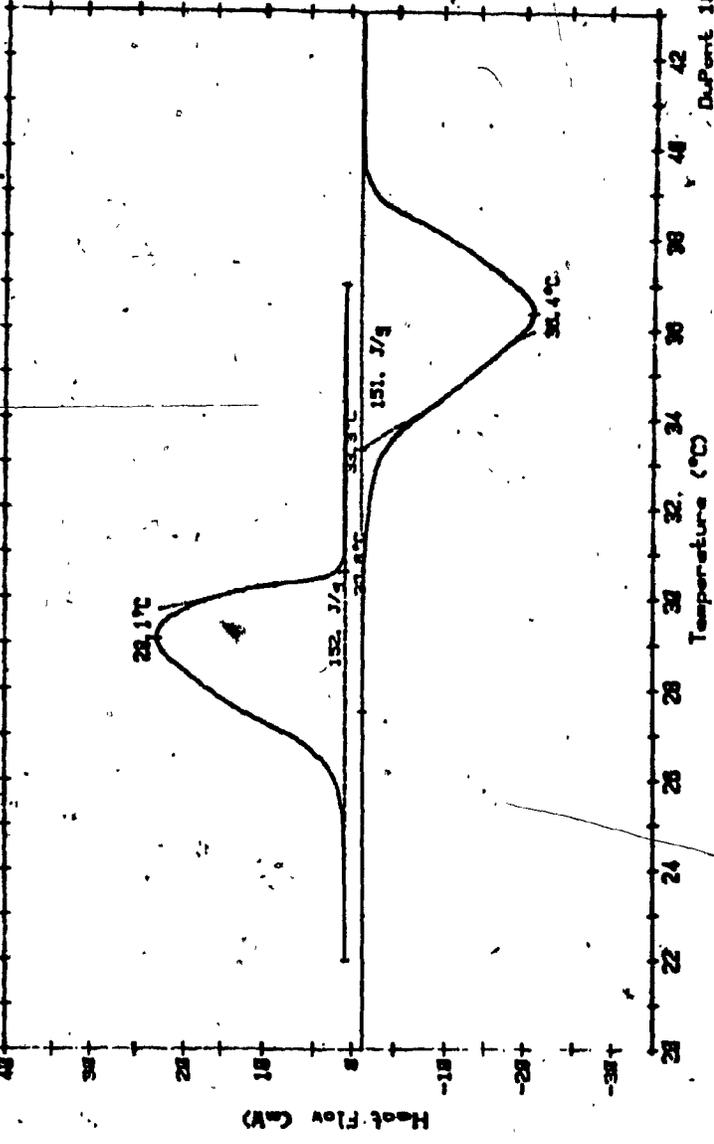


Fig. T-78. Thermogram of 81 mole % lauric : 19 mole % palmitic. DuPont 1898

Sample: 82.5L/17.5P  
Size: 5.845 M6  
Rate: 2 C/min  
Program: Interactive DSC V2.8

# DSC

Date: 28-Jun-83 Time: 22:58:28  
File: FUKS.25 DISK TWO  
Operator:  
Plotted: 18-Sep-83 21:58:24

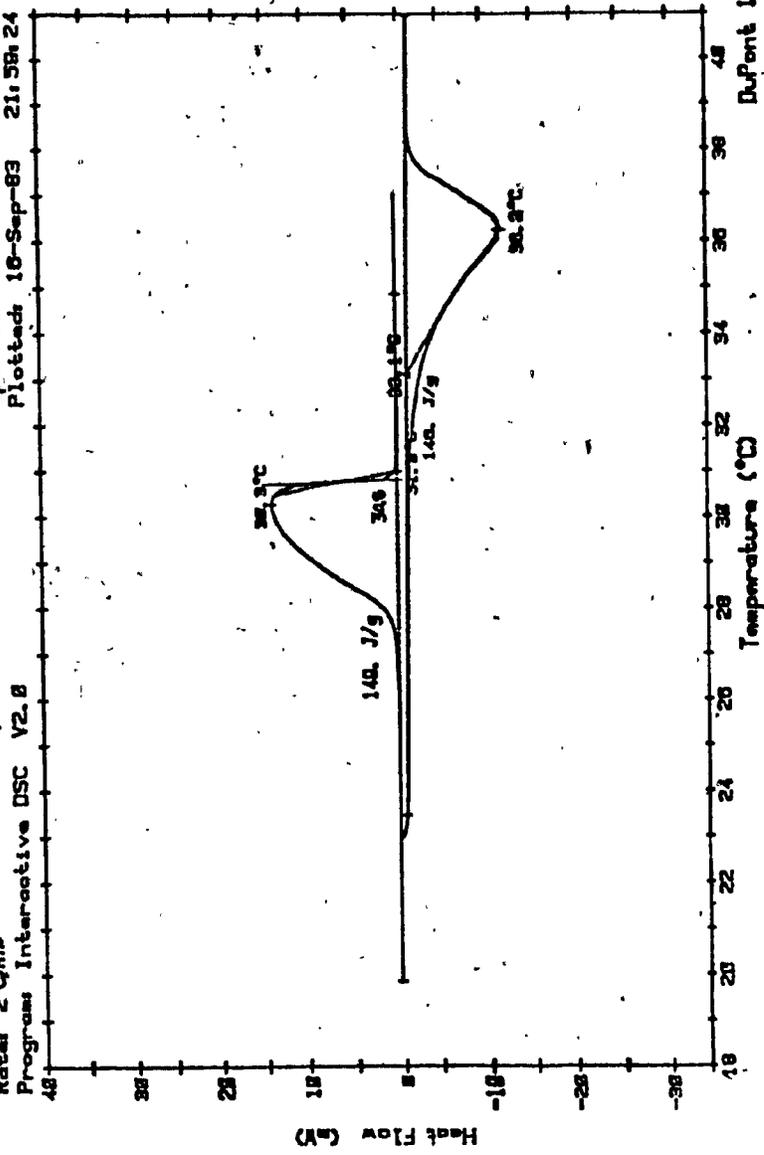


Fig. T-79. Thermogram of 82.5 mole % lauric; 17.5 mole % palmitic.

Sample: 85L/15P  
Size: 6.363 MG  
Rate: 2C/MIN  
Program: Interactive DSC V2.8

### DSC

Date: 29-Jun-83 Time: 11:35:54  
File: FUNKS.11 DISK TWO  
Operator:  
Plotted: 29-Jun-83 12:11:49

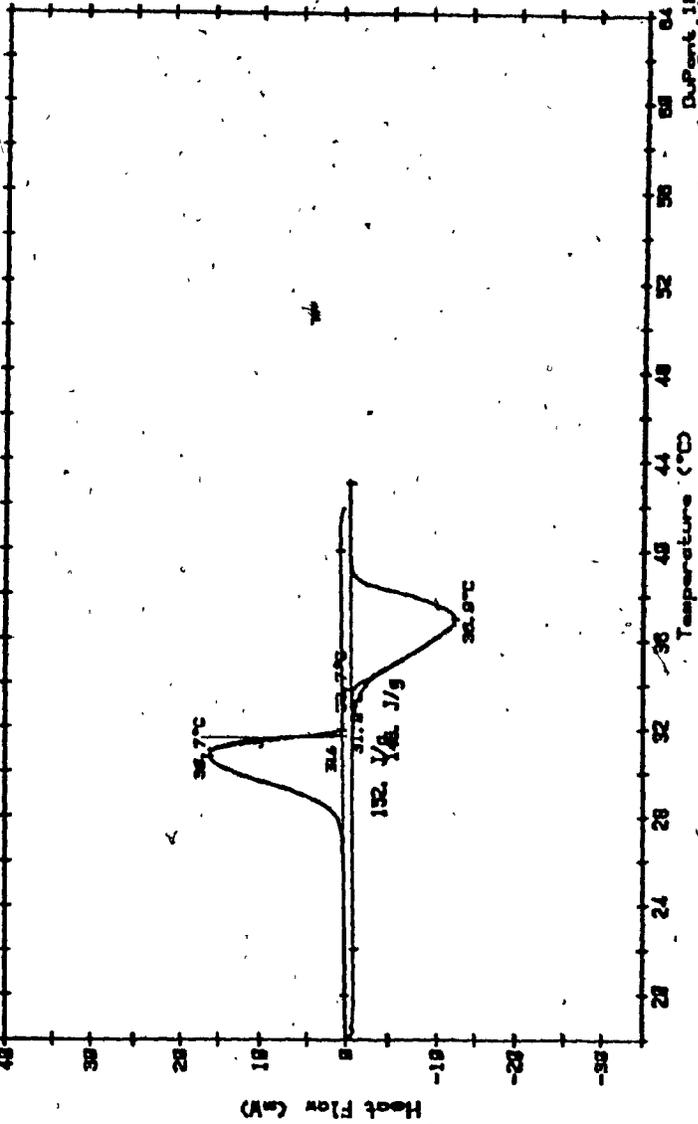


Fig. T-80. Thermogram of 85 mole % lauric ; 15 mole % palmitic.

