INHIBITING DOPAMINE REUPTAKE BLOCKS THE INDUCTION OF LONG-TERM POTENTIATION AND DEPRESSION IN THE LATERAL ENTORHINAL CORTEX OF AWAKE RATS

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Running Head: Dopaminergic suppression of synaptic plasticity in the entorhinal cortex

Number of Text Pages: 18
Number of Figures: 2

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Acknowledgements: This research was funded by grants to D.A.C and C.A.C. from the Natural Sciences and Engineering Research Council of Canada.

Keywords: memory, olfaction, piriform, parahippocampal, ventral tegmental area, rat.
ABSTRACT

Synaptic plasticity in olfactory inputs to the lateral entorhinal cortex may result in lasting changes in the processing of olfactory stimuli. Changes in dopaminergic tone can have strong effects on basal evoked synaptic responses in the superficial layers of the entorhinal cortex, and the current study investigated whether dopamine may modulate the induction of long-term potentiation (LTP) and depression (LTD) in piriform cortex inputs to layer II of the lateral entorhinal cortex in awake rats. Groups of animals were pretreated with either saline or the selective dopamine reuptake inhibitor GBR12909 prior to low or high frequency stimulation to induce LTD or LTP. In saline-treated groups, synaptic responses were potentiated to 122.4 ±6.4% of baseline levels following LTP induction, and were reduced to 84.5 ±4.9% following induction of LTD. Changes in synaptic responses were maintained for up to 60 minutes and returned to baseline levels within 24 hours. In contrast, induction of both LTP and LTD was blocked in rats pretreated with GBR12909. Dopaminergic suppression of synaptic plasticity in the entorhinal cortex may serve to restrain activity-dependent plasticity during reward-relevant behavioral states or during processing of novel stimuli.
The entorhinal cortex provides an interface between cortical association areas and the hippocampus and is involved in the formation of olfactory memory. Monosynaptic projections from the olfactory bulb and piriform cortex carry olfactory information directly to the entorhinal cortex [6], and lesions of the parahippocampal region that include the entorhinal cortex can produce deficits on olfactory tasks involving odor discrimination [30], delayed non-matching-to-sample performance [29, 35], and social recognition [3]. The entorhinal region, therefore, may make important contributions to olfactory memory, and persistent changes in synaptic strength in the entorhinal cortex may provide a mechanism for the modification of processing of olfactory information [5, 10, 12, 21].

Midbrain dopamine neurons of the ventral tegmental area and substantia nigra project to the superficial layers of the entorhinal cortex [4, 15, 27] and are well-poised to modulate responses to olfactory inputs that also terminate in these layers. Indeed, our recent work has demonstrated that increasing dopaminergic tone in the entorhinal cortex can facilitate synaptic responses to olfactory inputs in the superficial layers in vitro at low doses via D$_1$ receptors, and can suppress responses at higher concentrations mainly via D$_2$ receptors [8]. However, the effect of dopamine on lasting forms of synaptic plasticity in olfactory afferents to the entorhinal cortex is not known. Dopamine can have strong modulatory effects on both long-term potentiation (LTP) and depression (LTD) in cortical regions. In the hippocampus, dopamine facilitates the induction of LTD [11] and is also required for the long-term maintenance of LTP [37]. Similarly, dopamine facilitates both LTD [28] and LTP [20] in the prefrontal cortex. In most reports the enhanced plasticity in these areas has been linked to activation of D$_1$-like receptors [19]. Although dopamine typically facilitates both LTP and LTD, it can also inhibit plasticity; D$_1$ receptor activation can prevent LTP of the population spike in the dentate gyrus [41] and block maintenance of LTD in the CA1 region [25].

To determine the effects of dopamine on the induction and maintenance of synaptic plasticity
in the lateral entorhinal cortex, rats were pretreated with either saline or the selective dopamine reuptake inhibitor GBR12909 and field excitatory postsynaptic potentials (fEPSPs) evoked by stimulation of the piriform cortex were recorded before and after low or high frequency stimulation to induce LTD or LTP. We have shown previously that systemic administration of GBR12909 enhances extracellular levels of dopamine in the lateral entorhinal cortex in vivo [8] and the chronic field potential recording techniques used here provide a way to assess the effects of dopamine on synaptic plasticity in the awake rat.

