

**Evolutionary based Classification of Fungal Lipases as a Framework for Structure
and Function Prediction of Putative Lipases.**

Bahareh Behdad

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ABSTRACT

Evolutionary based Classification of Fungal Lipases as a Framework for Structure and Function Prediction of Putative Lipases.

Bahareh Behdad

Lipases are a family of enzymes which catalyze the hydrolysis of lipids and they exhibit esterase type activities such as phospholipase, lysophospholipase, cutinase and amidase. These enzymes have applications in detergent, the production of oil and fat, baked products, organic synthesis, hard surface cleaning, leather, paper, and cocoa butter. Lipases are versatile and have become a major source of industrial lipolytic enzymes. I performed a comparative analysis of protein sequences of well characterized fungal lipolytic enzymes from SWISS-PROT and developed a classification system to assist structural and functional characterization of newly identified putative lipase gene sequences. The distance-based UPGMA method was used for constructing the phylogenetic tree. Eight sequences of known secondary and tertiary structure were used to predict the secondary structure of similar sequences with unknown structure. Using sixteen different tools from SWISS-PROT and NPS@, the secondary structures of sequences with unknown 3-dimensional structures were predicted. The evolutionary based clustering of lipase protein sequences resulted in seven major families with the largest family being divided into five subfamilies and two single member branches. This analysis allowed us to: (i) perform a comparative study of well-characterized fungal lipases to develop a comprehensive classification system for fungal lipases, (ii) assess

various protein secondary structure prediction tools to select suitable tool(s) for predicting secondary structures of lipases, (iii) predict structural features important for specific function of lipases such as residues forming the catalytic sites, disulfide bonds and salt bridges, and (iv) develop a framework to predict putative function and reaction conditions of newly identified lipase gene sequences.

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DEDICATION

I would like to dedicate this thesis to my loving parents...

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A. INTRODUCTION

Lipases, a group of enzymes which catalyze hydrolysis of insoluble oil droplets into soluble products, are ubiquitous in nature. They are found in organisms ranging from bacteria and fungi to plants and animals. These enzymes aid in processes such as digestion, membrane phospholipid metabolism, and inflammatory reactions (Tojo *et al.*, 1997). Moreover, these enzymes are commercially important in industries such as detergent, oil and fat, baking, organic synthesis, paper and cocoa butter (Schmid and Verger, 1998; Bornscheuer and Kazlauskas, 1999; Anderson *et al.*, 1998; Clausen *et al.*, 2000; Rubingh, 1998; Jaeger, 1998). Fungal lipases have become a major source of industrial lipolytic enzymes because of their versatility and ease of production. Such new found uses for fungal lipases have spurred growth in patent filings, driving the need for an improved understanding of the structure-function relationships of fungal lipases.

Fungi produce various families of lipolytic enzymes including true lipases (EC 3.1.1.3), carboxylesterases (EC 3.1.1.3), secretory lipases (EC 3.1.1.3), and a variety of phospholipases, namely, phospholipase A1 (EC 3.1.1.32), phospholipase A2 (EC 3.1.1.4), lysophospholipase (EC 3.1.1.5), phospholipase C or 1-phosphatidylinositol-4, 5-bisphosphate phosphodiesterase 1 (EC 3.1.4.11), and phospholipase D (EC 3.1.4.4). The first three classes of enzymes belong to the structure-based super family of α/β -hydrolase's (Ollis *et al.*, 1992), a variety of enzymes whose activities rely mainly on a catalytic triad usually formed by serine, histidine and aspartic acid residues.

Three developments have advanced our knowledge of the structure and function of lipases and esterases: increased availability of gene sequences, biochemical

characterization of lipases, and resolution of numerous crystal structures (Grochulski *et al.*, 1993; Schrag and Cygler, 1993; Cygler *et al.*, 1993).

These classifications are based upon comparative studies of the structure and function of well-characterized enzyme groups. They aid in the identification of unknown structural motifs and also help predict the functions of newly-identified genes. Although a structure and function based classification of bacterial lipases already exists (Arpigny and Jaeger, 1999), no such study is yet available for fungal lipases.

Our objectives are to:

- (1) Perform a comparative study of well-characterized fungal lipases to develop a classification system for fungal lipases.
- (2) Select the best tools for predicting secondary structure of lipases.
- (3) Predict structural features important for function of lipases such as residues forming the catalytic sites, disulfide bonds and salt bridges.
- (4) Develop a framework to predict function and reaction conditions of newly-identified lipase gene sequences.

B. LITERATURE REVIEW

B.1 Background Information and Definition of lipases

Lipases were first identified in 1856 by Claude Bernard. They hydrolyse insoluble oil droplets into soluble products. The presence of lipolytic enzymes in blood plasma was first noted by Hahn in 1943. Initially, the enzyme was described as a “clearing factor”. Nine years later, Anfinsen and co-workers associated the clearing factor with the presence of lipolytic enzymes (Verger *et al.*, 1984). Since then, there has been a growing interest in both the lipolytic enzymes and their substrates. The lipase enzymes consist of different families showing the same overall structural folding (Ollis *et al.*, 1992; Derewenda *et al.*, 1994a). However, they differ in the versatility of their loop structures that contact the substrate. They also show variable substrate specificities. Lipases, also known as lipolytic enzymes, are capable of hydrolyzing lipid substrates: cutinases, lysophospholipases and enzymes hydrolyzing ester substrates of lipid nature. Lipases, in contrast to esterases, become activated when absorbed in a water/lipid interface. They display low activity with their substrates in a monomeric state (Verger and de Hass, 1976). A “true” lipolytic enzyme exhibits two characteristics (EC 3.1.1.3):

- It should be activated by the presence of an interface. That is, its activity should sharply increase as soon as the triglyceride substrate forms an emulsion (interfacial activation) (Sarda *et al.*, 1958).
- It should contain a “lid” (see below), which consists of hydrophobic residues covering the active site. By movement of the lid, the substrate can enter the cavity for enzyme reaction.

What make lipases so attractive? First, they usually display exquisite chemoselectivity, regioselectivity and stereoselectivity (Rogalska *et al.*, 1993). Secondly, they can readily be produced in high yields from microbial organisms-- fungi and bacteria. Thirdly, the crystal structures of many lipases have been solved, facilitating the design of rational engineering strategies. Finally, they usually do not require cofactors, and do not catalyze side reactions. These properties make lipases the most widely used group of biocatalysts in organic chemistry.

B. 2 Structural studies on lipases

To determine the structure of a protein it is necessary to understand its function and role. The mechanism of catalysis and the molecular nature of interfacial activation was not understood until the 3-D structures of lipases were determined. The lipases from two fungi-- *Aspergillus niger* (Fukumoto *et al.*, 1963) and *Geotrichum candidum* -- (Tsujiisaka *et al.*, 1973) were first to be crystallized. These crystals were unstable and of poor quality, likely a result of heterogeneity in the enzyme preparations. Since then, many lipases have been crystallized in a form suitable for high resolution X-ray diffraction studies.

Triacylglycerol lipases are α/β proteins, with a central β -sheet with the active site serine placed in a loop, in which the nucleophilic residue is essential for catalysis (Cygler *et al.*, 1992). This putative hydrolytic site is covered by a surface loop and is therefore, inaccessible to solvent. Interfacial activation, a property of lipolytic enzymes acting on water-insoluble substrates at water-lipid interfaces, probably involves the movement of this flap region in lipoprotein lipases. This movement changes the surface at the entrance

of the active site, making it more hydrophobic and changing the lipid-binding properties.

The activities of these enzymes rely on a catalytic triad usually formed by three residues following the order Ser-Asp/Glu-His. The active site serine is found in the consensus pentapeptide Gly-X-Ser-X-Gly (X being any amino acid). This sole presence of this motif has identified many serine hydrolases. The active site serine is embedded in a secondary structure element: β -strand-turn- α -helix. This general element was recognized in previous studies (Derewenda and Derewenda, 1991; Schrag *et al.*, 1991). It was found not only in lipases, but also in other hydrolytic enzymes (Schrag *et al.*, 1991; Ollis *et al.*, 1992).

The two glycine residues of the Gly-X-Ser-X-Gly consensus sequence are critical in maintaining the tight bend between the β -strand and the α -helix. These two residues face each other (Figure 1); the distance between their C $_{\alpha}$ atoms is very short (~ 4.5 Å). Figure 1 shows the comparison of *Rhizomucor miehei* lipase (RML) and *Geotrichum candidum* lipase (GCL), showing the conserved Gly-X-Ser-X-Gly and the short distance between the two Gly residues. Not only should the Gly residue be conserved, but also right after this Gly, there is always a small amino acid such as alanine or another glycine. This keeps the outlook or nature of the β -strand and α -helix unchanged (Cygler *et al.*, 1992). This conserved motif is seen in all known esterase structures: acetyl cholinesterase (Sussman *et al.*, 1991), cutinase (Nicolas *et al.*, 1996) *streptomyces scabies* esterase (Wei *et al.*, 1995).

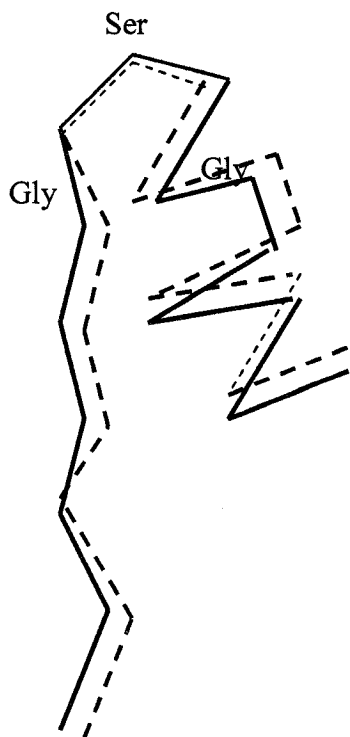


Figure 1: Superposition of strand-serine-helix super secondary motif. GCL (thick line) and RML are shown in dashed line. Red indicates the α -helix, and green indicates the β -strand.

Although the scaffolds and the active sites of lipases and cutinases share common features, accessibility to the active sites in each lipase differs. For example, in RML (*Rhizomucor miehei* lipase) the active site is covered by a short region (flap) and is about 10 Å from the protein surface. A similar structure was seen for human pancreatic lipase (HPL) (Terzyan *et al.*, 2000) with a longer loop and greater distance to the surface (about 14 Å). In fact, a different structure is seen for GCL, where two flap regions from different surfaces cover the active site. Yet no flap region is found in cutinase, and further, the active site is on the surface of the molecule (Martinez *et al.*, 1992). The histidine residue is often part of a special sequence pattern (Svendsen *et al.*, 1995) and mostly found in the C-terminal end of lipases. However, this residue in *Geotrichum candidum* lipase (GCL) comes from the N-terminal part of the sequence. In cutinases, the histidine is located in the same loop as its triad partner. Cutinases have a shorter C-terminus than the above

mentioned lipases (Martinez *et al.*, 1992). Moreover, the Asp/Glu residue would be found in the triacylglycerol lipases between the serine and the histidine residue.

In some lipase-related enzymes, the active site residues are arranged differently. In secretory lipases (discussed later in this study), there are only two catalytic residues, instead of three. Also, in the *Streptomyces scabies* bacteria esterase, the Asp residue is 'replaced' by carbonyl oxygen. Enzymes including lipases, proteases, esterases, peroxidases and lysases are members of the " α/β hydrolase fold" clan (Ollis *et al.*, 1992; Schrag *et al.*, 1997), although not all lipases belong to the same structural family. As an example, the structures of PLA₂, PLA-D, lysophospholipases and phospholipases C differ from triacylglycerol lipases.

B. 3 Function

Because lipases are water-soluble enzymes and function at interfaces, evaluation of substrate specificity is governed by several other factors (Brady *et al.*, 1990). Few lipases show strict substrate specificity. The mode of presentation of the substrate is also important (Brockman, 1984). The same enzyme may show different selectivities when the substrate is added in variable forms, like micelles, monolayer films or dissolved in organic solvents. Classically, types for specificities of lipases are classified by factors such as: a) substrates, i.e., preference for long or short fatty acids; b) positional: including sn1, sn2 and sn3 such that the regioselectivity is rather high for positions sn1 and sn3 so that sn2 is degraded rarely (Figure 2); c) stereospecificity: faster hydrolysis of one primary sn ester as compared to its counterpart and d) combination of all of the

specificities mentioned above (Benjamin and Pandey *et al.*, 1998; Svendsen, 2000).

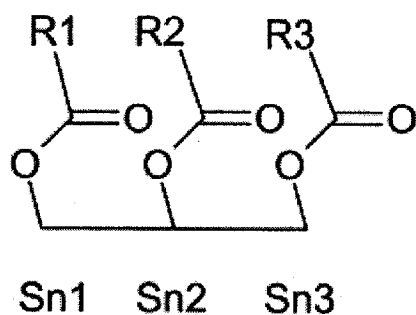


Figure 2: The triacylglycerol type of lipid presented using the *sn* designation for positions of the glycerol moiety (Adopted from Svendsen, 2000).

B. 4 Importance of lipases

Industrial and Biological applications

Lipases have evolved to be efficient catalysts for lipolytic reactions concerning the hydrolysis of ester linkages of mono-, di- and triglycerides in aqueous emulsions. The hydrolysis of these bonds is important in industrial applications:

- Generation of fatty acids from natural oils for the production of soaps
- Removal of oils and fats from fabrics, machinery, hides and waste water
- Production of mono- and diglycerides for food emulsifiers.

Lipases are also used to conduct transesterification reactions in commercial applications, such as the production of cocoa butter substitutes (Sharma *et al.*, 2001). Since lipases have the unique ability to act at oil-water interfaces, their structures are of particular interest. The selectivity of lipases towards the length of the fatty acids or the number and location of unstaurations in the fatty acids is employed to produce high-value fats or oils (Macrae *et al.*, 1983). The high stereoselectivity of lipases is also exploited to synthesize specific compounds-- in particular, enantiomerically pure compounds

containing ester bonds. For thermodynamic reasons, these synthesis reactions must be conducted in environments containing little water. There have been many reports of lipases in organic solvents synthesizing a variety of compounds (Boland *et al.*, 1991), including precursors for biologically active therapeutics, herbicides, and pesticides.

Though lipolytic enzymes are widely distributed in the plant kingdom, knowledge of lipases from plants is limited when compared with those from mammalian systems and micro-organisms. The low abundance of these proteins makes it difficult to purify them in amounts sufficient to get access to amino acid sequence information. During post-germination of oil seed plant (eg. *Arabidopsis thaliana*), the growth of the seedling is supported mainly by hydrolysis of the oil reserve (Huang *et al.*, 1990). Lipases are involved in the first step of this series of reactions and may control a crucial step in post-germination of seed. Plant lipases play a role in the production of goods such as bread and beer. They may decrease the shelf life of plant-derived foods, due to their participation in the processes that govern spoilage (Mukherjee *et al.*, 1994). Clearly, lipases are widely employed in the food, cosmetic, detergent and pharmaceutical industries.