Date: 20-Jun-83 Time: 19:48:24  
File: FUKS.10 DISK TWO  
Operator:  
Plotted 10-Sep-83 21:17:03

Sample: 90L/18P  
Size: 4.975 MG  
Rate: 2C/MIN  
Program: Interactive DSC V2.0

DSC

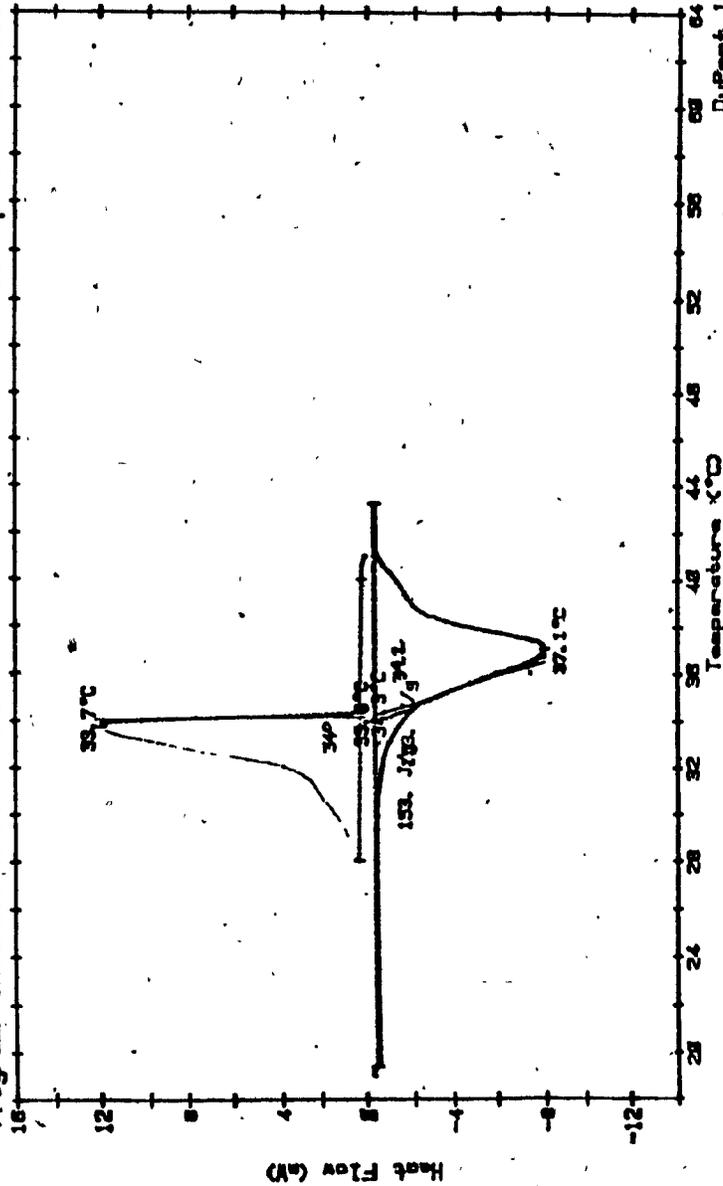


Fig. T-81. Thermogram of 90 mole % lauric / 10 mole % palmitic.

X-RAY DIFFRACTOMETRIC SPECTRA

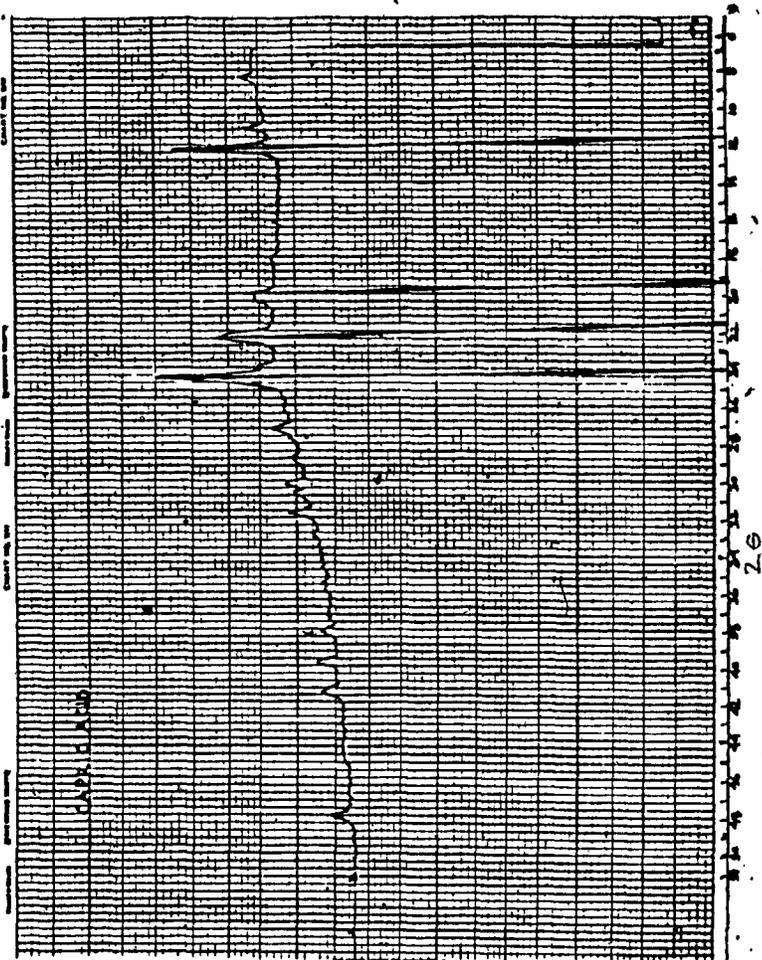


Fig. S-1. X-ray spectrum of capric acid.