MATERIALS AND METHODS

Experiments adhered to the guidelines of the Canadian Council on Animal Care, and surgical procedures were conducted according to methods described previously [8]. Briefly, male Long-Evans rats (300 to 350g; n =34) were anesthetized with sodium pentobarbital (65mg/kg, i.p.), and placed in a stereotaxic frame. A bipolar stimulating electrode was lowered into the right piriform cortex (P, 3.6mm; L, 6.5mm; V, 9.0mm relative to bregma), and a bipolar recording electrode was lowered into the superficial layers of the lateral entorhinal cortex (P, 6.5mm; L, 6.5mm; V, 7.5 to 8.5mm). Vertical placements were adjusted to optimize evoked responses. A stainless-steel screw in the contralateral frontal bone served as a reference electrode, and a screw in the occipital bone served as ground. Electrode leads were mounted in a connector and the assembly was embedded in dental cement.

Biphasic constant current square-wave pulses (0.1ms) were delivered via a stimulus isolator (A-M Systems, Model 2200) using a computer DAC channel or pulse generator (AMPI, Master 8 or A-M Systems, Model 2100). Evoked responses were filtered (0.1Hz to 5kHz passband), amplified (A-M Systems, Model 1700), and digitized at 20kHz for storage on computer hard disk (Datawave Tech.).
Animals were placed in a 40x40x60cm Plexiglas chamber inside a Faraday cage, and recordings were obtained after animals had habituated. Stability of responses was assessed using input/output tests every 2 days over a 5-day baseline period. During each test, 10 responses to stimulation of the piriform cortex were recorded and averaged at each of 6 intensities (0 to 1000µA) using a 10 sec inter-trial interval.

Following the final baseline input/output test, animals were injected with either the selective dopamine reuptake inhibitor GBR12909 (10mg/kg, i.p.; Sigma) or physiological saline (0.9%, 1ml/kg, i.p.) followed by another input/output test 20 min later. GBR12909 was prepared fresh by dilution in distilled water. Stability of responses was monitored over a 20 min baseline period prior to high frequency stimulation to induce LTP. During the baseline period, single stimulation pulses were delivered every 30 sec at an intensity set to evoke responses ≈50% of maximal. To induce LTP, ten high-frequency stimulation trains (16 pulses at 400Hz) were delivered every 2 min [10]. Post-tetanic effects were assessed during the 2 min inter-train intervals by delivering single pulses every 10 sec. Responses following LTP induction were monitored every 30 sec for a 60-min follow-up period, and input/output tests were administered 1 hour, and 1, 3, and 5 days post-tetanization.

Procedures to induce LTD were similar. Animals were pretreated with saline or GBR12909, and synaptic responses were monitored during the baseline period by delivering single stimulation pulses at an intensity that evoked responses ≈75% of maximal. Low frequency stimulation to induce LTD consisted of 900 pairs of stimulation pulses (30ms inter-pulse interval) delivered at 1Hz over a 15 min period [5].

Electrode placements were verified by light microscopy [8] and showed stimulating electrodes in the piriform cortex, and recording electrodes in superficial layers (I to III) of the lateral entorhinal cortex (not shown). Peak amplitudes of evoked fEPSPs were measured relative to the prestimulus
baseline [5, 10] and responses evoked during each input/output test were normalized to responses obtained at the highest stimulation intensity during the last baseline test. Field EPSPs recorded during LTP and LTD induction were normalized to the mean of responses obtained during the baseline period. Repeated measures ANOVAs compared averaged responses during the baseline period to responses evoked during the first and last 10 min of the 60-min follow-up period. Repeated measures ANOVAs also compared pre- and post-induction input/output tests.