B. 5 Structural Bioinformatics

The Nomenclature Committee of the International Union of Biochemistry (IUB) has classified hydrolases according to substrates recognized. The term lipase commonly refers to triacylglycerol hydrolases (EC 3.1.1.3). However, as pointed out by the Committee, many of the enzymes hydrolyse a wide variety of ester substrates-- e.g., phospholipases, cholesterol esterase (EC 3.1.1.13), cutinase, amidase and other esterase

type activities that makes the classification somewhat arbitrary (Schmid and Verger, 1998; Bornscheuer and Kazlauskas, 1999). All of these enzymes can be regarded as lipases.

Hydrolases are enzymes that catalyze the hydrolysis of various bonds. Some of these enzymes pose problems because of their wide specificity or substrate compatibility. Thus, deciding whether two preparations described by different authors are the same can be difficult. While the systematic name always includes 'hydrolase', the common name is, in most cases, formed by the name of the substrate with the suffix "ase". It is understood that substrates with this suffix, and no other indicator, indicate a hydrolytic enzyme.

Fungi produce different classes of lipolytic enzymes, including carboxylesterases (EC 3.1.1.-), true lipases (EC 3.1.1.3) and a diverse group of phospholipases A1, A2, B, C and

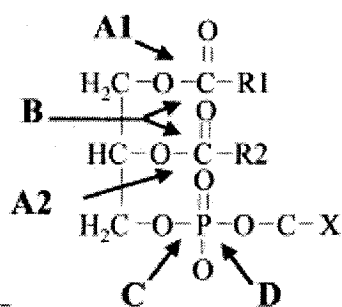
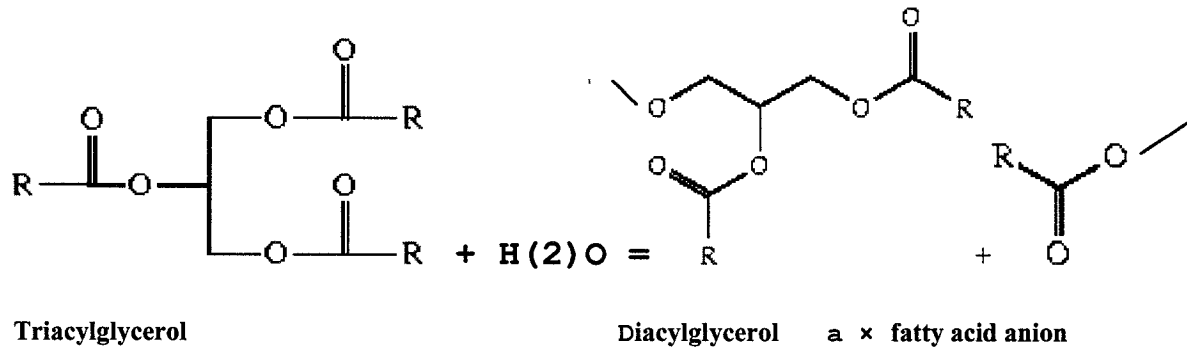


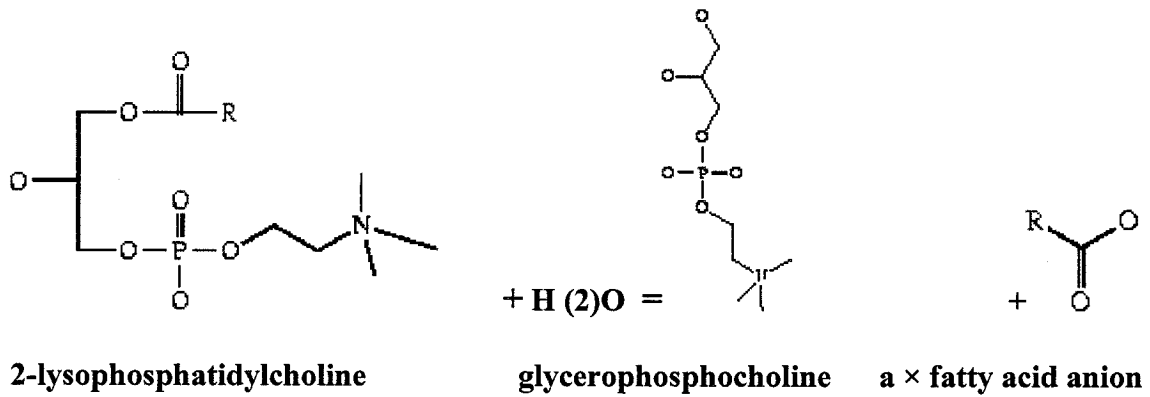
Figure 3: Sites of action of phospholipase A1, A2, B, C and D. The generic phospholipase structure is shown with the corresponding action sites of phospholipases designated by an arrow (adapted from Cox *et al.*, 2001).

D (Figure 3). ---

Carboxylesterases are enzymes which hydrolyze small ester containing molecules at least partly soluble in water. True lipases are enzymes that hydrolyse triglycerides into free fatty acids and glycerol. Triacylglycerol lipases are present in animals, plants, fungi, and bacteria. They have mainly triacylglycerol activity and are classified in the EC 3.1.1.3 group. The enzyme reaction is shown below by the following equation:



Phospholipids, on the other hand, have an important role in maintaining the structure and function of the cell membrane. Phospholipases generate second messengers, participate in cytotoxicity, and hydrolyze phospholipids in the gastrointestinal tract. They assist in regulating cellular functions in both mammalian and plant cells, and lower eukaryotes, including yeast and slime moulds. Phospholipases are a heterogeneous group of enzymes that are able to hydrolyse one or more ester linkages in glycerophospholipids. The actions of phospholipases can result in the destabilization of membranes, cell lyses and release of lipid second messengers (Ghannoum, 2000; Schmiel and Miller, 1999). These enzymes are categorized according to the specificity of the ester link that is cleaved (Figure 3). Five kinds of phospholipase activity have been described, including Phospholipase A1 and A2 (3.1.1.4); phospholipase B or lysophospholipase (3.1.1.5); phospholipase C or phosphatidylinositol-specific phospholipase C (EC 3.1.4.11) and phospholipase D (3.1.1.4). In this study, all phospholipases will be described except phospholipase A1 and A2 which were not experimentally found according to SWISS-PROT data base, but still they will be compared to phospholipase B.



Three-dimensional (3-D) and protein sequences are used to predict aspects of molecular function and evolution. Secondary structure prediction is a base of structure-based sequence analysis. Therefore, finding tools that accurately predict secondary structures are essential (Chen *et al.*, 1999; Klepeis *et al.*, 2003; McGuffin and Jones, 2003). Early studies by Dickerson *et al.* (1976) showed that multiple sequence alignments could be used to predict proteins structures. Zvelebil *et al.* (1987) integrated this theory into an automatic prediction method. These early studies suffered from lack of data and were used only on single sequences. Given the large families of homologous sequences now available and advancements in computing techniques, prediction accuracy is now above 75%.¹ More successful recent methods are PHD (Rost and Sander, 1994) and PSIPRED (Jones, 1999), reporting accuracy above 76%. Frishman and Argos (1997) hit 74.8% using PREDATOR and 79% using PORTER.

The results of the public prediction service (Predict Protein) have been used to determine protein structures (chain tracing in X-ray crystallography), as well as to formulate hypotheses about protein structure and function that guided experiments in molecular biology (Rost, 1998).

¹ Accuracy varies between different proteins (72% ±9%).

Secondary structure prediction tools used in this study are available on the SWISSPROT and NPS@ server. The SWISSPROT database is a manually annotated protein knowledgebase established in 1986 which strives to provide a high level of annotation (such as the description of the function of a protein, its domains structure post-translational modifications, etc.). NPS@ is the IBCP (Institute of Biology and Chemistry of Proteins) server contribution to PBIL in Lyon, France, and is a server developing several methods dedicated to protein secondary structure prediction, detection of fuzzy motifs in protein sequences, protein sequence analysis software, and so forth. From the SWISSPROT database, secondary structure tools used in this study include, SOPM (Geourjon and Delage, 1994), SOPMA (Geourjon and Delage, 1995), PSI-PRED (Jones, 1999), PROF. Prediction (Rost, 2001), JUFO (Meiler, 2002), PORTER (Pollastri *et al.*, 2002), NNPREDECTION (Kneller *et al.*, 1990), SSPRO (Baldi and Pollastri, 2003) and from NPS@ server, HNN (Guermeur, 1997), PHD (Rost, 1996), PREDATOR (Argos *et al.*, 1996), SIMPA 96 (Levin *et al.*, 1996), MLRC (Guermeur *et al.*, 1998), DSC (Guermeur *et al.*, 1998) and GORIV (Guermeur, 1996).

B. 5.1 Secondary Structure Prediction tools available from SWISS-PROT

B. 5.1.1 SOPM

Self-optimized method (SOPM) is a server for protein secondary structure prediction and was developed by Geourjon and Deleage in 1994. This method checks against an updated release of the Kabsch and Sander database, Data base of secondary structure prediction (DSSP), comprise protein chains. There are four steps in this method:

- (1) Building a sub-database drawn from DSSP using binary comparisons

of all protein sequences.

- (2) Taking into account the prediction of structural classes of proteins.
- (3) Predicting the secondary structure using an algorithm based on sequence similarity.
- (4) Iteratively determining the predictive parameters that optimize the prediction quality on the whole sub-database and applying the final parameters to the query sequence.

This method correctly predicts the secondary structures 69% of amino acids.

B. 5.1.2 SOPMA

SOPM was improved a year later in 1995 by Geourjon and Deleage to predict all the sequences from a set of aligned proteins belonging to the same family. The result was SOPMA, which had 69.5% accuracy.

B. 5.1.3 PSI-PRED

PSIPRED is a straightforward and reliable secondary structure prediction method using the output obtained from PSI-BLAST (Position Specific Iterated - BLAST). It incorporates this output with feed-forward neural networks for analysis. This program achieved the highest accuracy of 80.6% across all 40 submitted target domains with no obvious sequence similarity to structures present in PDB, which placed PSIPRED in first place out of 20 evaluated methods (an earlier version of PSIPRED was also ranked first in CASP3 held in 1998).

B. 5.1.4 PROF. Prediction

This program uses neural network on large data using different PSI-BLAST profiles, using the same concept implemented in PHD program. However, it does use more input data and has a third network layer for more accuracy.

B. 5.1.5 JUFO

JUFO offers a protein secondary structure prediction with 75% accuracy from its primary sequence. A neural network was trained with an amino acid property profile plus the position-based scoring matrix of a blast run (Jones, 1999).

B. 5.1.6 SSPRO

SSPRO (advanced recursive neural network system) is a server for protein secondary structure prediction based on an ensemble of 11 bi-directional recurrent neural networks (BRNNs). The only method that appears to improve prediction accuracy significantly via particular algorithms, as opposed to more divergent profiles, is SSpro. The algorithm to obtain multiple alignments of homologue sequences, based on PSI-BLAST instead of BLAST, is exploited. Experiments on an independent test set yields 78% accuracy (Pollastri *et al.*, 2002).

B. 5.1.7 PORTER

Porter is a new system for protein secondary structure prediction in three classes—it is an evolution of SSPRO. It increases the BRNN's (Bi-directional Recurrent Neural Networks) from 11 to 45. Porter was tested by a rigorous validation procedure

and achieves 79% accuracy (Pollastri *et al.*, 2002).

B. 5.1.8 NNpredict

This program predicts the secondary structure type for each residue in an amino acid sequence. Similar to other programs, it uses sequence as input. The output is a secondary structure prediction for each position in the sequence. To increase accuracy, it takes the tertiary class to account for the possible options (none, all-alpha, all-beta, or alpha/beta) for prediction. The best case prediction was 79% for the class of all-alpha proteins.

B. 5.2 Secondary structure prediction tools from NPS@

B. 5.2.1 HNN

The HNN (Hierarchical Neural Network) prediction method is made up of two networks: a sequence-to-structure network and a structure-to-structure network. Its prediction is based only on local information. Thus, the first layer network predicts the secondary structure of the central residue-- this is called the sequence-to-structure network. The second layer network, called the structure-to-structure network, filters the outputs from the first one and produces the final prediction results. To improve the accuracy of this tool, physico-chemical data have been taken into account for the structure-to-structure network.

B.5.2.2. PHD

PHD is neural network system (a sequence-to-structure level and a structure-structure level) to predict secondary structure (PHDsec), relative solvent accessibility (PHDacc) and trans membrane helices (PHDhtm) (Rost and Sander, 1993). The NPS@ server only uses PHDsec. PHDsec focuses on predicting hydrogen bonds. This program uses the BLASTP search for sequences and filters the result by aligning them with CLUSTALW. It uses these results as the input for the neural network.

B. 5.2.3. PREDATOR

PREDATOR is a secondary structure prediction method based on recognition of potential hydrogen-bonded residues in a single amino acid sequence (Frishman and Argos, 1996). The unique feature of this approach involves statistics on residue type occurrences in different classes of beta-bridges to describe interaction of beta-strands. The alpha-helical structures are also recognized on the basis of amino acid occurrences in hydrogen-bonded pairs.

B. 5.2.4. SIMPA96

This secondary structure prediction algorithm assumes that short homologous sequences of amino acids have the same secondary structure tendencies. Comparisons are made between secondary structure predictions from an X-ray database and an empirically determined similarity matrix which assigns a sequence similarity score between any two sequences of seven residues in length. This homologue method had a prediction accuracy of 62.2%.

B. 5.2.5. MLRC

MLRC (Multivariate Linear Regression Combination) is a secondary structure prediction method which combines GOR4, SIMPA96 and SOPMA (Guermeur *et al.*, 1999). It post-processes the outputs of protein secondary structure prediction methods and generates class posterior probability estimates. Experimental results establish that it can increase the recognition rate of methods that provide inhomogeneous scores, even if their individual prediction successes are largely different.

B. 5.2.6. DSC

Discrimination of protein Secondary structure Class (DSC) generates probabilities for helix and strand secondary structural states of each residue in a domain sequence. For input, it uses associated multiple sequence alignment CLUSTALW. This makes the prediction method comprehensible and allows the relative importance of the different sources of information used to be measured (King & Sternberg, 1996). The DSC method from multiply aligned homologous sequences has an overall per residue three-state accuracy of 70.1% accuracy.

B. 5.2.7. GORIV

GOR IV is the fourth version of GOR secondary structure prediction methods based on the information theory (Garnier *et al.*, 1996). GOR IV uses all possible pair frequencies within the window of 17 amino acid residues. GOR IV has a mean accuracy of 64.4% for a three state prediction.

C. METHODS

C.1 Comparative Study of Fungal Lipases

C.1.1 Multiple Sequence Alignment

Protein sequences selected for this study includes fungal lipases from “true” lipases, lysophospholipase and phospholipase C whose activities have been experimentally determined. Fifty-two such lipases from diverse fungal species were selected by surveying publications on search engines such as SWISS-PROT (<http://us.expasy.org/sprot>) and NCBI (<http://www.ncbi.nlm.nih.gov>), and by browsing sequence databases such as GeneBank, and SWISS-PROT. The amino acid sequences of these lipases were aligned using Multiple Sequence Alignment program, CLUSTAL W (Thompson *et al.*, 1994) at <http://www.ebi.ac.uk/clustalw>. Upon completion of sequence alignment, the data were saved as TEXT-FILE ONLY format prior to transfer to a data exploration program, MacClade 4.03 (Maddison and Maddison, 2002), and then to a phylogenetic tree-building and analysis program, PAUP 4.0, beta 8 (Swofford *et al.*, 2001). All the alignments were visually inspected and manually edited, as needed, in MacClade 4.03 before tree reconstruction. The BLAST tool (Altschul *et al.*, 1997) was also used for sequence similarity searches. The experimentally-determined 3-D structures were retrieved from the Protein Data Bank (Berman *et al.*, 2000). The protein structures were displayed by the program Cn 3-D from NCBI.