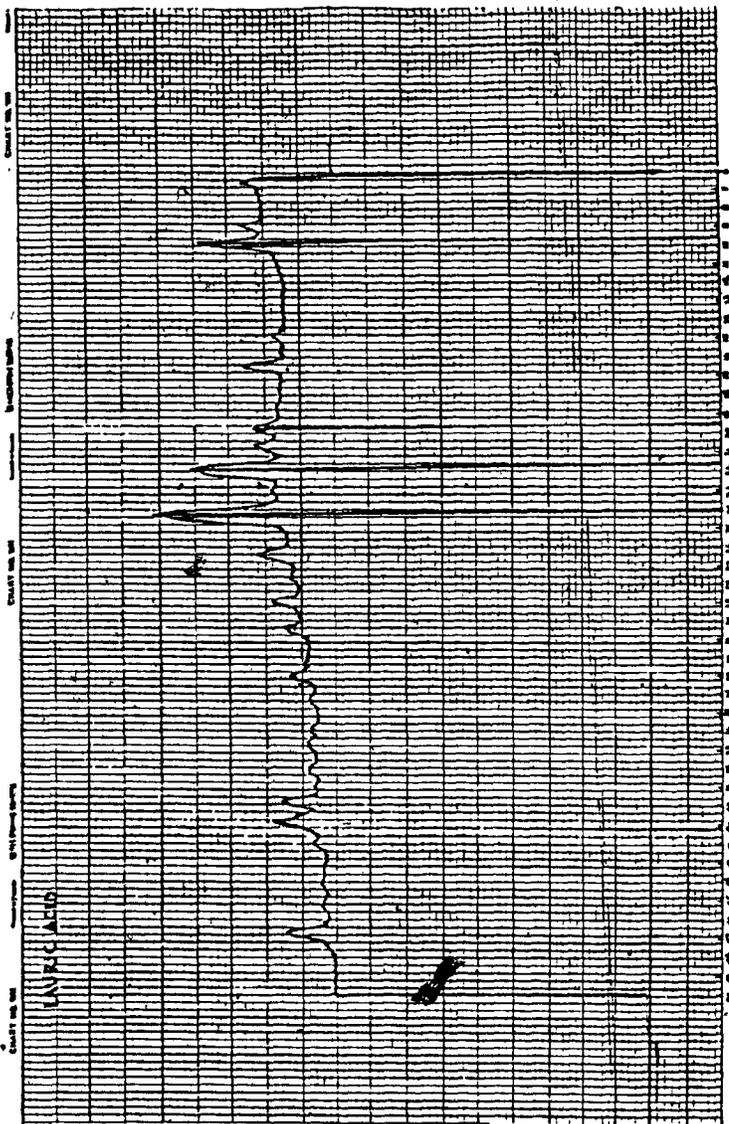


Fig. S-2. X-ray spectrum of lauric acid.

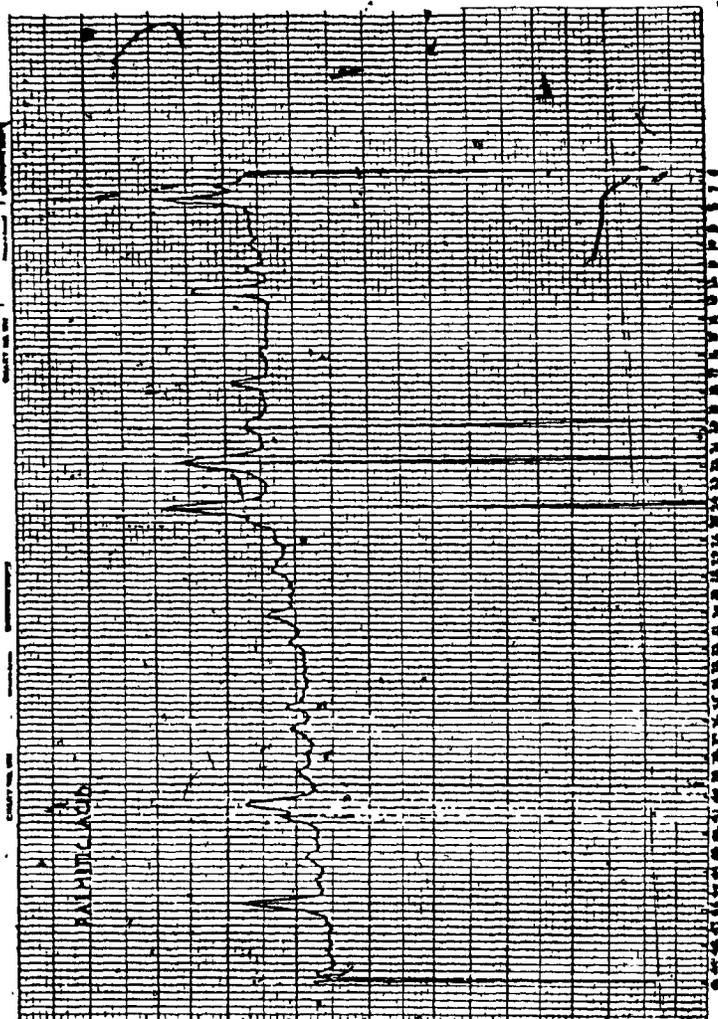


Fig. S-3. X-ray spectrum of palmitic acid.



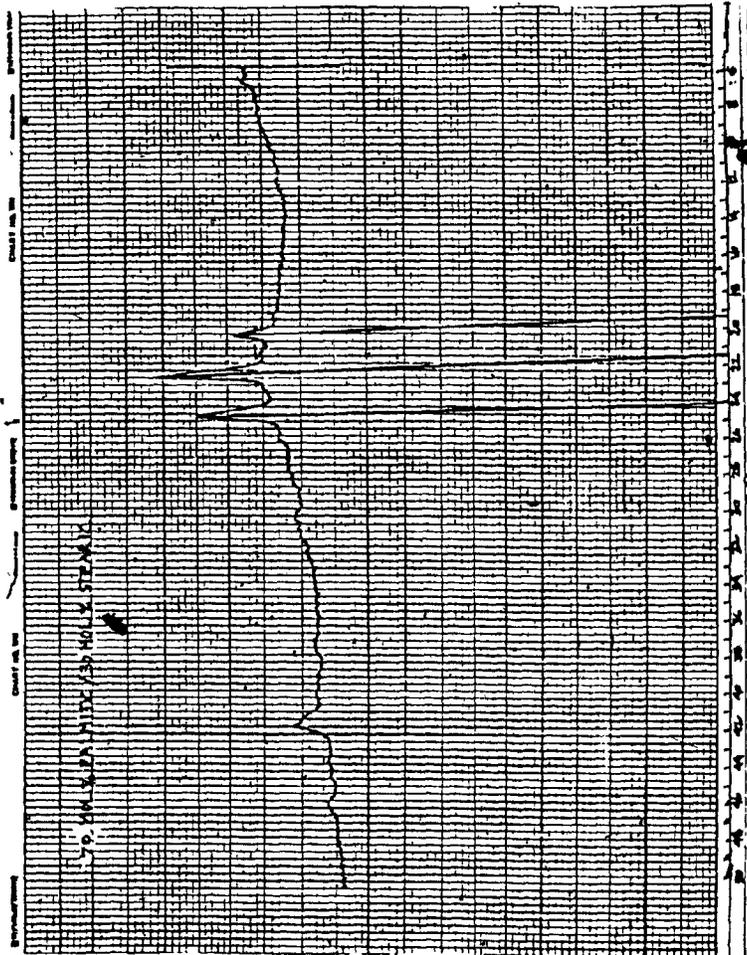


Fig. S-5. X-ray spectrum of 70 mole % palmitic : 25 mole % stearic.

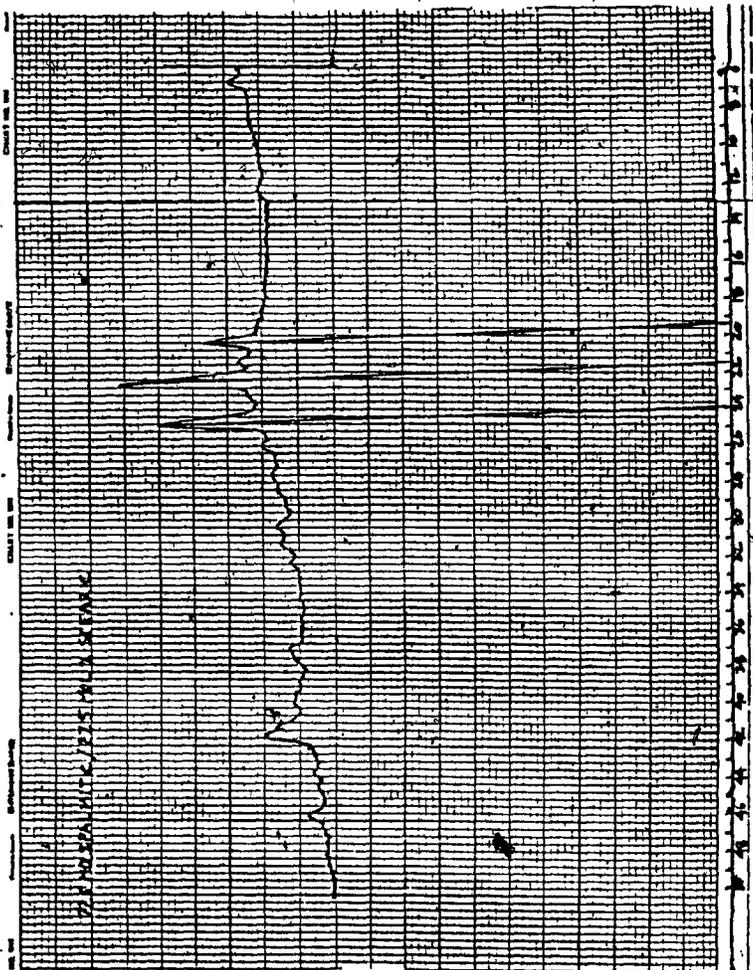


Fig. S-6. X-ray spectrum of 27.5 mole % palmitic : 27.5 mole % stearic.