RESULTS

Pretreatment with GBR12909 blocked the induction of LTP in piriform cortex inputs to the lateral entorhinal cortex (Fig. 1). Baseline fEPSPs had onset and peak latencies of 5.0 ±0.4 and 12.6 ±0.4ms and a mean peak amplitude of 1.03 ±0.14mV (e.g. Fig. 1A). High frequency stimulation potentiated synaptic responses in animals pretreated with saline (F2,16 =7.75, P<0.01; n =9) but did not significantly affect responses in animals pretreated with GBR12909 (n =8). The amplitudes of synaptic responses in saline-treated rats increased to 122.4 ±6.4% of baseline levels during the first 10 min following tetanization (Tukey, P <0.05) and were maintained at 125.1 ±7.0% of baseline during the last 10 min of the 60-min follow-up period (P<0.01). In contrast, responses in GBR12909-treated rats were stable and remained at 105.0 ±4.1% and 97.8 ±3.0% of baseline levels during the first and last 10 min of the follow-up period. Synaptic responses remained potentiated in saline-treated rats during the first follow-up input/output test (F5,40 =4.08, P<0.01; Fig. 1C) but responses decayed to baseline levels within 24 hours (not shown).

To determine whether pretreatment with GBR12909 influenced post-tetanic potentiation in the 2-min periods after each train, amplitudes of the first responses evoked after each of the 10 trains were compared between saline- and GBR12909-treated rats (Fig. 1B; black bar). Significant post-tetanic potentiation was evidenced in both groups by a decay in the amplitude of responses
during the 2-min inter-train intervals ($F_{11,165} = 5.91, P < 0.001$; not shown) but there was no significant difference between groups in the responses evoked immediately following each train. The development of LTP in saline-treated rats, however, resulted in larger overall averaged responses during the 2-min inter-train intervals ($F_{1,15} = 5.45, P < 0.05$; not shown).

Pretreatment with GBR12909 also blocked induction of LTD (Fig. 2). Low frequency paired-pulse stimulation depressed synaptic responses in saline-treated rats ($F_{2,18} = 5.87, P < 0.05; n = 10$) but had no significant effect on fEPSPs in animals pretreated with GBR12909 ($n = 7$). In control animals, responses were significantly reduced to $84.5 \pm 4.9\%$ of baseline levels during the first 10 min (Tukey, $P < 0.05$) and remained depressed at $82.8 \pm 6.8\%$ of baseline after 60 min ($P < 0.05$). In contrast, responses in GBR12909-treated animals remained stable at $94.7 \pm 5.9\%$ and $101.2 \pm 5.0\%$ of baseline after 10 and 60 min, respectively. Synaptic responses remained depressed during the first follow-up input/output test in saline-treated rats ($F_{5,45} = 2.30, P < 0.05$; Fig. 2C) but returned to baseline levels within 24 hours (not shown). Responses to conditioning pulses tended to be larger in control animals during LTD induction, but this difference was not significant (Fig. 2B, black bar), and GBR12909 also did not significantly affect the amount of paired-pulse facilitation during LTD induction.

DISCUSSION

We have found here that dopamine has a suppressive effect on the induction of both long-term potentiation and depression in olfactory inputs to the lateral entorhinal cortex of awake rats. Animals pretreated with saline showed levels of LTP and LTD that were comparable to previous reports that used similar stimulation protocols [5, 10, 21]. However, increasing dopamine levels in the entorhinal cortex with the selective dopamine reuptake inhibitor GBR12909 suppressed the induction of both LTP and LTD. In contrast to reports that have shown a facilitatory effect of dopamine on LTP and LTD in the hippocampus [11, 37] and prefrontal cortex [20, 28], the
suppression observed here suggests that synaptic plasticity is normally dampened in the entorhinal cortex during behaviors associated with increased activity in dopaminergic inputs. We have shown previously that dopamine has concentration-dependent biphasic effects on basal synaptic transmission in the entorhinal cortex; although high concentrations suppressed synaptic transmission, responses were facilitated by a lower concentration of dopamine [8]. This, together with our current findings, suggests that moderate elevations in extracellular dopamine may promote transmission of olfactory patterns into the hippocampus, while simultaneously limiting activity-dependent synaptic modifications in the entorhinal cortex. Although increased dopamine might be expected to enhance learning-related plasticity, a dopaminergic suppression of plasticity might be useful during periods of increased network excitability to prevent excessive changes in synaptic strength, and to maintain stable processing of physiologically relevant olfactory signals. Of course, the systemic injections used here may have elevated dopamine concentrations beyond physiologically relevant levels, and further work with moderate, temporally controlled elevations in cortical dopamine both in vivo [e.g., 20] and in vitro [7] is necessary.