C. 1.2 Phylogenetic Analysis and Statistics (Distance Analysis)

UPGMA (Unweighted Pair Group Method with Arithmetic Mean) was used in this study to determine the phylogenetic tree. The aligned amino acid sequences were analyzed using the default options of the program and all characters were weighted equally, gaps treated as missing characters, multi-state taxa interpreted as uncertain, and the distance measure was set to be equal to mean character difference.

C.2 Evaluation of Secondary Structure Prediction Tools for fungal Lipases

C.2.1 Secondary Structure Tools

The best way to assess the accuracy of a method is by carrying out “blind trials”. A “blind trial” was used for the “protein Structure Prediction Challenge” meeting (Asilomar, California; 4-6 December 1994), where numerous prediction tools were tested to assess the accuracy of secondary structure prediction from multiple alignments and protein-fold recognition. The input for my analysis was the eight sequences with known X-ray structures (Table 1) and we also carried out “blind trials”. For some lipase sequences there have been more than one crystal structure determined², thus we screened the Protein Data Bank (July 2005) to select lipase sequences with resolutions of 2.35 Å or better (Table 1). We used fifteen different programs available online from SWISS-PROT (<http://us.expasy.org/sprot>) and NPS@ (<http://umber.sbs.man.ac.uk/dbbrowser/bioactivity>) to predict the secondary structure of sequences with unknown 3-D structures (Table 2).

² For instance, *Candida rugosa* has seven known structures including opened and closed.

Table 1: Sequences with known crystal structures (Information from Protein Data

Bank)

| PDB# | AC# | EC# | Name | Protein chain | Release Date | Resolution |
|--|--------|---------|--------------------------------|---------------|--------------|------------|
| 1thg | P22394 | 3.1.1.3 | Hydrolase(carboxylic esterase) | 544 aa | 1993 | 1.80 A |
| 1gz7 | P32946 | 3.1.1.3 | Hydrolase | 534 aa | 2003 | 1.97 A |
| 1cle | P32947 | 3.1.1.3 | Cholesterol esterase | 534 aa | 1996 | 2.00 A |
| 1llf | P32947 | 3.1.1.3 | Hydrolase | 534 aa | 2003 | 1.40 A |
| 1dt3 | O59952 | 3.1.1.3 | Hydrolase | 269 aa | 2000 | 2.69 A |
| 1dt5 | O59952 | 3.1.1.3 | Hydrolase | 269 aa | 2000 | 2.40 A |
| 1dte | O59952 | 3.1.1.3 | Hydrolase | 269 aa | 2000 | 2.35 A |
| 1du4 | O59952 | 3.1.1.3 | Hydrolase | 269 aa | 2000 | 2.50 A |
| 1ein | O59952 | 3.1.1.3 | Hydrolase | 269 aa | 2000 | 3.00 A |
| 1lgy | P61871 | 3.1.1.3 | Hydrolase(carboxylic esterase) | 265 aa | 1996 | 2.20 A |
| 1lpm | P20261 | 3.1.1.3 | Hydrolase | 534 aa | 1995 | 2.20 A |
| 1lpn | P20261 | 3.1.1.3 | Hydrolase | 534 aa | 1995 | 2.20 A |
| 1lpo | P20261 | 3.1.1.3 | Hydrolase | 534 aa | 1995 | 2.20 A |
| 1lpp | P20261 | 3.1.1.3 | Hydrolase | 534 aa | 1995 | 2.05 A |
| 1trh | P20261 | 3.1.1.3 | Hydrolase(carboxylic esterase) | 534 aa | 1994 | 2.10 A |
| 1crl | P20261 | 3.1.1.3 | Hydrolase(carboxylic esterase) | 534 aa | 1994 | 2.06 A |
| 1lps | P20261 | 3.1.1.3 | Hydrolase(carboxylic esterase) | 534 aa | 1995 | 2.20 A |
| 1tca | P41365 | 3.1.1.3 | Hydrolase(carboxylic esterase) | 317 aa | 1994 | 1.55 A |
| 1tcb | P41365 | 3.1.1.3 | Hydrolase(carboxylic esterase) | 317 aa | 1994 | 2.10 A |
| 1tcc | P41365 | 3.1.1.3 | Hydrolase(carboxylic esterase) | 317 aa | 1994 | 2.50 A |
| 3tgl | P19515 | 3.1.1.3 | Hydrolase(carboxylic esterase) | 265 aa | 1993 | 1.90 A |
| 1tgl | P19515 | 3.1.1.3 | Hydrolase(carboxylic esterase) | 265 aa | 1990 | 1.90 A |
| 4tgl | P19515 | 3.1.1.3 | Hydrolase(carboxylic esterase) | 265 aa | 1993 | 2.60 A |
| Abbreviations: AC#, Accession number; EC#, Enzyme commission; PDB#, Protein database bank; aa, amino acid; | | | | | | |
| | | | | | | |
| | | | | | | |

These programs classify each residue into three classes (H = α -helix, E/S = strand or β -sheets and the rest known as coils). Each sequence was analyzed separately using each of the above tools (Table 2) and an evaluation performed with respect to a scoring mechanism introduced in this study. A pair-wise similarity score in the range of (0, 1) is computed between all amino acid residues. Score "1" is given to amino acids that have correct matches for the two states, alpha and beta, (similar to crystal structure) and score "0" is given to others.

The tool that predicted secondary structure most similar to the known X-ray structure for each sequence was selected to run for the rest of the homologues sequences with unknown 3-D structure. These selected tool(s) predict secondary structures based on: statistical methods (Chou and Fasman, 1974), physico-chemical (Lim, 1974, Ptitsyn and Finkelstein 1983), sequence patterns, evolutionary conservation, and neural networks.

Table 2: Secondary structure prediction tools used in this study

| | Programs | Reference |
|------------|------------------|------------------------------------|
| SWISS-PROT | SOPM | (Geourjon and Delage, 1994) |
| | SOPMA | (Geourjon and Delage, 1995) |
| | PSI-PRED | (Jones, 1999) |
| | Porter | (Pollastri <i>et al.</i> , 2002) |
| | PROF.-Prediction | (Rost, 2001) |
| | JUFO | (Jens Meiler <i>et al.</i> , 2002) |
| | SSPRO | (Baldi <i>et al.</i> , 2003) |
| | NNPREDICTION | (Kneller <i>et al.</i> , 1990) |
| NPS@ | HNN | (Guermeur, 1997) |
| | GORIV | (Garnier <i>et al.</i> , 1996) |
| | SIMPA 96 | (Levin <i>et al.</i> , 1996) |
| | PREDATOR | (Argos <i>et al.</i> , 1996) |
| | PHD | (Rost <i>et al.</i> , 1994) |
| | DSC | (King and Stenberg, 1996) |
| | MLRC | (Guermeur <i>et al.</i> , 1998) |

C. 2.2 Three-dimensional (3-D) Structure Prediction

Vector Alignment Search Tool (VAST) from NCBI with cross-linking to the Molecular Modeling Data Base (MMDB) and Protein Data Bank (PDB) was used to obtain the 3-D structure and structural neighbors. In order to obtain these 3-D structures we entered the four-digit alphanumeric entry codes. To view the 3-D structures of fungal lipases we should select “view 3-D structure” box. In addition to the 3-D structure of the query, neighbors (similar structures) ranked by similarity will be shown.

D. RESULTS

D.1 Comparative Study of Fungal Lipases

The distance-based UPGMA analysis of 52 selected amino acid sequences of fungal lipases resulted in a single dendrogram (Figure 4), which revealed four major clusters of sequences and a branch with one sequence (P34163). The sequence P34163 is the only known fungal lipase of the super-family “AB hydrolase”:

- Cluster 1 consisted of ten sequences of secretory lipases (group A), ten sequences of carboxylesterases (group B) and a single sequence, P54857, which occupied a basal position within this cluster.
- Cluster 2 consisted of six sequences of lipase.
- Cluster 3 consisted of 20 sequences of Phospholipase B.
- Cluster 4 consisted of three sequences of Phospholipase C and a single sequence of Phospholipase D, which was basal within this cluster.

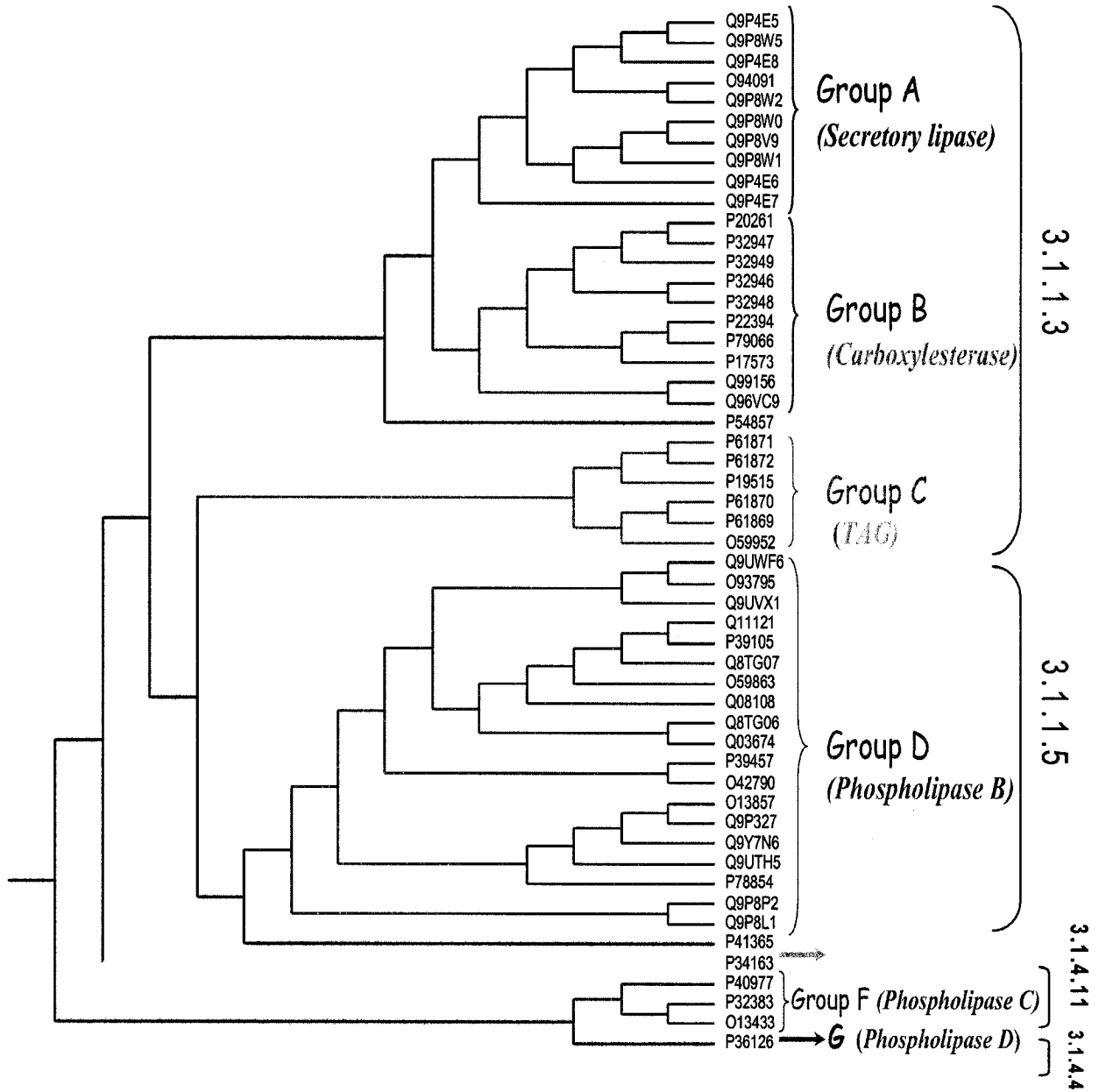


Figure 4: Phylogenetic tree using Neighbor joining method (UPGMA)

Table 3. Families and subfamilies of fungal lipolytic enzymes

| Family | Sub- | AC # | Organism | Properties | Molecular mass | Length | CATALYTIC ACTIVITY |
|------------------------|--------|--------|--|------------|-------------------|--------|---|
| | family | | | | | | |
| LIP (Secretory lipase) | AI | Q9P4E5 | Candida albicans (Yeast) | lipase 10 | 50 kDa | 465 | TAG+ H ₂ O = diacylglycerol + a carboxylate. |
| | AI | Q9P8W5 | Candida albicans (Yeast) | lipase 2 | 50 kDa | 466 | TAG+ H ₂ O = diacylglycerol + a carboxylate. |
| | AI | O94091 | Candida albicans (Yeast) | lipase 6 | 50 kDa | 463 | TAG + H ₂ O = diacylglycerol + a carboxylate. |
| | AI | Q9P8W2 | Candida albicans (Yeast) | lipase 1 | 50 kDa | 468 | TAG + H ₂ O = diacylglycerol + a carboxylate |
| | AI | Q9P8W2 | Candida albicans (Yeast) | lipase 3 | 51 kDa | 471 | TAG+ H ₂ O = diacylglycerol + a carboxylate. |
| | AII | Q9P8W0 | Candida albicans (Yeast) | (lipase 5) | 50 kDa | 463 | TAG+ H ₂ O = diacylglycerol + a carboxylate. |
| LIP (Secretory lipase) | AII | Q9P8V9 | Candida albicans (Yeast) | (lipase 8) | 50 kDa | 460 | TAG + H ₂ O = diacylglycerol + a carboxylate. |
| | AII | Q9P8W1 | Candida albicans (Yeast) | lipase 4 | 50 kDa | 459 | TAG + H ₂ O = diacylglycerol + a carboxylate. |
| | AII | Q9P4E6 | Candida albicans (Yeast) | lipase 9 | 49 kDa | 453 | TAG + H ₂ O = diacylglycerol + a carboxylate. |
| | A | Q9P4E7 | Candida albicans (Yeast) | lipase 7 | 49 kDa | 426 | TAG+ H ₂ O = diacylglycerol + a carboxylate. |
| Carboxylesterase | BI | P20261 | Candida rugosa (Candida cylindracea) | lipase 1 | 59 kDa | 549 | TAG + H ₂ O = diacylglycerol + a carboxylate. |
| | BI | P32947 | Candida rugosa (Candida cylindracea) | Lipase 3 | 59 kDa | 549 | TAG+ H ₂ O = diacylglycerol + a carboxylate. |
| | BI | P32949 | Candida rugosa (Candida cylindracea) | lipase 5 | 59 kDa | 549 | TAG + H ₂ O = diacylglycerol + a carboxylate. |