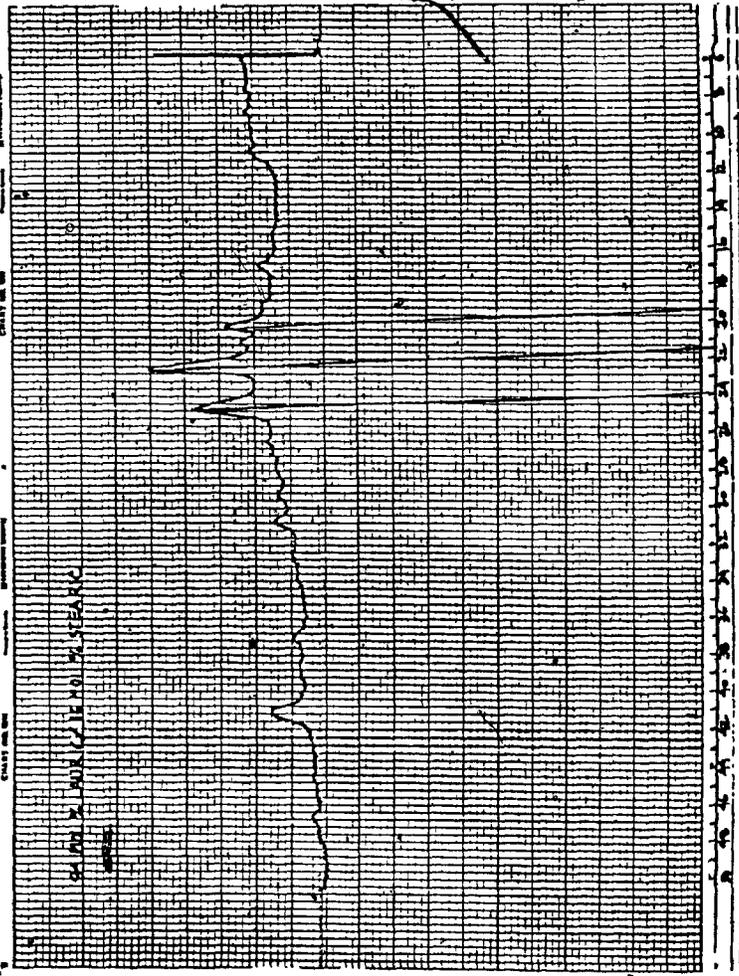


Fig. S-7. X-ray spectrum of 84 mole % lauric : 16 mole % stearic.

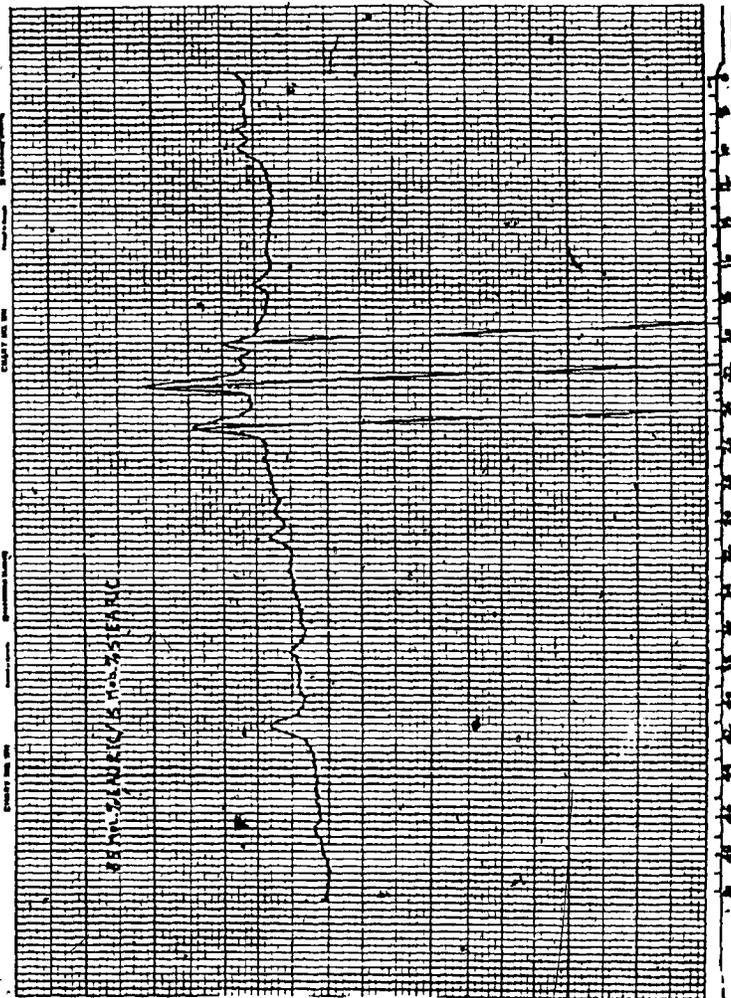


Fig. S-8. X-ray spectrum of 85 mole % lauric : 15 mole % stearic.

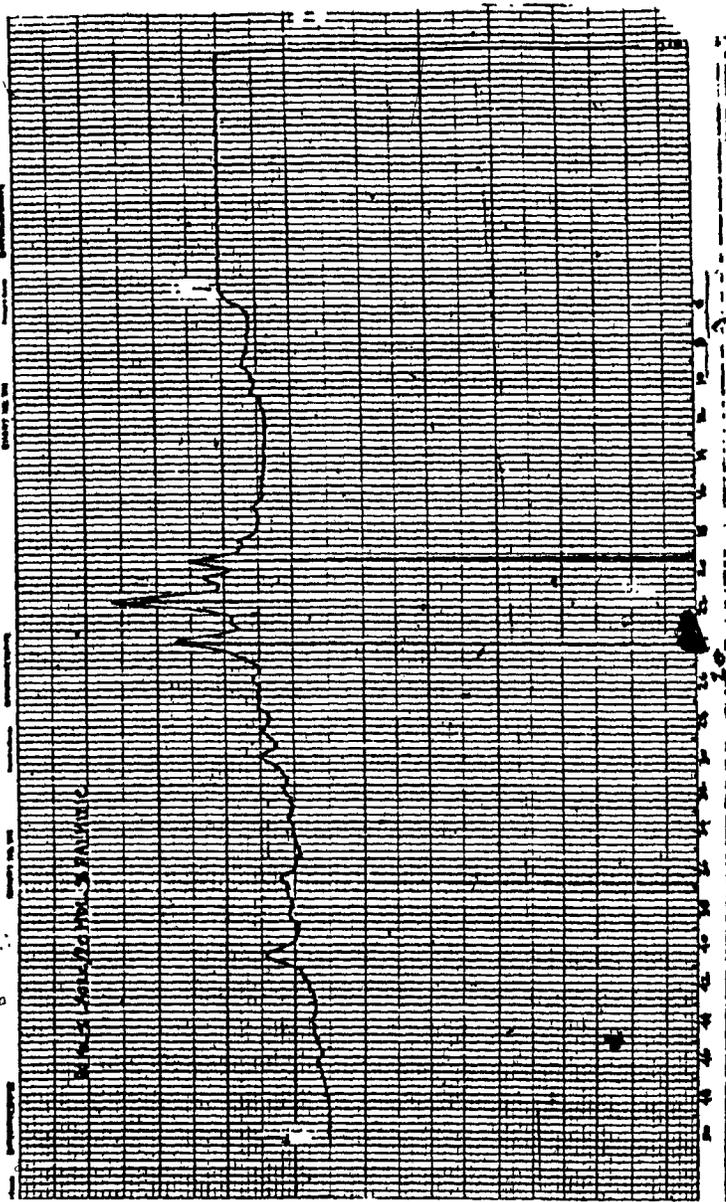


Fig. S-9. X-ray spectrum of 80 mole % lauric : 20 mole % palmitic.