Systemic administration of GBR12909 could have enhanced dopamine availability in terminal regions throughout the brain, but the suppression of plasticity observed here was likely due to effects of GBR12909 within the entorhinal cortex. Olfactory inputs from the piriform cortex terminate in the superficial layers of the lateral entorhinal cortex [6] where fibers from midbrain dopamine neurons also terminate [4, 15, 27]. Further, we previously monitored the effects of GBR12909 at the dose used here with in vivo microdialysis, and found that extracellular levels of dopamine in the entorhinal cortex were increased to 306% of basal levels [8]. However, GBR12909 injections could also have resulted in increased acetylcholine and/or serotonin in the entorhinal cortex. GBR12909 increases locomotor activity in rats [8, 26] that is associated with cholinergic-dependent theta activity in the entorhinal cortex [24] and, although cholinergic activation can
promote synaptic plasticity in some areas, we have recently found that muscarinic receptor
activation suppresses glutamatergic transmission in the entorhinal cortex [18]. This suppression
could have contributed to the block of LTP and LTD shown here. Similarly, systemic
administration of GBR12909 can increase firing of raphé neurons [23], and serotonin inhibits
synaptic transmission in the lateral entorhinal cortex in vitro [17]. Thus, although the block of
synaptic plasticity observed here was likely due primarily to the effects of increased dopamine
levels on local entorhinal circuitry [8], effects of systemic injections are always difficult to interpret,
and the current results will have to be extended using in vitro recordings.

The induction of both LTP and LTD in the entorhinal cortex is dependent on NMDA
receptors [1, 14, 21], and it is likely that GBR12909 may have interfered with plasticity by reducing
postsynaptic depolarization required for NMDA receptor activation. We showed previously that low
concentrations of dopamine facilitate synaptic responses in the lateral entorhinal cortex [8], and this
suggested to us that dopamine might enhance the induction of LTP. However, although 10µM
dopamine facilitates responses via a D₁ receptor-mediated mechanism, higher concentrations of 50
and 100µM suppress AMPA and NMDA responses via a D₂-mediated reduction in glutamate
release [7, 8]. A suppression of transmitter release could help block plasticity by reducing
postsynaptic depolarization during stimulation trains. In the present study, pretreatment with
GBR12909 did not significantly enhance basal responses, and this could be due to a D₂-mediated
suppression of transmitter release [7, 8, 31, 36]. Strong activation of D₁ receptors could also have
reduced levels of postsynaptic depolarization during trains [8]; while D₁ receptors mediate the
facilitation of responses at low-concentrations of dopamine, they also contribute to the suppression
of EPSPs observed at higher concentrations. This is similar to the inverted U-shaped relationship
described for prefrontal cortex responses [2, 16]. We cannot know what the effective concentration
of dopamine was during LTP and LTD induction in the present groups of animals, but both D₁ and
D₂ receptor activation may have contributed to the block of plasticity observed here. In the prefrontal cortex and hippocampus, dopamine typically enhances plasticity through an intracellular signal cascade involving a D₁ receptor-mediated increase in cAMP via activation of adenylate cyclase and resultant activation of PKA [19]. However, D₁ receptor activation can also inhibit NMDA-mediated synaptic currents in cultured hippocampal neurons through a direct coupling of D₁ receptors to NMDA receptors [9], and selective D₄ receptor activation in prefrontal cortex slices and cultures can also suppress NMDA receptor currents [40]. D₄ receptors are also present in the entorhinal cortex [13, 32, 39] and a D₄-mediated suppression of NMDA receptor currents may have blocked the induction of LTP and LTD in the current study.