| | | | | | | | |
|---------------------------|------|--------|--|---------------------------------|--------|-----|--|
| Carboxylesterase | BI | P32946 | <i>Candida rugosa</i> (<i>Candida cylindracea</i>) | lipase 2 | 59 kDa | 548 | TAG + H ₂ O = diacylglycerol + a carboxylate. |
| | BI | P32948 | <i>Candida rugosa</i> (<i>Candida cylindracea</i>) | lipase 4 | 59 kDa | 549 | TAG + H ₂ O = diacylglycerol + a carboxylate. |
| | BII | P22394 | <i>Geotrichum candidum</i> (Oospora lactis) | (lipase 2) | 61 kDa | 563 | TAG + H ₂ O = diacylglycerol + a carboxylate. |
| | BII | P79066 | <i>Geotrichum fermentans</i> (Trichosporon fermentans) | Lipase1 | 61 kDa | 563 | TAG+ H ₂ O = diacylglycerol + a fatty acid anion |
| | BII | P17573 | <i>Geotrichum candidum</i> (Oospora lactis) | Lipase1 | 61 kDa | 563 | TAG + H ₂ O = diacylglycerol + a fatty acid anion |
| | BIII | Q99156 | <i>Yarrowia lipolytica</i> (<i>Candida lipolytica</i>) | lipase 1 | 55 kDa | 486 | TAG + H ₂ O = diacylglycerol + a carboxylate |
| | BIII | Q96VC9 | <i>Yarrowia lipolytica</i> (<i>Candida lipolytica</i>) | Lipase 3 | 56 kDa | 498 | TAG+ H ₂ O = diacylglycerol + a carboxylate |
| | CI | P61871 | <i>Rhizopus niveus</i> | Lipase, TAG, Lipase II | 42 kDa | 392 | TAG + H ₂ O = diacylglycerol + a carboxylate. |
| | CI | P61872 | <i>Rhizopus oryzae</i> (<i>Rhizopus delemar</i>) | Lipase, TAG | 42 kDa | 392 | TAG+ H ₂ O = diacylglycerol + a carboxylate. |
| | CI | P19515 | <i>Rhizomucor miehei</i> | Lipase, TAG | 40 kDa | 363 | TAG + H ₂ O = diacylglycerol + a carboxylate. |
| Lipase_3 (Lipase class 3) | CII | P61870 | <i>Penicillium camembertii</i> | Mono- and diacylglycerol lipase | 33 kDa | 305 | -- |
| | CII | P61869 | <i>Penicillium cyclopium</i> | Mono- and diacylglycerol lipase | 33 kDa | 305 | -- |

| | | | | | | | | | |
|--|-----|--------|--|---|--------|-----|---|--|--|
| PLA2_B (Lysophospholipase catalytic domain) | | | <i>Thermomyces</i> | | | | | | |
| | CII | O59952 | <i>lanuginosus</i> (<i>Humicola lanuginosa</i>) | Lipase, TAG | 32 kDa | 291 | TAG + H2O = diacylglycerol + a carboxylate. | | |
| | DI | Q9UWF6 | <i>Candida albicans</i> | Lysophospholipase 1, phospholipase B1 | 66 kDa | 605 | 2-lysophosphatidylcholine + H2O = glycerophosphocholine + a carboxylate | | |
| | DI | O93795 | <i>Candida albicans</i> | Lysophospholipase 2, phospholipase B 2 | 67 kDa | 608 | 2-lysophosphatidylcholine + H2O = glycerophosphocholine + a carboxylate | | |
| | DI | Q9UVX1 | <i>Candida albicans</i> | Lysophospholipase 3, phospholipase B 3 | 81 kDa | 754 | 2-lysophosphatidylcholine + H2O = glycerophosphocholine + a carboxylate | | |
| | DII | Q11121 | <i>Torulasporea delbrueckii</i> (<i>Saccharomyces rosei</i>) | Lysophospholipase, phospholipase B | 71 kDa | 649 | 2-lysophosphatidylcholine + H2O = glycerophosphocholine + a carboxylate | | |
| | DII | P39105 | <i>Saccharomyces cerevisiae</i> | Lysophospholipase, phospholipase B1 | 72 kDa | 664 | 2-lysophosphatidylcholine + H2O = glycerophosphocholine + a carboxylate | | |
| | DII | Q8TG07 | <i>Candida glabrata</i> (<i>Torulopsis glabrata</i>) | Lysophospholipase 1, phospholipase B1 | 72 kDa | 659 | 2-lysophosphatidylcholine + H2O = glycerophosphocholine + a carboxylate | | |
| | DII | O59863 | <i>Kluyveromyces lactis</i> (<i>Yeast</i>) | lysophospholipase, Phospholipase B | 70 kDa | 640 | 2-lysophosphatidylcholine + H2O = glycerophosphocholine + a carboxylate | | |
| | DII | Q08108 | <i>Saccharomyces cerevisiae</i> | Lysophospholipase 3, phospholipase B 3 | 75 kDa | 686 | 2-lysophosphatidylcholine + H2O = glycerophosphocholine + a carboxylate | | |
| | DII | Q03674 | <i>Saccharomyces cerevisiae</i> | Lysophospholipase 2, phospholipase B 2 | 75 kDa | 706 | 2-lysophosphatidylcholine + H2O = glycerophosphocholine + a carboxylate | | |
| | DII | Q8TG06 | <i>Candida glabrata</i> (<i>Yeast</i>) (<i>Torulopsis glabrata</i>) | Lysophospholipase 2, phospholipase B 2 | 75 kDa | 695 | 2-lysophosphatidylcholine + H2O = glycerophosphocholine + a carboxylate | | |

| | | | | | | | | |
|-------------------|---|---------------------------------|--|---|--------|-----|---|--|
| Abhydro lase_1 | PLA2_B (Lysophospholipase catalytic domain) | | | | | | | |
| | DIII | P39457 | <i>Penicillium chrysogenum</i> (<i>Penicillium notatum</i>) | Lysophospholipase , phospholipase B | 66 kDa | 612 | 2-lysophosphatidylcholine + H2O = glycerophosphocholine + a carboxylate | |
| | DIII | O42790 | <i>Neurospora crassa</i> | lysophospholipase, Phospholipase B | 70 kDa | 653 | 2-lysophosphatidylcholine + H2O = glycerophosphocholine + a carboxylate | |
| | DIV | O13857 | <i>Schizosaccharomyces pombe</i> | Putative lysophospholipase, Phospholipase B | 69 kDa | 624 | 2-lysophosphatidylcholine + H2O = glycerophosphocholine + a carboxylate | |
| | DIV | Q9P327 | <i>Schizosaccharomyces pombe</i> | Phospholipase B | 75 kDa | 673 | 2-lysophosphatidylcholine + H2O = glycerophosphocholine + a carboxylate | |
| | DIV | Q9Y7N6 | <i>Schizosaccharomyces pombe</i> | Phospholipase B | 68 kDa | 633 | 2-lysophosphatidylcholine + H2O = glycerophosphocholine + a carboxylate | |
| | DIV | Q9UTH5 | <i>Schizosaccharomyces pombe</i> | Probable lysophospholipase, Phospholipase B | 70 kDa | 644 | 2-lysophosphatidylcholine + H2O = glycerophosphocholine + a carboxylate | |
| | DIV | P78854 | <i>Schizosaccharomyces pombe</i> | Lysophospholipase 1, phospholipase B1 | 67 kDa | 613 | 2-lysophosphatidylcholine + H2O = glycerophosphocholine + a carboxylate | |
| | DV | Q9P8P2 | <i>Cryptococcus neoformans</i> var. <i>grubii</i> | Phospholipase B, Lysophospholipase | 69 kDa | 637 | 2-lysophosphatidylcholine + H2O = glycerophosphocholine + a carboxylate | |
| | DV | Q9P8L1 | <i>Cryptococcus neoformans</i> | Phospholipase B | 69 kDa | 637 | 2-lysophosphatidylcholine + H2O = glycerophosphocholine + a carboxylate | |
| E | P34163 | <i>Saccharomyces cerevisiae</i> | - | 63 kDa | 548 | | | |

| PI-PLC-X, PI-PLC-Y | | | | | | | |
|--------------------|---------------|---|---|---------|------|---|--|
| F | P32383 | <i>Saccharomyces cerevisiae (Baker's yeast)</i> | 1- phosphatidylinositol-4,5-bisphosphate phosphodiesterase 1, Phosphoinositide phospholipase C | 100 kDa | 869 | 1-phosphatidyl-1D-myo-inositol 4,5-bisphosphate + H ₂ O = 1D-myo-inositol 1,4,5-trisphosphate + diacylglycerol | |
| F | O13433 | <i>Candida albicans</i> | 1- phosphatidylinositol-4,5-bisphosphate phosphodiesterase 1, Phosphoinositide phospholipase C | 124 kDa | 1099 | 1-phosphatidyl-1D-myo-inositol 4,5-bisphosphate + H ₂ O = 1D-myo-inositol 1,4,5-trisphosphate + diacylglycerol | |
| F | P40977 | <i>Schizosaccharomyces pombe</i> | 1- phosphatidylinositol-4,5-bisphosphate phosphodiesterase 1, Phosphoinositide phospholipase C | 102 kDa | 899 | 1-phosphatidyl-1D-myo-inositol 4,5-bisphosphate + H ₂ O = 1D-myo-inositol 1,4,5-trisphosphate + diacylglycerol | |
| G | <u>P36126</u> | <i>Saccharomyces cerevisiae (Baker's yeast)</i> | Phospholipase D1, PLD1 | 160 kDa | 1380 | phosphatidylcholine + H ₂ O = choline + a phosphatidate | |
| | P54857 | <i>Saccharomyces cerevisiae (Baker's yeast)</i> | Other Lipase Lipase 2, Triacylglycerol lipase | 38 kDa | 326 | Triacylglycerol + H ₂ O = diacylglycerol + a carboxylate | |
| | P41365 | <i>Candida antarctica (Yeast) (Trichosporon oryzae)</i> | Other Lipase Lipase B (CALB) | 36 kDa | 342 | Triacylglycerol + H ₂ O = diacylglycerol + a carboxylate. | |

Abbreviations: AC#, Accession number; EC#, Enzyme commission;

D.1.1 Group A (Secretory lipase)

The group A consists of lipase 1 (O94091), lipase 2 (Q9p8W5), lipase 3 (Q9P8W2), lipase 4 (Q9P8W1), lipase 5 (Q9P8W0), lipase 6 (Q9P4E8), lipase 7(Q9P4E7), lipase 8 (Q9P8V9), lipase 9 (Q9P4E6), and lipase 10 (Q9P4E5) -- all members of a single gene family (Hube *et al.*, 2000). These gene sequences were identified from *Candida albicans* and have a molecular mass of 49-51 kDa. They show high similarity in sequence and the amino acid identities ranged from 36% between lipase (Q9P8V9 and Q9P4E7) to 85% (between Q9P8W0 to Q9P8V9) (Table 4).

| Table. 4 | Q9P4E5 | Q9P8W5 | Q9P4E8 | O94091 | Q9P8W2 | Q9P8W0 | Q9P8V9 | Q9P8W1 | Q9P4E6 | Q9P4E7 |
|----------|--------|--------|--------|--------|--------|--------|--------|--------|--------|--------|
| Q9P4E5 | 0 | 68% | 67% | 56% | 58% | 53% | 52% | 51% | 50% | 38% |
| Q9P8W5 | | 0 | 65% | 58% | 57% | 52% | 52% | 50% | 50% | 39% |
| Q9P4E8 | | | 0 | 59% | 63% | 52% | 53% | 51% | 49% | 38% |
| O94091 | | | | 0 | 73% | 55% | 55% | 55% | 53% | 39% |
| Q9P8W2 | | | | | 0 | 52% | 51% | 50% | 49% | 39% |
| Q9P8W0 | | | | | | 0 | 85% | 81% | 76% | 36% |
| Q9P8V9 | | | | | | | 0 | 82% | 73% | 36% |
| Q9P8W1 | | | | | | | | 0 | 76% | 37% |
| Q9P4E6 | | | | | | | | | 0 | 36% |
| Q9P4E7 | | | | | | | | | | 0 |

Table 4. Pairwise comparison of lipase1-lipase 10 using the program NCBI (Align two sequences BLAST).

These ten isoenzymes could be further divided in to two subfamilies (A.I and A.II):

Sub-family A.I includes Q9P4E5, Q9P8W5, Q9P4E8, O94091, and Q9P8W2 with more than 56% identity (Table 4) while sub-family A.II includes, Q9P8W0, Q9P8V9, Q9P8W1 and Q9P4E6 which were at least 49% identical to each other (Table 4). Lipase 7 (Q9P4E7) of the “family A” occupied a basal position to A.I and A.II, (Table 3 and Figure 4). The lipases of sub-family A.I have a molecular mass of 50-51