INFRARED SPECTRA

17

SPECTRUM NO. \_\_\_\_\_

DATE \_\_\_\_\_

SAMPLE 1. 74% CA - 26% LA  
2. 85% LA - 15% SA  
3. 72.5% PA - 27.5% SA  
4. 80% LA - 20% PA

SOURCE \_\_\_\_\_

STRUCTURE \_\_\_\_\_

PATH \_\_\_\_\_ mm

SOLVENT \_\_\_\_\_

CONC. \_\_\_\_\_

PHASE \_\_\_\_\_

COMMENTS *KBr disk*  
*10mg/1000mg*  
*74% CA - 26% LA*  
*cappilary film between*  
*KBr disks.*

ANALYST \_\_\_\_\_

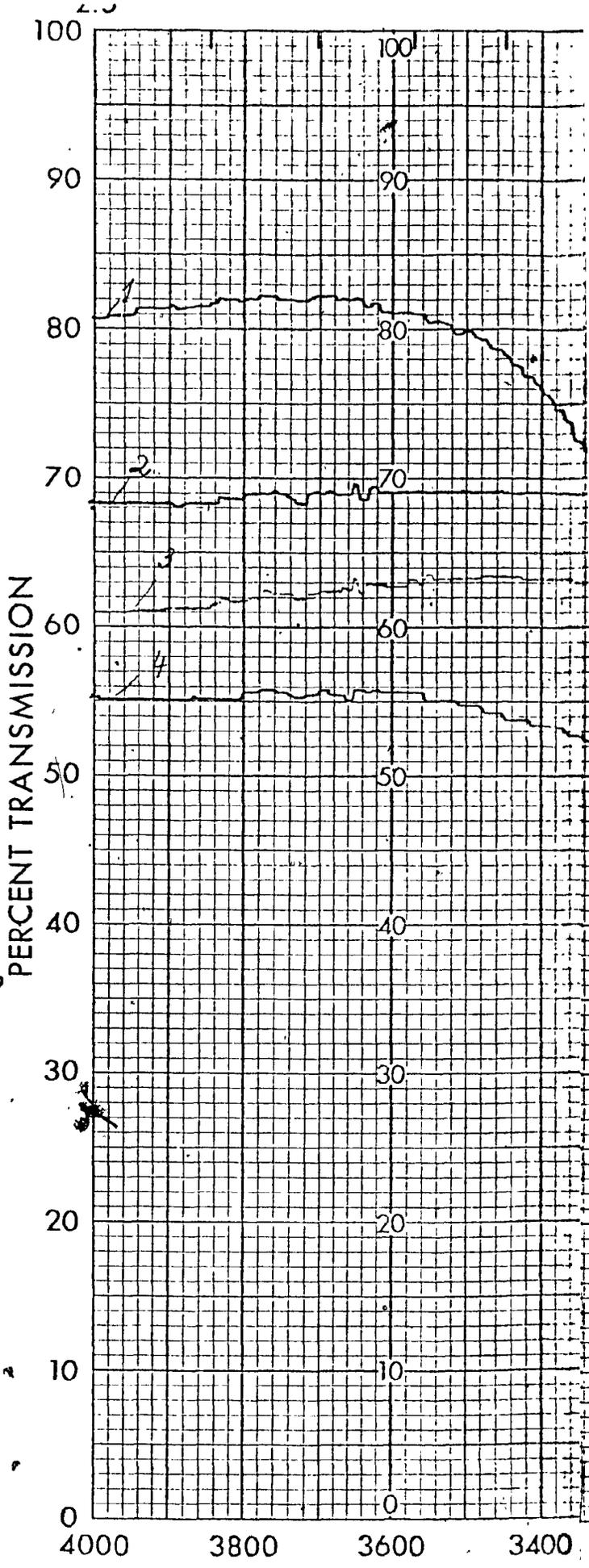
SPEED *600* cm<sup>-1</sup>/min.

GAIN *2*

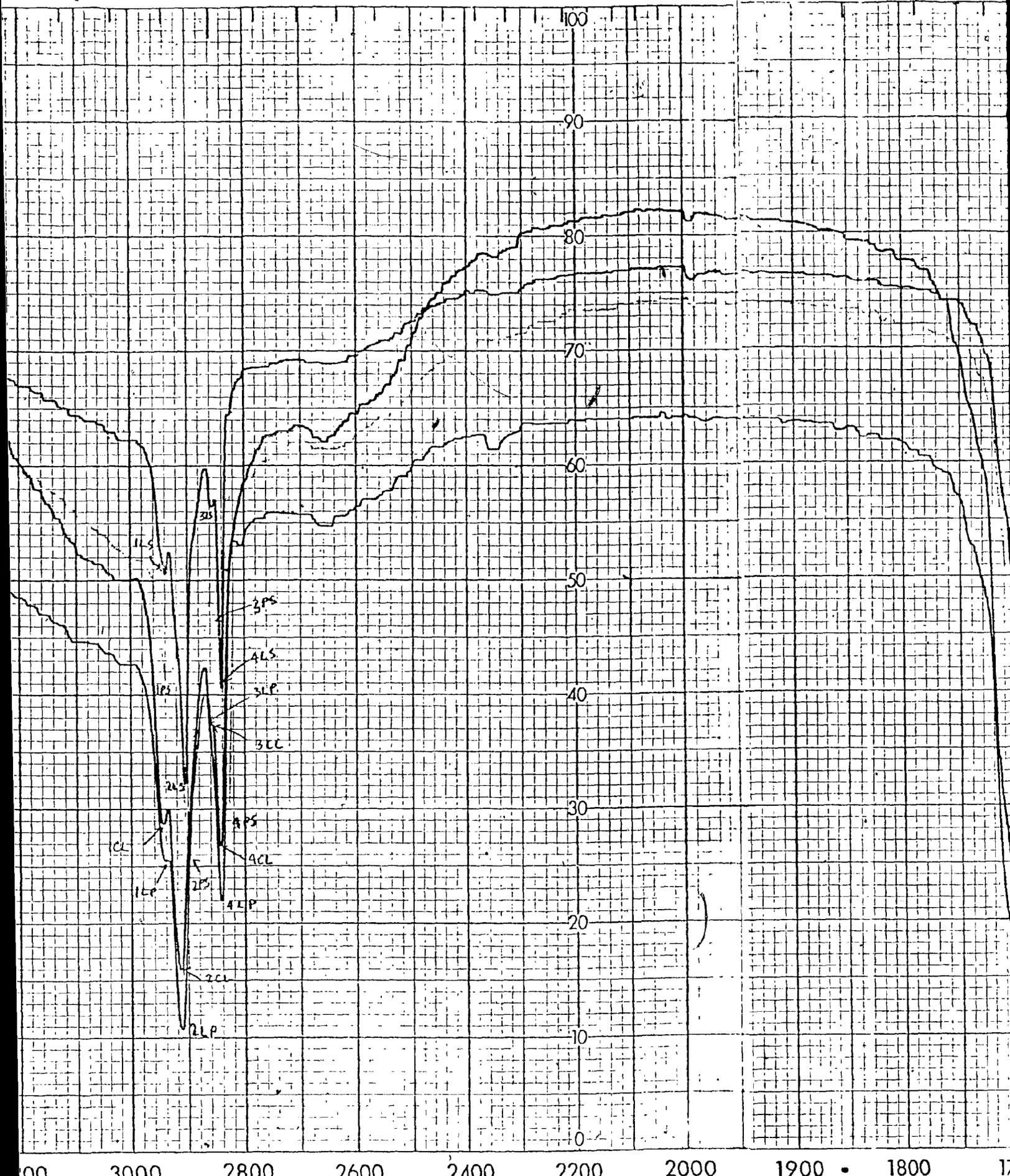
PERIOD *1* SEC.