Relative to control animals, there was a non-significant reduction in responses evoked following each high-frequency train in GBR12909-treated rats (Fig. 1B), as well as a non-significant reduction in responses to conditioning pulses during low frequency paired-pulse stimulation to induce LTD (Fig. 2B). In layer V neurons of the lateral entorhinal cortex dopamine increases the Iₕ current, and this reduces postsynaptic excitability during repetitive synaptic stimulation by reducing temporal summation of EPSPs [33]. Layer II neurons also show inward rectification that reflects Iₕ [7, 34, 38] but we have found that dopamine significantly reduces Iₕ in layer II neurons rather than enhancing it [7]. It is not yet clear, then, if these effects on temporal summation may be expressed in the superficial layers.

We have used systemic injections here, but the precise timing of dopamine application relative to LTP or LTD induction has been a critical factor in other studies. Plasticity is generally enhanced when transient D₁ receptor activation occurs before or during stimulation [11, 20, 28, 37] but the maintenance of LTD is blocked when D₁ receptors are activated shortly after the trains [25]. Multiple intracellular mechanisms are likely to have been activated by the injections used here, and
it is unknown if similar time-dependent effects control the modulatory actions of dopamine in the entorhinal cortex.

Activation of midbrain dopamine neurons during appetitive behaviors is likely to have complex effects on the processing and encoding of olfactory representations by the entorhinal cortex. We found previously that dopamine has bidirectional effects on synaptic transmission via D<sub>1</sub> and D<sub>2</sub> receptors [8], and we have shown here that enhancing dopaminergic tone with GBR12909 blocks the induction of LTP and LTD in the lateral entorhinal cortex of awake animals. In the hippocampus, dopamine efflux is triggered in response to novelty, and it has been suggested recently that this can enhance the encoding of new information within CA3 projections to the CA1 region [22]. Thus, enhanced basal transmission in the entorhinal cortex could promote transfer of sensory information into the hippocampus and enhance the integration of this information into elaborated representations carried by the CA3 and CA1 regions [8, 22, 37]. At the same time, the inhibitory effect of dopamine on LTP and LTD that we have observed here suggests that dopamine may protect the entorhinal cortex from plasticity that could follow from increased neuronal activity during intense sensory processing, and may also shift the site of plasticity in novel or reward-relevant situations to the hippocampal region [22].
REFERENCES


FIGURE CAPTIONS

**Figure 1.** Enhancing extracellular dopamine with GBR12909 blocks the induction of long-term potentiation in olfactory inputs to the lateral entorhinal cortex. **A:** Representative traces from rats pretreated with saline (A₁) or GBR12909 (A₂) before or after high frequency stimulation to induce LTP. Numbered traces in A correspond to time points indicated in B. Note the potentiation observed in the saline-treated animal (A₁; 1+2) but not the GBR12909-treated rat (A₂; 1+2). **B:** Mean response amplitudes (±SEM) recorded before, during, and after high-frequency stimulation trains in rats pretreated with saline (open circles) or GBR12909 (filled circles). Amplitudes of fEPSPs were expressed as a percentage of the entire baseline period and averaged every 5-min for plotting. Averaged responses recorded immediately following each stimulation train showed no significant difference between groups in post-tetanic potentiation (black bar). **C:** Synaptic responses remained potentiated for 60 min in saline-treated rats (C₁) and were stable in rats pre-treated with GBR12909 (C₂). Responses in C are expressed as a percentage of responses to the highest stimulation intensity during the last baseline test.
Figure 2. Pretreatment with GBR12909 blocks induction of long-term depression in the lateral entorhinal cortex. A: Representative fEPSPs from saline- (A₁) and GBR12909-treated (A₂) rats were recorded before and after repetitive low frequency paired-pulse stimulation to induce LTD. Depression of the fEPSP was observed in the control animal (A₁), but not in the animal pretreated with GBR12909 (A₂). B: Mean fEPSP amplitudes before, during, and after low frequency stimulation in saline- (open circles) and GBR12909-treated (filled circles) animals. Amplitudes of responses to conditioning pulses during repetitive paired-pulse low frequency stimulation (PP LFS, black bar) were not significantly different between groups. C: LTD was maintained for 60 min in saline-treated animals in response to the highest stimulation intensities (C₁). Amplitudes of fEPSPs remained stable in animals pretreated with GBR12909 (C₂).