kDa, and contain a sequence of 16 amino acids as a signal peptide. In contrast to the typical catalytic triad (Ser, Asp and His) found in other lipases, the family A.I lipases have a dyad catalytic site with two residues for the catalytic activity at positions 196 (serine) and 344 (histidine). Similar to other lipases, the group A.I lipases have a conserved motif, Gly -X₁- Ser -X₂- Gly around the serine residue. The residues X₁ and X₂ of sub-family A.I lipases are glycine and tyrosine respectively. This sub-family includes conserved *N*-glycosylation site at positions 231 and 319 and at a few more positions as given in Figure 5. They consist of four conserved cysteine residues and a conserved lipase motif which may form disulfide bridges, yielding a similar 3-D structure. However, no crystallography-based structures are available for these protein sequences.

```

Q9P4E5      MKTLLIFLAFLLSSIFASLIGLTPPSKSTFYSPPVCFATAPPCDILIRNTPSAPSSLYLP 60
Q9P8W5      MKGLVFLGLLPTIYASLVHITPASEDDFYNPPAGFESARNCDILKLRNSPNRLASFYFP 60
Q9P4E8      MRDLILFLSLLHTIFASLFLSKPPSQDDFYKAPACFKAAKPCDILRTRKSPNKPSLYAP 60
O94091      MRGIAVFLAFISLIFASPLTVKSPLVDDFYTAPDQYESARLCEILLRLRKTPSKLSSMFFE 60
Q9P8W2      MKTLVVLCTLLSIFASPLSLKSPLVDDFYNPPRCYESARLCEILLRLRKTPGKISSLFIP 60
* : : : : : * : * * . . . . * . * . . * * : : * * * : * * * * : * : * : :

Q9P4E5      IVVKNAWQLLIRSEDSFGNPNAFVATLIQPLNANPSKLVSYQSWEDASHIDCSPSYGMQF 120
Q9P8W5      IDVKNAWQLLVKSEDSFGNPNAFVTTLIEPYNADPSKVVSYQTWEDASNINCSPSYGAQF 120
Q9P4E8      VDVQNSWQLLVREDSFGNPNAFVTTIIQPKNADPSKVVSYQNWEDASNINCSPSYGSQL 120
O94091      IDIKNSWQLLVREDSFGNATAIVTTVIEPYNADPSKVLSYQTFEDSANIECSPSYGMQY 120
Q9P8W2      VEVKNSWQLLVREDSFGNAAAIVTTVIEPFNADPSKVVSYQSWEDAANIECSPSYGMQF 120
: : * : * * * : * * * * * . * : * : * * * * * : * * * * * : * * * * * : * * * * * *

Q9P4E5      KSPATVTTQIDMTLIVPLLQNGYVVIIPDYEGPKSTFTVGRQSGKATLNSIRAALQTGA 180
Q9P8W5      GSPLSTITTQIDMTLIVPLRSGGYVVTPDYEGPKATFAVGRQSGQATLDSVRAILKSGS 180
Q9P4E8      GAPLSTILTQLDMTFIVPLKSGYVVLPDYEGPKSTFGVGRQSGKATLDSIKAVLTKD 180
O94091      GAPWSTVATQIDMALMVPMLKQGYVVSPDYEGPKSTFTVGRQSGKATLDSIRALILKSN 180
Q9P8W2      GAPLSSVQTQVDMIFIVPLLDKGCFVLPDYEGPKSTFGVGRQSGKATLDSIKAVLTKD 180
* : : : * * : * * : : * * * . * : * : * * * * * : * * * * * : * * * * * : * :

Q9P4E5      FSGIKKTAKVALWGYSGGSLATGWAISLQSKYAPELKENLIGAAVGGFATNTIAVAEAVD 240
Q9P8W5      FSGINEDAKVALWGYSGGSLATGWAALQPVYAPELQKNIVGAAVGGFAANTIAIAESVD 240
Q9P4E8      FSGINDAKVVLWGYSGGSFASGWAAVLQPEYAPELKDNLIGAALGFAANTIGIAESVD 240
O94091      FTGIKSDAKVAMWGYSGGSLASGWAALQPKYAPELKKNLIGAALGFVNTITATAEATD 240
Q9P8W2      FSGINDAQVAMWGYSGGTIAAGWAATLQPKYAPELKKNLIGAALGFVNTITATAEATD 240
* : * * . * : * : * * * * * : * * * * * * * . * * * * * : * * * * * *

Q9P4E5      GTVFSGFIPLALNGLANEYPDFKRLYGEVKLSARSTMEKGSQNCLAASLVGYPMSQYFT 300
Q9P8W5      GTIFSGLITLALNGLANEYPDLKTAFYEELSDFAVPEFKAGAENCLAENIFHYPLHQYFT 300
Q9P4E8      GEVFSGFIPLALNGLANEYPDFKRLYEEVKPGAKADLQKGAENCLAASLISYPMYQYFT 300
O94091      GTLFAGLVPNALSGLANEYPEFKEILYQKVSKAATDNLRQGTEHCIGGAILYFAEDQYFT 300
Q9P8W2      GTLFAGLIPNALNGLANEFPDFKKRMYEVVEKRYEGALQQTQHCLGGAILHFADQVFT 300
* : * : * : . * * : * * * * * : * * * * * : * * * * * : * * * * * : * * * * *

Q9P4E5      GQNRAFEKGWLLQDEVFNKTIEDNLLKLDKTYLPQVPVLIYHGTIDEIIPIKDANAQY 360
Q9P8W5      GPKRAFEKGWLLKEDIFNKSIQDNLLIGLNKTYLPQVPVLIYHGTVDEIIPIKDPHAQY 360
Q9P4E8      GPRRVFEKGWSLLEDKTIGKTLEDNLLIALSKEHMPQIPIFVIYHGTIDKIIPIKSIKIY 360
O94091      GDDRAFPGGYGLLKEVVNKTISENNLMQMDKDYLPDIPIFVIYHGALDSIVPISNVHVTY 360
Q9P8W2      GDHRYFEQGYGLLEEVVFNRTISGNSLLYMDQEYLPDIPIFVIYHGSLDGIVPIPDVHGVY 360
* * * * * : * * * * * : . . . . . * : : : : * * * * * : * * * * * : * * * * *

Q9P4E5      QIWCDRGIQSLEFAEDLSAGHLAETFTGAPAALSWIDARFSGKPAVNGCQRTIRSSNVLY 420
Q9P8W5      QLWCDWGIESLEFAEDLSTGHLAETFTGAPAALAWIDARFDGKTPIQGCSHTTRLTNLLY 420
Q9P4E8      KNWCDWGIGSFEFSEDKSNGHTTETVVGAPAALTWIDARFAGKPAVEGCSFTTRASNFLY 420
O94091      KNWCDWGINSFEFSEDLLNGHTTETIVGAPAAITWLEARFDGEPVVKGCKKTSRITNFSY 420
Q9P8W2      KNWCDWGIDSFEFAEDSLNGHLTEIVVGAPAAITWLDARFDGQPVVEGCKKTTRITNFSY 420
: * * * * * : * * * * * : * * * * * : * * * * * : * * * * * : * * * * * : * * * * *

Q9P4E5      PGISITIRIYFEGISKTIFGVNLGSGVNAD-KSISNKFFAYIRKYI----- 465
Q9P8W5      PNTSDSTHSYFLGIYQAVFGTPLGPGINDNITINSGLLGLVSSII----- 466
Q9P4E8      PNISESAASYFKGIYQTILRSKLGSGVTSDDVSVN-GLRSLYHT----- 463
O94091      PNISDSTSSIFEGILNSVTGSELGPGVTSDNITLD-GLTGLGNFIDLK--- 468
Q9P8W2      PNISDSTRNFFKGIDSLTASQLGPGVTSDNVTLS-GLTGFMGLSKFKSV 471
* . * : * * * : : * * * . . . * : : : : * * * * * : * * * * * : * * * * *

```

Figure 5. CLUSTALW sequence alignment of Secretory lipase (sub-family A.I). The 16 amino acid signal peptide, the active site (Ser, His) along with the consensus lipase sequence GxSxG, the potential N-glycosylation site and cysteine residues available for disulfide bond formation are shown in this figure.

```

Q9P8W0      MLYLILFLIPIYAGLIFPTKPSDDPFYNPPKGFENAAVGDILQSRATPKSITGGFTPLK 60
Q9P8V9      MLFLLFLLITPIYAGLIFPTKPSDDPFYNPPKGFEEAAVGDILQSRATPKSITGRFAPLK 60
Q9P8W1      MLFLLFLLVAPIYAGLILPTKPSNDPFYNAPAGFEKAAVGDILQSRATPKSITGVFVVK 60
Q9P4E6      MLYLILFLIPIYAGVLLPTKPSIDPFYNAPAGFEKNAVGDILQFRKTPKSITGGFVPLN 60
           **:::***:*****::***** *****.* **::*:***** * ***.*** *.*::

Q9P8W0      IQNSWQLLVRSEDSFGNPNVIVTTVIEPVNADPSKIASYQVFEDAADKADCAPSYALQFGS 120
Q9P8V9      IQNSWQLLVRSEDSFGNPNVIVTTVIEPVNADPSKIASYQVFEDAADKADCAPSYALQFGS 120
Q9P8W1      IQNSWQLLVRSEDSFGNPNVIVTTVMEPFNADPSKLASYQVFEDSAKADCAPSYALQFGS 120
Q9P4E6      VQNSWQFLVRSEDSFGNPNVIVTTVIEPVNADPSKIASYQVSENAARADCAPSYALQFGS 120
           :*****:*****:*****.*.*****:***** *:*:*****:*****

Q9P8W0      DLTTFVTQAEMYLMAPLLDQGYVVSPDYEGPKSTFTIGKQSGQAVLNSIRAALKSGKIT 180
Q9P8V9      DLTTFVTQAEMYLMAPLLDQGYVVSPDYEGPKSTFTIGKQSGQAVLNSIRATLKSKIT 180
Q9P8W1      DVTTIATQVETYLLAPLLDQGYVVSPDYEGPKSTFTVGKQSGQAVLNSIRAALKSGKIT 180
Q9P4E6      DVSTLATQAETYLLAPLLDKGYVVSPDYEGPKSTFTVGKQSGQAVLNSIRASLKSKIT 180
           *:*.***.* **::*****:*****:*****:*****:*****:*****:*****:*****

Q9P8W0      NIKDDAKVVMWGYSGGSLASGWAAALQPSYAPELGGNLLGAALGGFVNTITATAQADTGT 240
Q9P8V9      NIKEDAKVVMWGYSGGSLASGWAAALQPSYAPELSSLLGAALGGFVNTITATAQAADGT 240
Q9P8W1      NLAENAKVVMWGYSGGSLASGWAAALQPNYAPELGGNLLGAALGGFVNTITATAEATDGT 240
Q9P4E6      NIAEDAKVLMWGYSGGSLASGWAAALQPDYAPELSRNLLGVALGGFITNVTATVEATDDT 240
           *: :*:*****:*****:*****.*.***.*****:***:***:***.*

Q9P8W0      VFAGIVANALGGVANEYPEFKSILQSDTDKKSVFEEFDGHCLIDGVLNYIGTSFLTGDHK 300
Q9P8V9      VFAGIVANALGGVANEYPEFKSILQSDTDKKSVFDEFDSSHCLADGVIDYINTSFLTGDNK 300
Q9P8W1      VFAGIMANALGGVANEYPEFKQILQNDTDKQSVFDQFDNHCLADGVINYIGKHFLSGTNK 300
Q9P4E6      IFAGTAANVLGGIANEYPEFKSILQNDTNKSSIFNKINNHCLTDSFIKYVGARFLTGDNK 300
           :*** **.*.***:*****:*****.*.***.*.*:***:***.***.***.*

Q9P8W0      IFKTGWDILKNPKIGKVVEDNGLVYQQLVPKIPVFIYHGSIDQIVPIVDTKKTYQNWCD 360
Q9P8V9      IFKTGWDILKSPTIAKIVEDNGLVYQQLVPKIPIFVYHGSIDQIVPIVNVKKTYQNWCE 360
Q9P8W1      IFKSGWNILKNPTISKIVEDNGLVYQQLVPKIPIILIYHGAIDQIVPIVNVKKTYQNWCD 360
Q9P4E6      VFKSGWNIFKNLVVSKIVKDNGLVYQQLIPTIPVFIYHGSMDQISFILNPKKTYQNWCD 360
           :***:***:***. .*:***:*****:***:***:***:*** **:: *****:

Q9P8W0      AGISSLEFAEDASNGHLTEAIMGAPAALTWIIDRFNGKQTVSGCQHQRFSNLEYPNIPS 420
Q9P8V9      GGISSLEFAEDGTNGHLTETVVGAPAALTWIIDRFNGKQTVSGCQHDKRLSNFQYPNISS 420
Q9P8W1      AGIASLEFSEDATNGHITETIVGAPVALTWIIDRFNGKQTVSGCQHVKRTSNFEYPNIPP 420
Q9P4E6      AGISSIEFAEDLTNGHFTESIVGAPAALTWIIDRFNKNPPVDGCQHVVRTTNYEYPNVSS 420
           .***:***:*** **::*****:*****:*****:***.*.*** * :*:***:..

Q9P8W0      SIANYFKAAMDVVLHGLGPDVQKDQVSPGKIKKLSIEMRWL 463
Q9P8V9      SILKYFVALDMMNSNGLGSDIQDKRITPDDLRFK--LLGW- 460
Q9P8W1      SILNYFKAALNLIQKGLGPDQKDVNPDGLKKSILV---- 459
Q9P4E6      SILDYFKAAMDVVAQQGLGPNIQKQLEIKSNL----- 453
           ** .***.*:: : ***.***:***:..

```

Figure 6. CLUSTALW sequence alignment of Secretory lipase (sub-family A.II). The 14 amino acid signal peptide, the active site (Ser, His) along with the consensus lipase sequence Gly-x-Ser-x-Gly, the potential N-glycosylation site and cysteine residues available for disulfide bond formation are shown in this figure.

The A.II sub-family is similar to A.I sub-family, but with a few key differences. They include sequences with accession numbers of Q9P8W0, Q9P8V9, Q9P8W1 and Q9P4E6 of lipases. They have fourteen amino acids as the signal peptide, (in contrast to sixteen amino acids in A.I). An *N*-glycosylation site was only conserved at residue 229. The four cysteine residues, (residues 110-281 and 359-404) and the conserved dyad catalytic sites are identical to A.I sub-family (Figure 6).

D.1.2 Group B (Carboxylesterases)

The family B comprises ten lipases from *Ascomycota*, which could be further divided into three sub-families (B.I, B.II and B.III). The sub-family B.I includes five sequences from *Candida rugosa* with accession numbers of P20261, P32947, P32949, P32946 and P32948. All five isozymes have similar structure with a molecular mass of 59 kDa and contain 534 amino acids in their mature protein (main chain). The sequences in the sub-family B.II are from *Geotrichum*, and include sequences with accession numbers of P22394, P79066 and P17573. The molecular mass of these lipases are 61kDa. The sub-family B.III comprises of two sequences from *Yarrowia lipolytica* (*Candida lipolytica*); yeast commonly used in genetic studies. The molecular mass of these two sequences (55 kDa) are lower than other sequences in the family B. The sequences in sub-family B.III include sequences with accession numbers of Q99156 and Q96VC9 (Figure 4 and Table 3). Although Q99156 and Q96VC9 are distinct from the rest of the sequences based on the distance dendrogram, they share common properties with the rest of the sequences in this family such as having the same EC number (3.1.1.3) and hydrolyzing triacylglycerol into diacylglycerol and a carboxylate. Unlike other lipases, the catalytic triad in group B (Carboxylesterase) is made up of the amino acids