SLIT WIDTH *3* mm

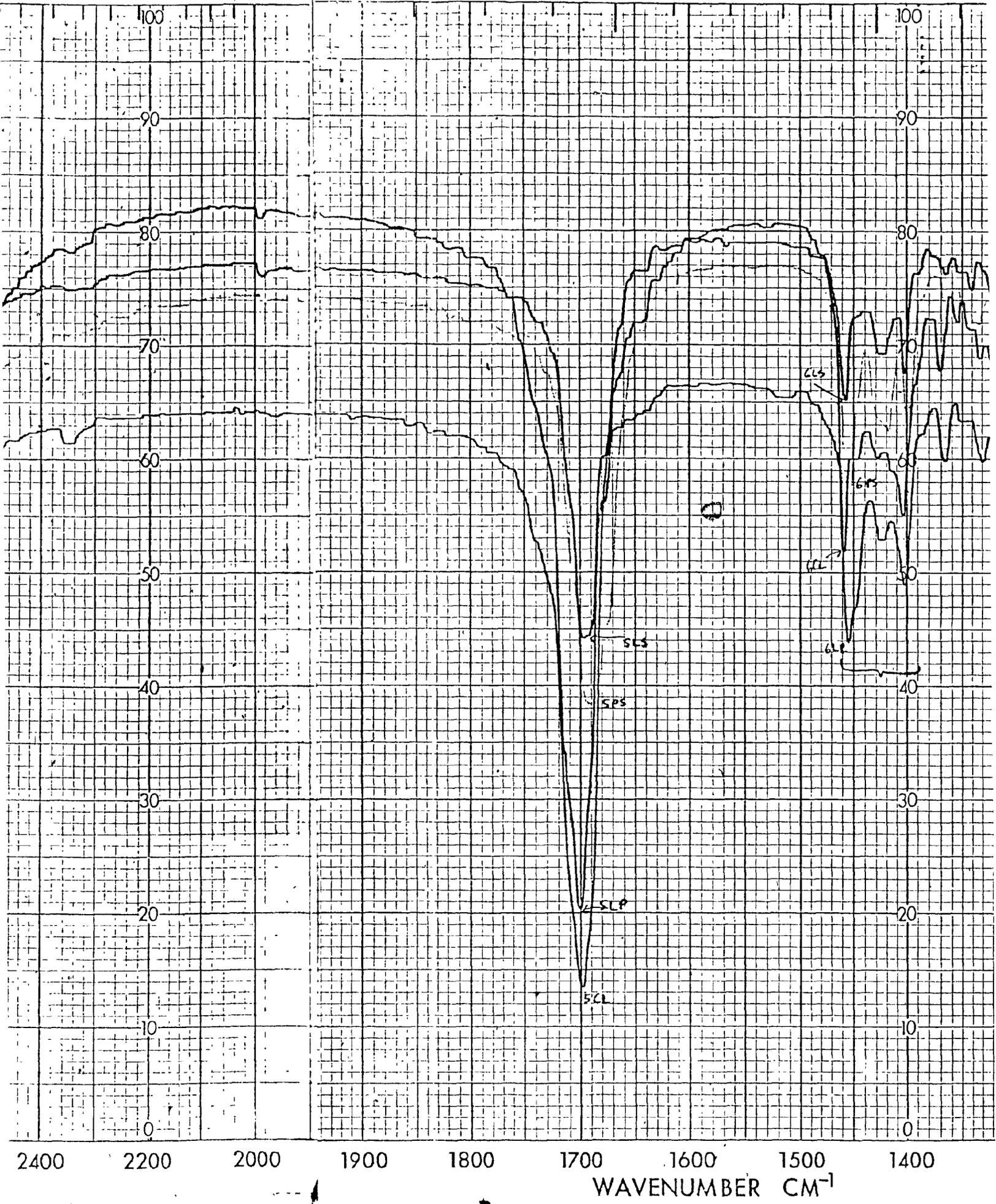
@ *3000* cm<sup>-1</sup>

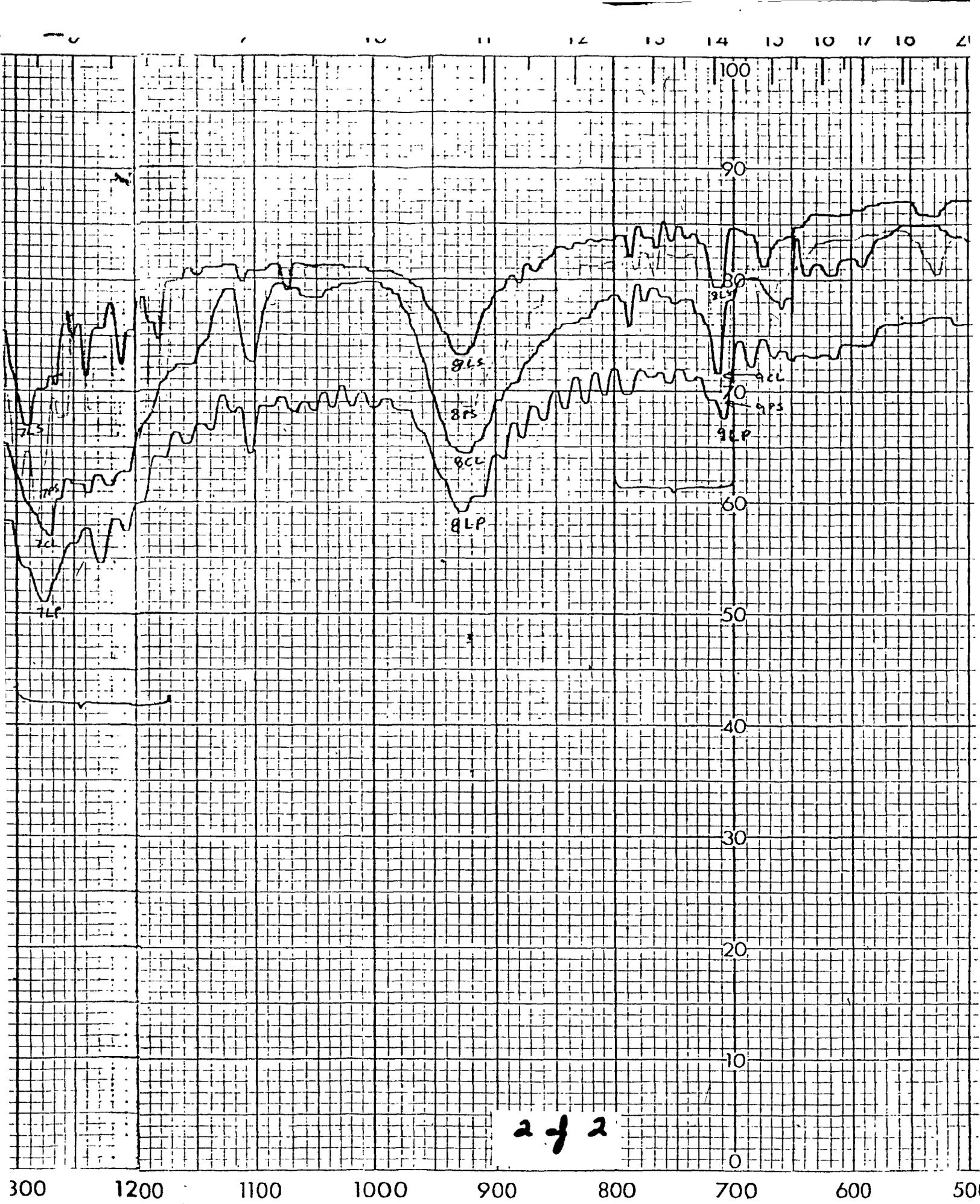


**BECKMAN**  
SPECTROPHOTOMETER



00 3000 2800 2600 2400 2200 2000 1900 • 1800 1700





2 of 2

300 1200 1100 1000 900 800 700 600 500

SPECTRUM NO. 107

DATE \_\_\_\_\_

SAMPLE 1. Copric Ac.  
2. Stearic Ac.  
3. Palmitic Ac.  
4. Lauric Ac.