Ser-Glu-His at positions 223, 355 and 365. The aspartate residue common to most lipases is replaced with glutamic acid in this group. However, in sequence Q99156, the aspartate catalytic residue remains unchanged. The motif Gly-X₁-Ser-X₂-Gly at the serine site (residue 193); (as given in figure 7) is conserved in this family. X₁ in this family is glutamic acid and X₂ is alanine. Family B.I and B.II include two disulfide bonds at regions (74-111) and (282-291) while there is only one disulfide bond found in sub-family B.III (Figure 7). *N*-glycosylation sites were conserved in sub-family B.I at position 365 in all five sequences, but at position 329 sequences P32946 and P32948 has diverged to a hydrophobic amino acid (phenylalanine). Moreover, these lipases show an α/β hydrolase fold, which is a common 3-D fold in several other hydrolases (Ollis *et al.*, 1992).

```

P22394      MVSKSLFLAAAVNLAGVLAQAPRPSLNGNEVISGVLEGKVDTFKGI PFADPPLNDRFKH 60
P79066      MVSKSLFLAAAVNLAGVLAQAPTAVLNGNEVISGVLEGKVDTFKGI PFADPPLNDRFKH 60
P17573      MVSKTFFLAAALNVVGTLAQAPTAVLNGNEVISGVLEGKVDTFKGI PFADPPVGLDRFKH 60
P20261      -----MELALALSLIASVAAAPTATLANGDTITGLNAINAEFLGIPFAEPPVGNLRFKD 55
P32947      -----MKLALALSLIASVAAAPTAKLANGDTITGLNAINAEFLGIPFAEPPVGNLRFKD 55
P32949      -----MKLALALSLIASVAAAPTATLANGDTITGLNAINAEFLGIPFAEPPVGNLRFKD 55
P32946      -----MKLCL-LALGAAVAAAPTATLANGDTITGLNAINAEFLGIPFAEPPVGLTRFKP 54
P32948      -----MKLALVLSLIVSVAAPTATLANGDTITGLNAINAEFLGIPFAQPPVGNLRFKP 55
Q96VC9      -----MPLELPSLNASIVGNTVQNG-----AVEQFLNIRYADIPG---KFEK 39
Q99156      -----MSVTSTSLNGTFNGISED-----GIEIFKGIKYANIPYRWAYAE 40
          : : : : : * . * : * : * :
          : : : : :

P22394      PQPFTGSYQGLKANDFS PACMQLD FGNSLTL LDKALGLAKVIPEEFRGPLYDMAKGTVSM 120
P79066      PQPFTGSYQGLKANDFS PACMQLD FGNSLTL LDKALGLAKVIPEEFRGPLYDMAKGTVSM 120
P17573      PQPFTGSYQGLKANDFS SACMQLD FGNAISLLDKVVLGKII PDNLRGPLYDMAQGSVSM 120
P20261      PVPYSGSLDQKFTSYGSPCMQMN FEGTYEENLPKAAALDLVMQS-----KVFEAVSP 108
P32947      PVPYSGSLNGQKFTSYGSPCMQMN FEGTYEENLGKTAALDLVMQS-----KVFAVLPQ 108
P32949      PVPYRGS LNGQSF TAYGSPCMQMN FEGTYEENLPKVALDLVMQS-----KVFAVLPN 108
P32946      PVPYASLNGQKFTSYGSPCMQMN FMGSFEDTL PKNARHLVLS-----KIFQVVLPN 107
P32948      PVPYASLNGQKFTSYGSPCMQMN FLGNWSSLPKAAINSLMQS-----KLFQAVLPN 108
Q96VC9      PVLKN-DWNGAEIDATKVGVPVCPQRT FNFVSPDDLWEKVNV-----TYQD 87
Q99156      ----IDDYDNGVFDCTQEGMACP-QVL F DYNI EKGP KEMP FDE----- 79
          : :
          : :

P22394      NEDCLYLN VFRPAGTKPDAKL PVMWVI YGGAFVYGSSA--AYPGNSYVKESINMGQP VVF 178
P79066      NEDCLYLN VFRPAGTKPDAKL PVMWVI YGGAFVYGSSA--AYPGNSYVKESINMGQP VVF 178
P17573      NEDCLYLN VFRPAGTKPDAKL PVMWVI YGGAFVYGSSA--SYPGNGYVKESVEMGQP VVF 178
P20261      SEDCLTIN VVRPPGTKAGANLPVMLWIFGGGF E VGGTS--TFPPAQMITKSIAMGKPI IH 166
P32947      SEDCLTIN VVRPPGTKAGANLPVMLWIFGGGF E IGSPT--IFPPAQMVTKSVLMGKPI IH 166
P32949      SEDCLTIN VVRPPGTKAGANLPVMLWIFGGGF E IGSPT--IFPPAQMVSKSVLMGKPI IH 166
P32946      DEDCLTIN VVRPPGTRASAGLPVMLWIFGGGF E LGGSS--LFPGDQMVAKSVLMGKPI IH 165
P32948      GEDCLTIN VVRPSGTPKGANLPVMLWIFGGGF E VGGSS--LFPPAQMITASVLMGKPI IH 166
Q96VC9      GLLCDNLI VTRPKGVSANARLPTV VVW IHGGSNIEGSI YNLI YE PQLVAESVRV GKPI V 147
Q99156      -FECSNLMITR PQGATN---LPVFVW IHGGSNLAGNGYCS DHPV PPFVKHSIVAGR PVLH 135
          * : : * * * . * * * : * * * . * : : * : * : * :
          * : : * * * . * * * : * * * . * : : * : * : * :

P22394      VSIN YRTGPF GFLGGDAITAE GNTNAGLHDQRKGLEWVSDN IANFGGDPDKVMI F GESAG 238
P79066      VSIN YRTGPF GFLGGDAITAE GNTNAGLHDQRKGLEWVSDN IANFGGDPDKVMI F GESAG 238
P17573      VSIN YRTGPF GFLGGDAITAE GNTNAGLHDQRKGLEWVSDN IANFGGDPDKVMI F GESAG 238
P20261      VSVN YRVSSWGFLAGDE IKAEGSANAGLKDQRLGMQWVADN IAAFGGDPTKVTF GESAG 226
P32947      VAVN YRVASWGFLAGDD IKAEGSGNAGLKDQRLGMQWVADN IAGFGGDP SKVTIF GESAG 226
P32949      VAVN YRLASFGFLAGPDI KAEGSSNAGLKDQRLGMQWVADN IAGFGGDP SKVTIF GESAG 226
P32946      VSMN YRVASWGFLAGPDI QNEGSGNAGLHDQRLAMQWVADN IAGFGGDP SKVTIY GESAG 225
P32948      VSMN YRVASWGFLAGPDI KAEGSGNAGLHDQRLGLQWVADN IAGFGGDP SKVTIF GESAG 226
Q96VC9      VCIE YRLGLAGFLT----KNGKGNWGTWDQYTG CQWVNRHIQDFGGDPLNVTLT GESAG 202
Q99156      VMIE YRLS AFGYLA VPD TNGN WGNW GARDQY TALQWISKHIVEFGGDP SQITIG GESAG 195
          * : : * * * . * * * : * * * . * : : * : * : * :

P22394      AMSVAHQ LIAYGGDNTYNGKKLFHSAILQSGG PLYPHDSSSVGPD ISYNRFAQYAGCDTS 298
P79066      AMSVAHQ LIAYGGDNTYNGKKLFHSAILQSGG PLYPHDSSSVGPD ISYNRFAQYAGCDTS 298
P17573      AMSVAHQ LVAYGGDNTYNGKQLFHSAILQSGG PLYFDSTSVGPESAYS RFAQYAGCDAS 298
P20261      SMSVMCHILWNDGDNTYK GKPLFRAGIMQSGAMVPSDAVDGIY GNEIFDLLASNAGCGSA 286
P32947      SMSVLCHL I WNDGDNTYK GKPLFRAGIMQSGAMVPSDPVDGT YGNEIYDLFVSSAGCGSA 286
P32949      SMSVLCHL I WNDGDNTYK GKPLFRAGIMQSGAMVPSDPVDGT YGTQIYD TLVASTGCSSA 286
P32946      SMSTFVHL V WNDGDNTYNGKPLFR AAIMQSGAMVPSDPVDGT YGTEIYNQVVASAGCGSA 285
P32948      SMSVMCQLL WNDGDNTYNGKPLFR AAIMQSGAMVPSDPVDG PYGTQIYDQVVASAGCGSA 286
Q96VC9      SVAVHNMLIK---DSMNGRKLFRNAVMMSGTLETITP QPKWHARLEEKVAKVTGKEVA 258
Q99156      SIGLHALMVHESMKPKEE--CIIHNVLSSGTMDRMG--TGTISENAFKPIYDGIKTLVG 251
          : : : : :
          : : : : :

P22394      ASANDTLECLRSKSSSVLHDAQNSYDLKDL FGLLPQFLGFGPRPDGNI IPDAAYELFRSG 358
P79066      ASANDTLECLRSKSSSVLHDAQNSYDLKDL FGLLPQFLGFGPRPDGNI IPDAAYELFRSG 358
P17573      AGDNETLACLRSKSSSVLHSAQNSYDLKDL FGLLPQFLGFGPRPDGNI IPDAAYELYRSG 358
P20261      S---DKLACL RGSVSDTLE DATN--NTPGFLAYSSLRLSYLPRPDG VNI TDDMYALVREG 341
P32947      S---DKLACLRSASD TLLDATN--NTPGFLAYSSLRLSYLPRPDGKNI TDDMYALVREG 341
P32949      S---NKLACL RGLSTQALLDATN--DTPGFLSYTSLRLSYLPRPDGANI TDDMYALVREG 341
P32946      S---DKLACL RGLSQTLYQATS--DTPGVLAYPSLRLSYLPRPDGTFIT TDDMYALVREG 340
P32948      S---DKLACLRSISNDKLFQATS--DTPGALAYPSLRLSYLPRPDGTFIT TDDMYALVREG 341
Q96VC9      D-----LASLSDKELLDAQIKLNVAVCMTC D-----DGDFFEPGWKQHLTPD 300
Q99156      D-----INTCSADELLEAQIKAGLDLGFY LQ-----DFFFPDWRNV R--- 289
          : * . * . * : : : :
          : : : : :

```



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P22394      RYAKVPYISGNQEDECTAFAFVALNATTPPHVKKWLQYIFYDASEASIDRVLSLYPQTLS 418
P79066      RYAKVPYISGNQEDECTAFAFVALNATTPPHVKKWLQYIFYDASEASIDRVLSLYPQTLS 418
P17573      RYAKVPYITGNQEDECTILAPVANATTPPHVKKWLKYICSEASDASLDRVLSLYPGSWS 418
P20261      KYANIPVLIIGDQNDECTFFGTSSLNVTTDAQAREYFKQSFVHASDAEIDTLMAYPGDIT 401
P32947      KYASVPVLIIGDQNDECTIFGLSSLNVTTNAQARAYFKQSFIFHASDAEIDTLMAYPGDIT 401
P32949      KYASVPVLIIGDQNDECTFLGLSSLNVTTEADAEAYLRKSFIFHATDADITALKAAAYPSDVT 401
P32946      KYAHVPVLIIGDQNDECTFLGLSSLNVTTDAQARAYFKQSFIFHASDAEIDTLMAYTSDIT 400
P32948      KCANVPVLIIGDQNDECTVFALSSLNVTTDAQARQYFKESFIFHASDAEIDTLMAYPSDIT 401
Q96VC9      WLDKL--IISDCKEGMLYFLPVN----AQDDELLAKVAKSPVGKEISELYGIKEGGDI 354
Q99156      -FKVSRVLLSDVIVDGTNF-----KNKINPAVRVTPENDEFDHKVFKLYNISTEDTW 339
      . . . : * . . . : :
P22394      VGSPFRTGILNALTPQFKRVAAILSDMLFQSPRRVMLSATKDVNRWTYLSTHLHNLVPFL 478
P79066      VGSPFRTGILNALTPQFKRVAAILSDMLFQSPRRVMLSATKDVNRWTYLSTHLHNLVPFL 478
P17573      EGAPFRTGILNALTPQFKRIAAIFDLLFQSPRRVMLNATKDVNRWTYLATQLHNLVPFL 478
P20261      QGSPFDTGILNALTPQFKRISAVLGDGLGFTLARRYFLNHYTGGTKYSFLSKQLSG-LPVL 460
P32947      QGSPFDTGIFNAITPQFKRISAVLGDGLAFIHARRYFLNHFFQGGTKYSFLSKQLSG-LPIM 460
P32949      QGSPFDTGILNALTPQLKRINAVLGDGLTFTLSRRYFLNHYTGGPKYSFLSKQLSG-LPIL 460
P32946      QGSPFDTGIFNAITPQFKRISALLGDGLAFTLARRYFLNYYQGGTKYSFLSKQLSG-LPVL 459
P32948      QGSPFDTGIFNAITPQFKRIAAVLDGLAFTLPRRYFLNHFFQGGTKYSFLSKQLSG-LPVI 460
Q96VC9      KSACLDLKTDAFNFYFNHLLFKKMEARNNGSTSRVYRLAVDEPNPHNPQRAHHAVDVL 414
Q99156      EDYHYKMMLFKGDETFIR--GNQQLLEFEQENIPVWRQLFDQIHPNDPSRLCHHAVDVL 397
      . . . : : . . . : :
P22394      GTFHGNELIFQFNVNIGPANSYLRYFISFANHHPNVGTNLLQWDQYTDE---GKEMLEI 535
P79066      GTFHGNELIFQFNVNIGPANSYLRYFISFANHHPNVGTNLLQWDQYTDE---GKEMLEI 535
P17573      GTFHGSDLLFQYYVDLGPSSAYRRYFISFANHHPNVGTNLKQWDMYTDS---GKEMLQI 535
P20261      GTFHSNDIVFQDYLLGSGSLIYNNAFIAFATDLDPNNTAGLLVKWPEYTSSSQSGNNLMMI 520
P32947      GTFHANDIWQDYLLGSGSVIYNNAFIAFATDLDPNNTAGLLVNWPKYTSSSQSGNNLMMI 520
P32949      GTFHANDIWQHFLGSGSVIYNNAFIAFATDLDPNNTAGLSVQWPKSTSSSQAGDNLMQI 520
P32946      GTFHGNDIWQDYLVGSGSVIYNNAFIAFANDLDPNKAGLWLNWPTYTSSSQSGNNLMQI 519
P32948      GTFHANDIWQDFLVSHSSAVYNNAFIAFANDLDPNKAGLLVNWPKYTSSSQSGNNLQI 520
Q96VC9      YMFNSTKFNEHGDKLSR---LFQSHFLRLAYGLEPWHDRNFGVYRNGGYQQQLPLSELNKV 471
Q99156      YMWDNWEMPEDKHAVAR---QYQDTLTKFVYGDQDPVVDKHLHYVHDNQFEILDKSKQFGDF 454
      . . . : : : * . . . : :

```

Figure 7: Amino acid sequence alignment carboxylesterase in family B including three sub-families (BI, BII, and BIII). The catalytic residues, N-glycosylation, the conserved proline residue (Pro 85 and phenylalanine residue 326) are shown in this figure.

D.1.3 Group C (AB hydrolase super family/TAG)

Family C comprises six sequences, further divided into two subfamilies (C.I and C.II). Group C.I has three sequences P61871 (*Rhizopus niveus*), P61872 (*Rhizopus oryzae*), and P19515 (*Rhizomucor miehei*). Sequences P61870 (*Penicillium camembertii*), P61869 (*Penicillium cyclopium*), and O59952 (*Thermomyces lanuginosus*) are in sub-family C.II (Figure 4). The 3-D structure for at least one sequence of each sub-family (P19515 and P61871 in sub-family C.I and O59952 in sub-family C.II) is known. The sub-family C.I consists of sequences from two filamentous fungi, *Rhizopus* and *Rhizomucor*, the only members of the phylum *Zygomycota* used in this study. The

lipases produced by these fungi are proteins of 363-392 amino acids with molecular masses of 40-42 kDa. The first known 3-D structure of sub-family C.I is P61871 from *Rhizopus niveus*. The X-ray crystallography structure of P61871 reported by Kohno (1996) showed that this sequence contains a catalytic center with a triad of three amino acids (Ser-Asp-His), similar to serine proteases such as chymotrypsin and subtilisin (Blow *et al.*, 1969). It also contains the consensus sequence (Gly-X₁-Ser-X₂-Gly, with X₁ = histidine and X₂ = leucine) as shown in Figure 8.