SOURCE \_\_\_\_\_

STRUCTURE \_\_\_\_\_

PATH \_\_\_\_\_ mm

SOLVENT \_\_\_\_\_

CONC. \_\_\_\_\_

PHASE \_\_\_\_\_

COMMENTS KBr disk  
10mg/1000mg

ANALYST \_\_\_\_\_

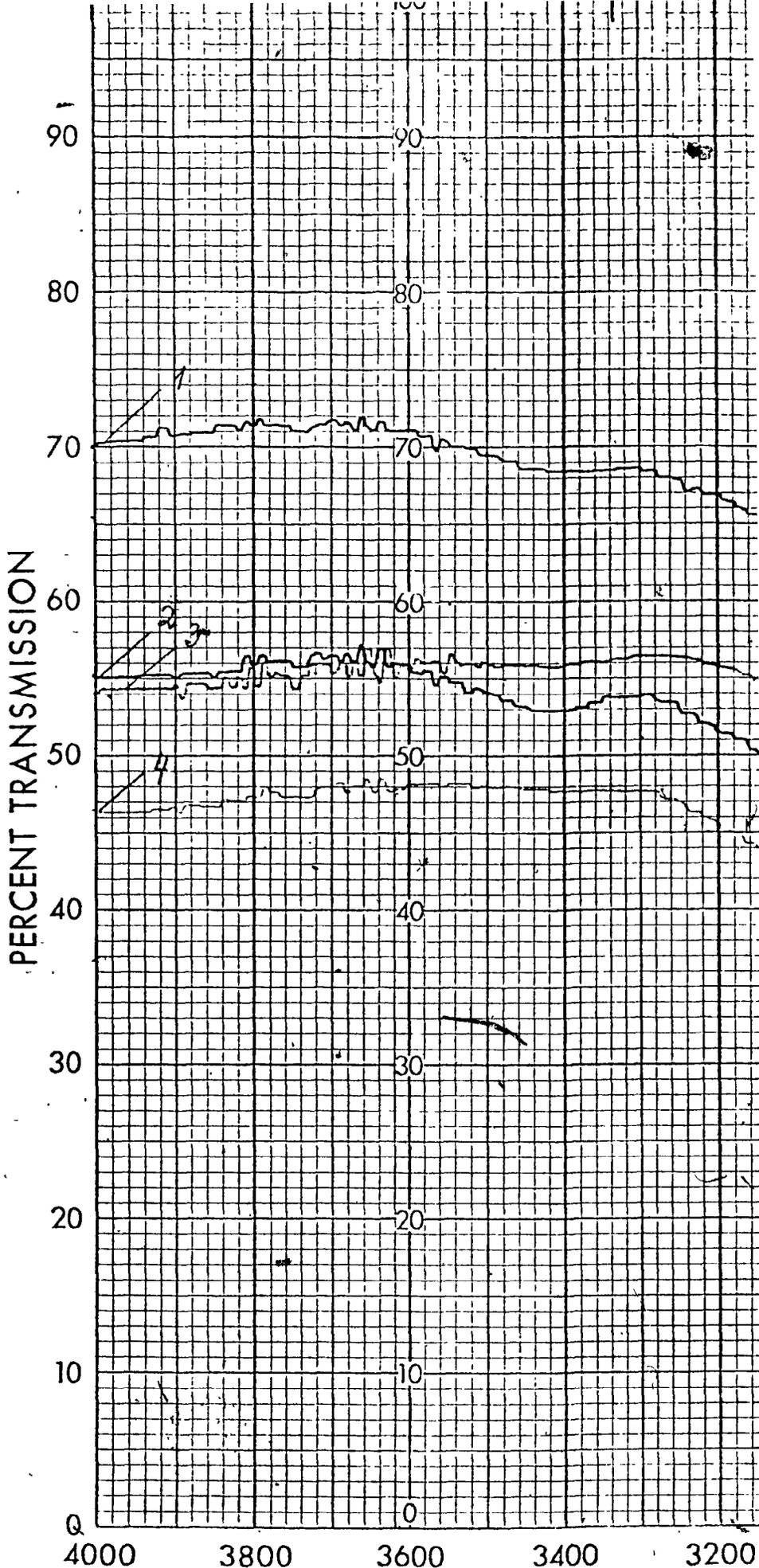
SPEED 600 cm<sup>-1</sup>/min.

GAIN 2

PERIOD 1 SEC.