```

P61871 MVSFISISQGVSLCLLVSSMMLGSSAVPVSGKSGSSNTAVSASDNAALPPLISSRCAPPS 60
P61872 MVSFISISQGVSLCLLVSSMMLGSSAVPVSGKSGSSNTAVSASDNAALPPLISSRCAPPS 60
P19515 MVLKQRANYLGFELIVFFTAFLV--EAVPIKRQS---NSTVDS-----LPPLIPSRTSAPS 50
** . * ::::::: .***. :* *::*. *****.* :.**

P61871 NKGSKSDLQAEFYNMQNTEWYESHGGNLTSIGKRDDNLVGGMTLDLPSDAPPISLSST 120
P61872 NKGSKSDLQAEFYNMQNTEWYESHGGNLTSIGKRDDNLVGGMTLDLPSDAPPISLSST 120
P19515 SSPSTTDPEAP--AMSRN-----GPLPS---DVETKYGMLNATSYPDSVQAMS- 95
..*.*:* * *.* * *.* * : **:*.* . . : *

P61871 NSASDGGKVVAATTAQIQEFTKYAGIAATAYCRSVVPGNKWDCVQCQKWVPDGKIITFT 180
P61872 NSASDGGKVVAATTAQIQEFTKYAGIAATAYCRSVVPGNKWDCVQCQKWVPDGKIITFT 180
P19515 ---IDGG-IRAATSQEINELTYTTLSANSYCRTVIPGATWDCIHCD-ATEDLKIITKS 150
*** : ***: :***:* * :*:*:*:* * . * ***.*:

P61871 SLLSDTNGYVLRSDKQKTIYLVFRGTNSFRSAITDI FNFSDYKPVKGAKVHAGFLSSYE 240
P61872 SLLSDTNGYVLRSDKQKTIYLVFRGTNSFRSAITDI FNFSDYKPVKGAKVHAGFLSSYE 240
P19515 TLIYDTNAMVARGDSEKTIYLVFRGSSIRNWIADL FVPVSYPPVSGTKVHKGF LDSYG 210
:*: **.* * *.*:***:***:.* * * * * . * **.*:*** **.*

P61871 QVVNDYFPVVQEQLTAHPTYKVI VTGHSLGGA QALLAGMDLYQREPRLSPKNLSIFTVGG 300
P61872 QVVNDYFPVVQEQLTAHPTYKVI VTGHSLGGA QALLAGMDLYQREPRLSPKNLSIFTVGG 300
P19515 EVQNELVATVLDQFKQYPSYKVA VTGHSLGGA TALLCALDLYQREGLSSSNLFLYTOGQ 270
:* * : ...* :*:. :*:*** ***** **.*:***** **.* ** :* *

P61871 PRVGNPTFAYYVESTGIPFQRTVHKRDIIVPHVPPQSFGFLHGPVESWIKSGTS-NVQICT 359
P61872 PRVGNPTFAYYVESTGIPFQRTVHKRDIIVPHVPPQSFGFLHGPVESWIKSGTS-NVQICT 359
P19515 PRVGDPAFANYVVSTGIPYRRTVNERDIIVPHLPPAAFGLHAGEEYWITDNSPETVQVCT 330
***:*:* ** *****:***:*****:* :*****.* * * * . . . . **:*

P61871 SEIETKDCSNSIVPFTSILDHLSYFDINEGSCL 392
P61872 SEIETKDCSNSIVPFTSILDHLSYFDINEGSCL 392
P19515 SDLETSDCSNSIVPFTSVLDHLSYFGINTGLCT 363
*:*:*.******:*****.* * *

```

Figure 8: ClustalW alignment, the first 26 potential residue in green are the signal peptide, cystein residues correspond to the sites for disulfide bonds. The red box shows the flap region for P61871 and P19515 and the predicted flap region for P61872. The amino acid residues which belong to the catalytic triad are also shown in red (Brady et al., 1990).

Based on comparative sequence analysis and the tertiary structure of P61871, these three catalytic triad residues interact through hydrogen bonds in their side chains as described in previous studies (Kohno *et al.*, 1999). The structure around the serine active center shown in figure 8 and 9 resembles the structure of lipases known as the α/β hydrolase fold (Ollis *et al.*, 1992) with the consensus pentapeptide of β - ϵ -Ser- α motif (Derewenda *et al.*, 1994 a; Derewenda *et al.*, 1994 b), which forms a sharp turn at the active site connecting the β -sheet and an α -helix lid (Figure 9-A-B). The crystallographic structure of *Rhizomucore miehei* (RmL) triglyceride lipase was resolved by Derewenda *et al.* (1992). The RmL (P19515) molecule falls into the category of parallel α/β domains as defined by Ollis *et al.* (1992) and is similar to P61871 sequence. It contains nine beta strands, and all strands are sequential and connected by either β -hairpin loops or right-handed turns involving the β - α - β motif (3-D structure Figure 9-B). Also, Figure 9 illustrates that the tertiary structures of P61871 and P19515 contain six α -helices. The folding of the polypeptide chain in RmL is stabilized by three disulfide bridges: Cys123-Cys362, Cys134-Cys 137 and Cys329-Cys338.

```

P61871      MVSFISISQGVSLCLLVSSMMLGSSAVPVSGKSGSSNTAVSASDNAALPPLISSRCAPPS 60
P61872      MVSFISISQGVSLCLLVSSMMLGSSAVPVSGKSGSSNTAVSASDNAALPPLISSRCAPPS 60
P19515      MVLKQRANYLGFLIVFFTAFLV--EAVPIKRQS---NSTVDS----LPPLIPSRTSAPS 50
          ** . * ::::: .***. :* *:::. ***** ** :.**

P61871      NKGSKSDLQAEFYNMQNTEWYESHGGNLT SIGKRDDNLVGGMTLDLPSDAPPISLSST 120
P61872      NKGSKSDLQAEFYNMQNTEWYESHGGNLT SIGKRDDNLVGGMTLDLPSDAPPISLSST 120
P19515      SSPSTDPEAP--AMSRN-----GPLPS----DVETKYGMALNATSYPDSVQAMS- 95
          ..*.:* :* *.:* * *.* * : ***: .* .: : *

P61871      NSASDGGKVVAATTAQIQEFTKYAGIAATA YCRSVVPGNKWDCVQCQKWVPDGGKIITFT 180
P61872      NSASDGGKVVAATTAQIQEFTKYAGIAATA YCRSVVPGNKWDCVQCQKWVPDGGKIITFT 180
P19515      ---IDGG-IRAATSQEINELTYITLSANS YCRTVIPGATWDCIHCD-ATEDLKIITKTS 150
          *** : *** : **:* * : :*: **:* ** * **:* : * * **.* :

P61871      SLLSDTNGYVLRSDKQKTIYLVFRGTSSFRSAITL IVFNFSYKPVKGAKVHAGFLSSYE 240
P61872      SLLSDTNGYVLRSDKQKTIYLVFRGTSSFRSAITL IVFNFSYKPVKGAKVHAGFLSSYE 240
P19515      TLIYDTNAMVARGDSEKTIYVFRGSSSIRNWIAL LTFVVPVSYPPVSGTKVHKGFLDSYG 210
          :* : ** . * * . : ** : ** : . * . : * . * * . : ** * ** . * *

P61871      QVVNDYFPVVQEQQLTAHPTYKVIVTSHSLGCAQALLAGMDLYQREPRLSPKNLSIFTVGG 300
P61872      QVVNDYFPVVQEQQLTAHPTYKVIVTSHSLGCAQALLAGMDLYQREPRLSPKNLSIFTVGG 300
P19515      EVQNELVATVLDQFKQYPSYKVAVTSHSLGCATALLCALDLYQREGLSSSNLFLYTQGG 270
          :* * : ...* :*: . :*: ** * * * * * * * * * * * * * * * * * * * * *

P61871      PRVGNPTFAYYVESTGIPFQRTVHKRDIVPHVPPQSFGLHHPGVESWIKSGTS-NVQICT 359
P61872      PRVGNPTFAYYVESTGIPFQRTVHKRDIVPHVPPQSFGLHHPGVESWIKSGTS-NVQICT 359
P19515      PRVGDEPAFANYVSTGIPYRRTVNERDIVPHLPPAAFGLHAGEEYWITDNSPETVQVCT 330
          ***:*:* ** * * * * * * * * * * * * * * * * * * * * * * * * * *

P61871      SEIETKDCSNSIVPFTSILDHLSYFDINEGSCL 392
P61872      SEIETKDCSNSIVPFTSILDHLSYFDINEGSCL 392
P19515      SDLETSDCSNSIVPFTSVLDHLSYFGINTGLCT 363
          * : ** . ***** : ***** . * * *

```

Figure 9-A: Sequence alignment of sub-family CI, the secondary structure based on the crystal structures of sequences P61871 and P19515 are available and its shown in bold “flashy green” for α helix and bold red for β sheet. For P61872 sequence secondary structure were based on PSI-PRED prediction. “Light green” indicate α -helices and orange indicate β sheets. The red box shows the flap region for P61871 and P19515 and the predicted flap region for sequence P61872. The blue arrows show the three disulfide bonds.

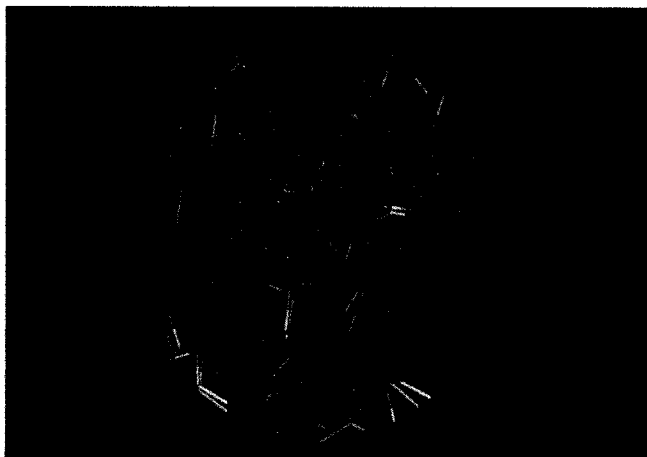


Figure 9-B: The stereo image of comparison between sequences P61871 and P19515 derived from VAST (NCBI). Red lines indicate identical sequences and blue lines indicate similarities.

Lipases of sub-family C.II, P61870 (*Penicillium camembertii*), P61869 (*Penicillium cyclopium*), and O59952 (*Thermomyces lanuginosus*), have the smallest molecular masses in the range of 32-33 kDa, compared to the other true lipases. In contrast to the other lipases, members of this sub-family show maximum activity at a wide range of both pH (from 5.5 to 10) and temperature. The catalytic triad of Ser, Asp and His was conserved at positions 171, 225 and 285 for all three sequences. Unlike sub-family C.I, this sub-family has only two disulfide bridges instead of three, and they are close together. As the disulfide bridges in sub-family C.II were very short in length, they may supply only local structural stability (Figure 10).

A.

```

P61870      MRLSFFTALSAVASLGYALPGKLRDVSSTSELDQFEFVWQYAAASYEADYTAQVGDKL 60
P61869      MRLSFFTALSAVASLGYALPGKLRDVSSTSELDQFEFVWQYAAASYEADYTAQVGDKL 60
O59952      MRSSLVLF--VSAWTALASPIR-REVSQDLFNQFNLFAQYSAAAYCGKNNNDAPAGTNI 56
             ** * : . * ** . : : * : : : : : : : : : : : : : : : : * . * : :
             ┌───┘
P61870      SCSKGNCPVEEATGATVSYDFSDSTITDTAGYIAVDHTNSAVVLAFRGYSYRNWVADAT 120
P61869      SCSKGNCPVEEATGATVSYDFSDSTITDTAGYIAVDHTNSAVVLAFRGYSYRNWVADAT 120
O59952      TCTGNACPEVEKADATFLYSFEDSGVGDVTFGLALDNTNKLIVLSFRGSRSENWIGNLN 116
             : : . ***** : . * . * . * : : . * : : : : : : : : : : : : : : : : * . * : :
             ┌───┘
P61870      FVHTNP-GLCDGCLAEELGEWSSWKLVRDDI IKELKEVVAQNPNYELVVGHSLGAAVATL 179
P61869      FVHTNP-GLCDGCLAEELGEWSSWKLVRDDI IKELKEVVAQNPNYELVVGHSLGAAVATL 179
O59952      FDIKEINDICSGCRGHDGFTSSWRSVADTLRQKVEDAVREHPDYRVVFTGHSLGALATV 176
             * : : . * . * . * . * * * * : * : : : : : * : : * : : * : : * : : * : : * : : * : : * : :
             * : : . * . * . * . * * * * : * : : : : * * : : * : : * : : * : : * : : * : : * : : * : :
P61870      AATDLRGKGYPSAKLYAYASPRVGNAAALAKYITAQ--GNNFRFTHTNDPVPKLPLLSMGY 237
P61869      AATDLRGKGYPSAKLYAYASPRVGNAAALAKYITAQ--GNNFRFTHTNDPVPKLPLLSMGY 237
O59952      AGADLRNGY-DIDVFSYGAPRVGNRAFAEFLTVQTGGTLYRITHTNDIVRLLPREFGY 235
             * : : * : : * : * . : : : : : * : : * : : * * . : : * : : * : : * : : * : : * : : * : : * : :
P61870      VHSVPEYWITSPNNATVSTSDIKVIDGVSFDGNTGTGLPLLTDFEAHIWYFVQVDAGKG 297
P61869      VHSVPEYWITSPNNATVSTSDIKVIDGVSFDGNTGTGLPLLTDFEAHIWYFVQVDAGKG 297
O59952      SHSSPEYWKSGTLVPVTRNDIVKIEG---IDATGGNNQPNIPDIPAHLWYFGLIGTCL- 291
             * ***** . * . . : : . * * * : * : . * . * * : : : : * : : * : : * : : * : :
P61870      PGLPFKRV 305
P61869      PGLPFKRV 305
O59952      -----

```

Figure 10-A: Protein sequence alignment of sub-family C.II

The blue line shows the disulfide bonds and the red box is the flap region for sequence O59952 and the predicted flap regions for sequences P61870 and P61869.

The secondary structure based on the crystal structure of O59952 is available and is shown in bold “flashy green” for α helix and bold red for β sheet. For P61870 and P61869 sequences secondary structure were based on PORTER prediction. “Light green” indicate α -helices and orange indicate β sheets.

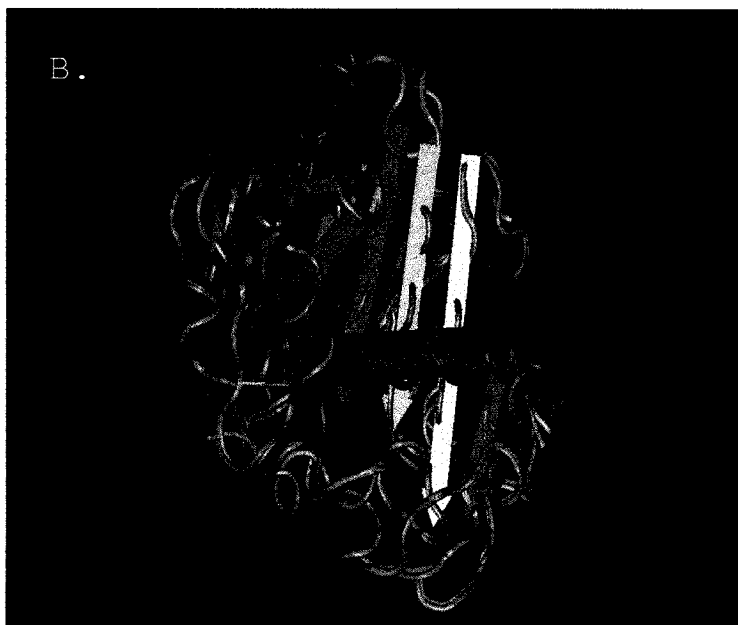


Figure 10-B: The 3D image of O59952 derived from VAST (NCBI).

D.1.4 Group D (Lysophospholipase catalytic domain)

Group D comprises 20 sequences, divided further into five sub-groups (D.I, D.II, D.III, D.IV and D.V). Since no structural information was available for these sequences, they were compared with “human cytosolic phospholipase A2”, a known lysophospholipase structure found in humans-- to find conserved motifs, the potential active site and important structural features.

Sub-family D.I contains three phospholipase B (PLB1, PLB2 and PLB3) sequences, all from *Candida albicans*. They include sequences with accession numbers of Q9UWF6 (PLB1), O93795 (PLB2) and Q9UVX1 (PLB3). The first two sequences have 605 and 608 amino acids with molecular masses between 66-67 kDa. The third sequence has a longer amino acid sequence (754 amino acids) with a molecular mass of 81 kDa. Sequences Q9UWF6 and O93795 contain 17 hydrophobic amino acids and 19 Q9UVX1 amino acids as signal sequence. These signal sequences are believed to target the proteins to the endoplasmic reticulum for subsequent processing leading to the secretory pathway (Leidich *et al.*, 1998). The sequence Q9UWF6 contains seven Asn-X-Ser/Thr motifs at residues 199, 261, 399, 451, 465, 492, and 573 that could potentially be *N*-glycosylated (Figure 11). One potential tyrosine phosphorylation site, Lys-Ser- Asn- Ile Asp- Val-Ser- Ala-Tyr, is shown in Figure 11 (Leidich *et al.*, 1998).