SLIT WIDTH 3 mm

@ 3000 cm<sup>-1</sup>



**BECKMAN**  
SPECTROPHOTOMETER

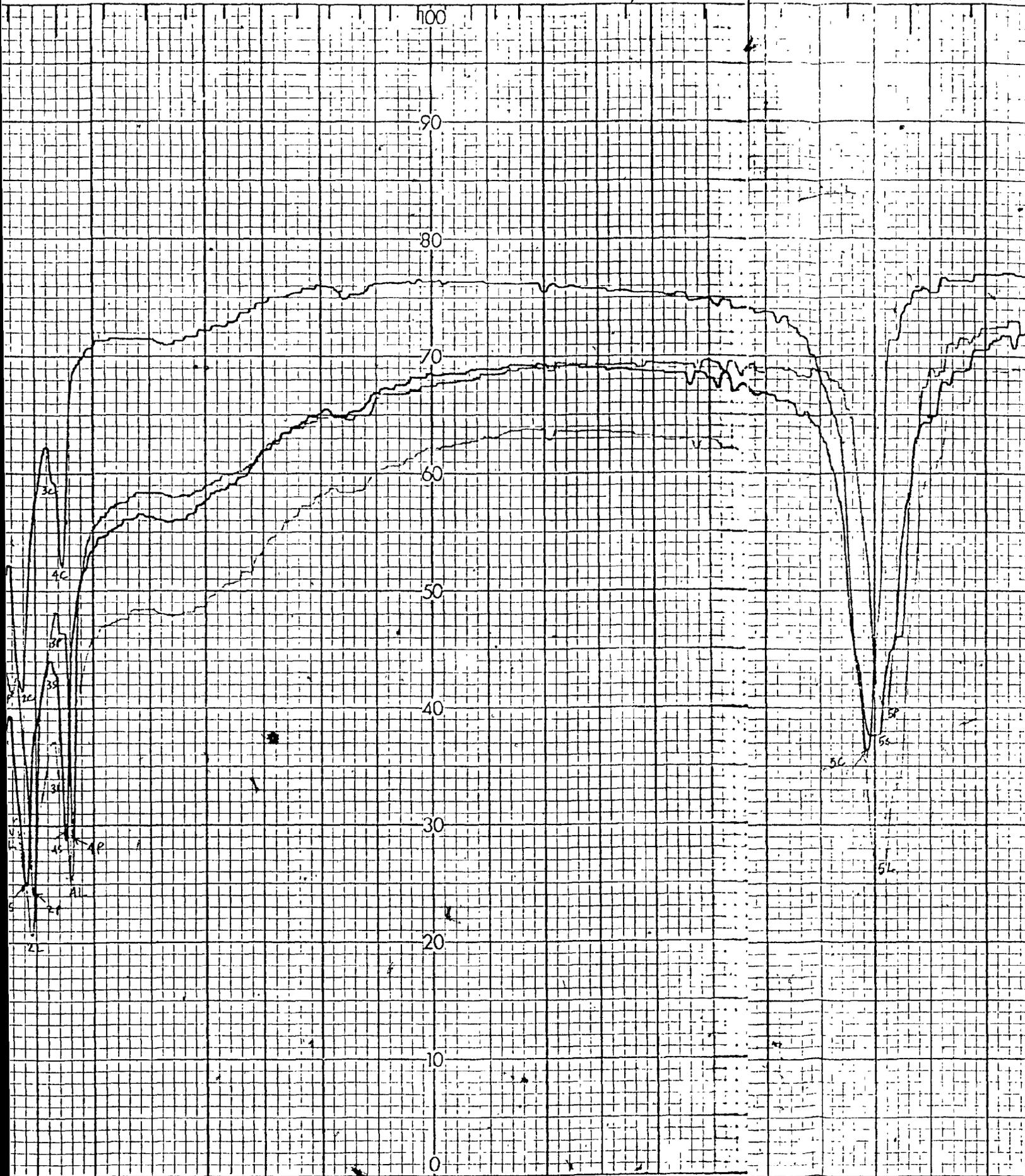
3.5

4

5

WAVELLENGTH μM

6



2800

2600

2400

2200

2000

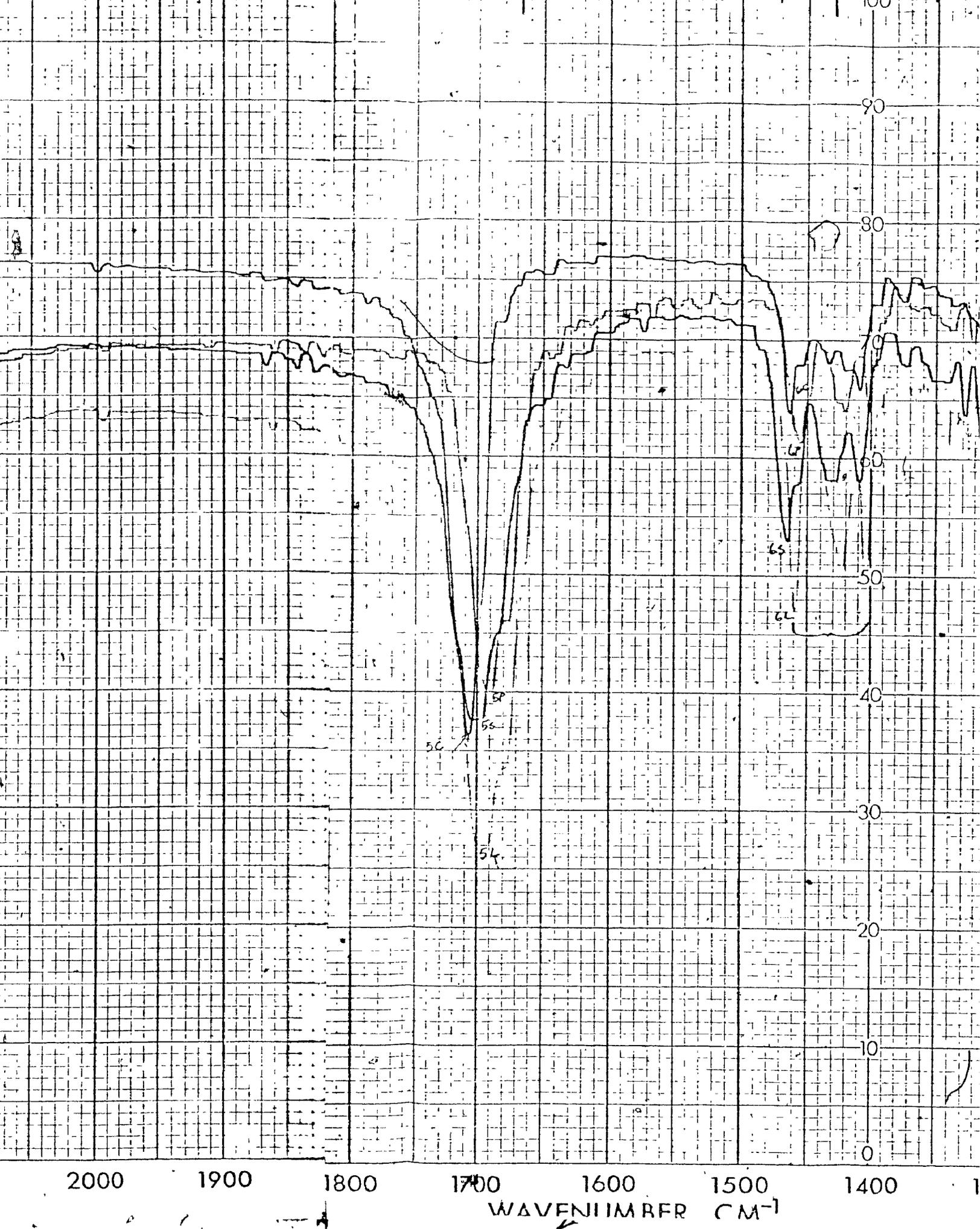
1900

1800

1700

1600

WAVELLENGTH μM



7

10

11

12

13

14

15

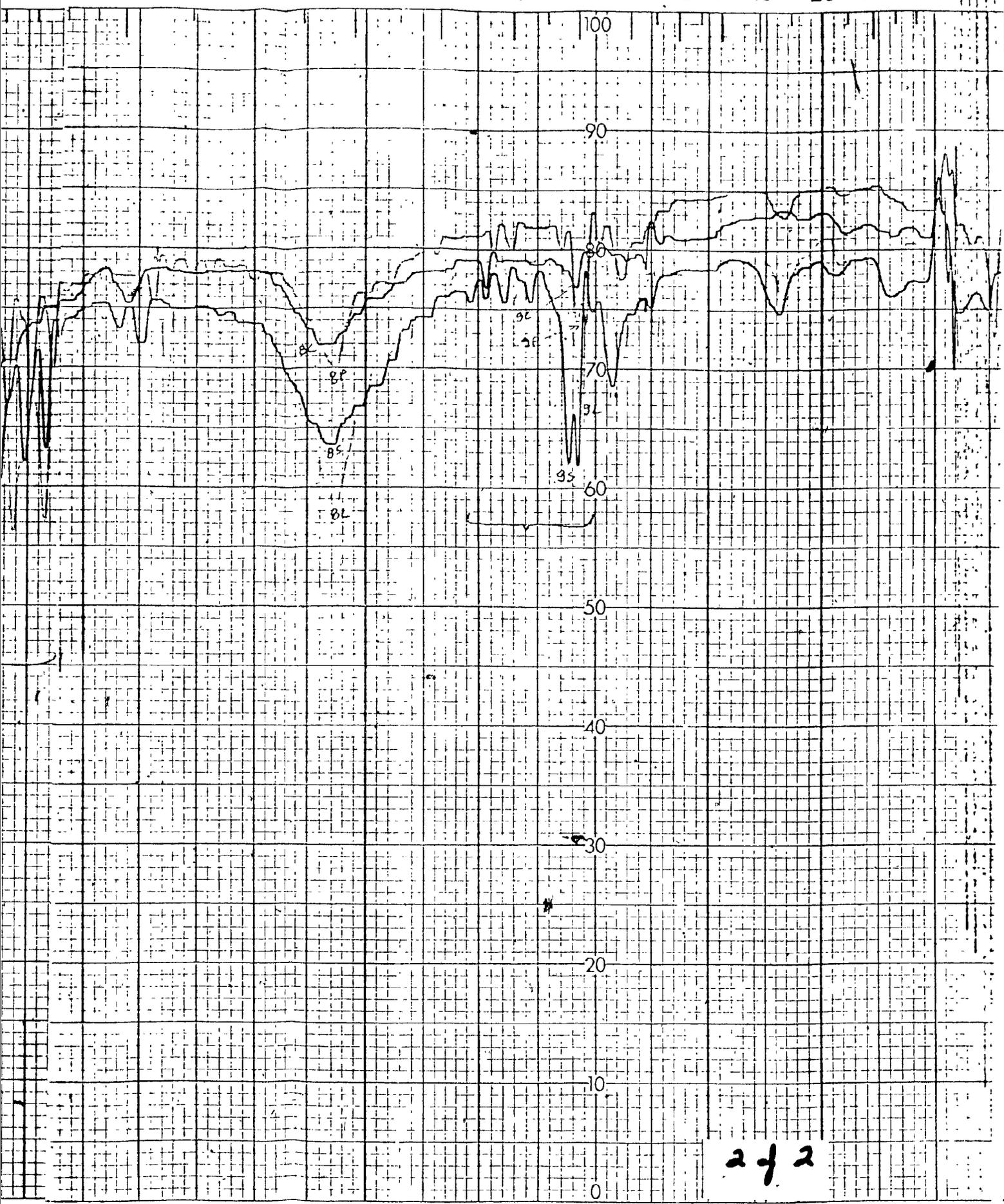
16

18

20

22

24



2 of 2

1200 1100 1000 900 800 700 600 500 400