```

Q9UWF6 -----MILHLLILLIINYCVATSPTN----- 22
O93795 -----MLVWQSILLFLVGVLSKSPTN----- 22
Q9UVX1 MKVNLKLIIGSILISQAQAIWFFDSSGSSSSSSDSSPSETGSSGGTFPFDLFGSGSSLTQS 60
      :: : : .. :.*.:

Q9UWF6 -----GYAPGPVSCPSSQL 36
O93795 -----LYTPGYVQCPEGKL 36
Q9UVX1 SSAQASSTKSTSDSASSTDSLFSNSGSSSWYQTFLDGDSGDQKTDYAPFNLTCPSSKKT 120
      *: * : ** :

Q9UWF6 IRSGSQGINPNEQSYINARIPIAKQALSFLHN-ANLQNFVDVDFLAH----SNPTIGLA 91
O93795 TRSSLDGINSNEKAYIDRRYANAKSELSRFLHN-AKMVDFVDVDFLN----SNPTIGLA 90
Q9UVX1 FIRTASELSQQEKDYIHKRQETTNNKLNIDFLSKRANLSDFDAKSFINDNAPNHNITIGLS 180
      . : . : * : * . * : : . * * : * : : * * . . * : * * * * :

Q9UWF6 FSGGGYRAMLTGAGEISSLDSRT-KTNTPLVLAGILQASSYIAGLSGGSWLVGSLASNNLN 150
O93795 FSGGGYRAMLAGAGELLALDSR--ATNPSVLGILQSSSYIVGLSGGSWLVGSLASNDLI 148
Q9UVX1 FSGGGYRAMLAGAGQILGLDGRYEDANKHGLGGLDSSTYVVGLSGGNWLVGSLALNDWL 240
      * * * * * : * * * : . * . * : * * * : * * * : * * * * * * * * :

Q9UWF6 SVDDMLSQG--LWELTHSFLSYGIEHPKQVEEWNVGNQVASKRANFNVS1LTDIYGR 208
O93795 PVDQLLREDK-LWDIQNSLVAYYGVN-IVRNTAMWGNINLQVQTKQLAGFTVSI2TDVYGR 206
Q9UVX1 SVGDIVNGKSTIWQLQDSILNPSGMR-IDKTIAYYYGLAQAVQAKEDAGFQTSV3TDTWGR 299
      . * : : : : * : : . * : : : : * : * . * * . * : * * * * :

Q9UWF6 LLSYPLLTNTED--EGDAYLWSDVTSASNFSQHQMPFPILISDGRAPDTTIINL4NSTVIE 266
O93795 ALSHQLLTNFDN--QGASFLWSDVTETTSFQNNEMPYPILAALGREPNTVLMN5NSTVFE 264
Q9UVX1 ALSYQFFEEEDSGTGGANITWSSIRNLS6SFQDHSMPYPIVVANGRTPGTYIIN7NSTIFE 359
      * * : : : : . * * * : : . : * * : * * * : : * * * * : * * * * : *

Q9UWF6 LTPYEFGSWDP8SLNEFVDTRYLGT9KL10DNGRP-TGK-CYNGFDNAGFFMGTSSALFNEAVL 324
O93795 LTPYEVGSWDP11SLRSFVDTKYIGTRLDDGAP-VSKRCVNGFDNAGFFMGTSSLFNIVLQ 323
Q9UVX1 ISPYELGSWDP12SLKSF13SNIQYLGSSV14NNGNPN15NTD16ICVNNFDNAGFIMGTSSSLFNQILL 419
      : : * * . * * * * . * : : * * : : * * * * . * * . * * * * * * * * :

Q9UWF6 SITEANIP17FLKDIIDDILVDPILKSNID18VSAYNP19PF20IKSSGS-NTAISQSKNLYLVDG 383
O93795 QLNNM21PIPPFLKELISKFTLDPVEKLNID22IAQYNP23PF24IKSNNS-DTKIAQSRTLYLADG 382
Q9UVX1 QLDNYSINSI25IKMILEKVLTD-VSDEEYDIAV26YEP27PF28IGADSAGIKSIT29NTD30LYLCDG 478
      . : : * . : * : : . . * . : : * . * * * * : . : . * : . * * * *

Q9UWF6 GEDGQNIPISP31LLH--RNVSAIFAFD32NSNDV33LN-WPDG34TSLVKTYERQFSSQNGIAFPY 440
O93795 GEDGQNVPLLPLIH--RKVSAI35FAFD36QSADK37NN-WPDG38SALIK39TFERQFSSQGDGIAFPY 439
Q9UVX1 GEDLQNVPFYPLIQ40KRGVDV41IFAFD42NSADT43NSSWPN44GT45SIQETYK46RQF47SKQ48GK49TP50FP51 538
      * * * * * : * * : * * . * * * * * * * . * * * : : : * * * * * * * * :

Q9UWF6 VPDQY52TFRN53L54TSK55PTFFGCD56AK57N58LSL59T-----KDIYD60VPL61VIY62LANR63P64F65Y66W67S68N69T 493
O93795 VPDQNT70FRN71L72TSK73PTFFGCD74AK75N76LSL77T-----ENIYD78V79PV80VIY81LANR82P83F84Y85S86N87I 492
Q9UVX1 APDYK88TFLDK89NMGDK90PVFFG91CN92SSD93LEDLVAWHENDKIN94V95TD96VPL97V98Y99TSN100TRMS101Y102NS103N104F 598
      . * * * : * : * * . * * * : : * * . * : : * * * * : * * * * : * * * *

Q9UWF6 STFKLYDDNERQGMISNGFEIATRSSGSLDDEWAACVGC105AIIRREQERQ106GIEQTEQ107CKR 553
O93795 STFKLKYS108DT109ERQGMISNGYDVASRLNGKLDNEWAACVGC110AIIRREQERL111GIEQTEQ112CKK 552
Q9UVX1 STFKLSYSDQEKFGAIRNGFETVTRN113L114TDEN115W116STCVGC117AIIRRQERL118GEEQ119SDECK 658
      * * * * . * * : * * * * : : * * . * : : * * * * : * * * * * * * * :

Q9UWF6 CFENY120CWDG-----TIYKGEPLGEN121FSDDGLTNSATEYNSNNVAGFNDGGTSILKK 604
O93795 CFENY122CWDG-----TIYKGEPLGD123NFSDEGLT124TSAAAYNSNNVAGINDGGIALV125KR 603
Q9UVX1 CFQEY126CWTGGFKDAASVSSVSGISGLAAK127HTSGGTSST128QQTSTTTGSSANGGSSSTGS 718
      * * : * * * * : : . * . : . * * * : . * . . . . * * :

Q9UWF6 A----- 605
O93795 DDLSN----- 608
Q9UVX1 SSSSKKKN129GGDLVNGGV130PSSI131FLVFN132LLGLIIAYL 754

```

Figure 11: Amino acid sequence alignment of lysophospholipase sub-family DI. Residues in red indicate the active site, and residues in pink indicate potential *N*-glycosylation sites. The red box shows potential tyrosine phosphorylation site with the Tyr residue shown in a black background.

Sub-family D.II includes three diverse phospholipases B, designated as PLB1, PLB2 and PLB3 from different species (Table 3). *Saccharomyces cerevisiae*, one of the species, has three phospholipases B: PLB1, PLB2, and PLB3. *Candida glabrata* the other species has two phospholipases B: PLB1 and PLB2, with only one phospholipase B (PLB) found from *Torulospora delbrueckii* and *Kluyveromyces*. The phospholipase B encoded by the *Saccharomyces cerevisiae* PLB1 gene catalyzes the deacylation of glycerophospholipids at both the *sn*-1 and *sn*-2 positions. Moreover, the PLB1 gene product exhibits phospholipase B, activity catalyzing the deacylation of phospholipase B and transacylase activity which catalyzes the transfer of an acyl chain from one phospholipase B to another and allocating the resynthesis of glycerophospholipids (Figure12 adopted from Fyrst *et al.*, 1999). Fyrst (1999) identified a gene, PLB2, which is functionally similar to PLB1, but contains no significant transacylase activity. Three highly similar genes (PLB1, PLB2 and PLB3 with >60% identity at the DNA level) that code for enzymes with diverse properties were found to exhibit three activities:

- 1, 2-acylhydrolase activity and diacyhydrolase activity on diacylphospholipids.
- Acylhydrolase activity on monoacylphospholipids.
- Acyltransferase activity on monoacylphospholipids to form the corresponding diacylphospholipids (Sreenivas *et al.*, 1998).

All three forms of the enzyme may be derived from a common protein component by using differential glycosylation (Sreenivas *et al.*, 1998). At their pH optimum (2.5-3.5), substrate preference is similar for PLB1p and PLB2p [PtdSer (phosphatidylserine)> PtdIns (phosphatidylinositol)>>PtdCho (phosphatidylcholine) >PtdEtn

(phosphatidylethanolamine)], whereas PLB3p accepts only PtdIns and PtdSer as substrates (Merkel *et al.*, 1999). Preference of PLB3p for PtdIns and PtdSer at low pH is because of the presence of histidine at the substrate-binding site of the enzyme. PtdSer and PtdIns (which is the most abundant anionic phospholipid of the plasma membrane) are tightly segregated to the internal leaflet of the plasma membrane in most cell types (Williamson *et al.*, 1994; Zwaal *et al.*, 1997). At pH values of 5 and above, the substrate preferences change to PtdCho=PtdEtn for PLB1 and PtdSer=PtdEtn for PLB2 (Merkel *et al.*, 1999).

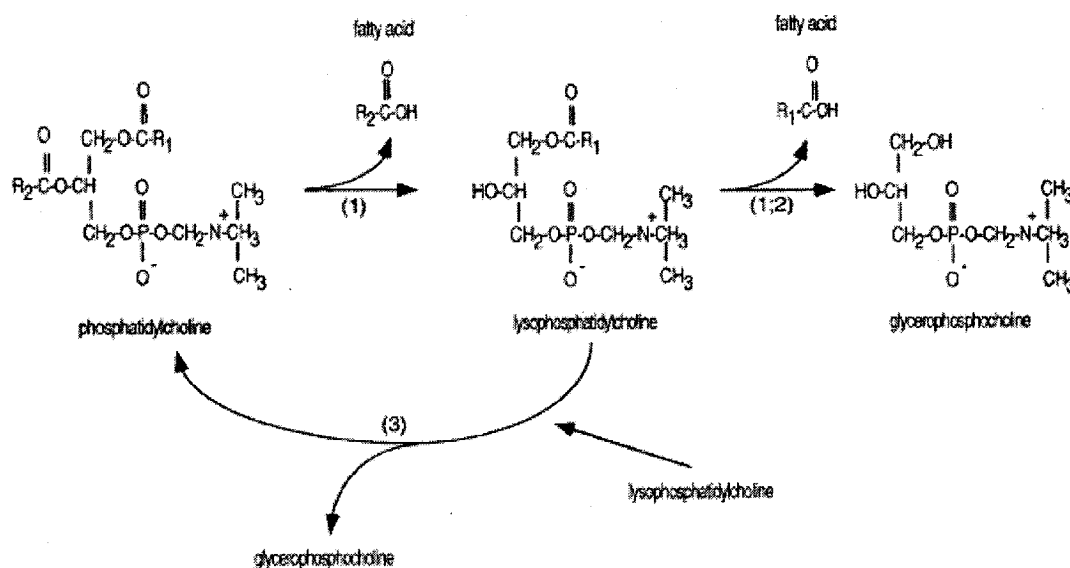


Figure 12: PLB1p activity toward phospholipid substrates containing a choline head group. Phospholipase B (1), lysophospholipase (2) and transacylase (3).

There are nine potential N-glycosidic (N-X-S/T) sites conserved in group D.II (Figure 13). Phospholipase B was reported to be glycosylated with these N-glycosidic linkages. Moreover, the lipase consensus sequence, Gly -X₁- Ser -X₂- Gly and two amino acid residues, Arg 112 and Asp 406, are comparable to catalytically essential residues of

cytosolic phospholipase A2 and are conserved in all sequences in this group (Appendix 2). Since no crystal structure of fungal phospholipase B is available, the functional residues are predicted by comparing to the phospholipases A2, which belong to lysophospholipase catalytic domain family.

```

Q11121      ---MNLKEWLLFSDAVF--FAOQTLAWSPS-NSYTPANVSCD-EDINLIR-QASGSPSDNE 52
P39105      ---MKLQS-LLVSAAVLTSLTENVNAWSPN-NSYVSPANVTCDD-NDINLVR-EASGLSDNE 53
Q8TG07      ---MQLQD-LVVTVSLLAAFNGGVEAWSPT-NSYVSPANVTCPC-NDINLLR-NATGLSQSE 53
O59863      -MWFLLNSVNLFLVCSVALHLDVAWNAWSPT-NGYAPGVVDCD-ENINLVR-KADAVSDDE 56
Q08108      MIRPLCSKIIISYIFAISQFLLAANAWSPT-DSYVPGTVSACP-DDINLVR-EATSISQNE 57
Q03674      ---MQLRN-ILQASSLISGLSLAADSSSTTGDGYAPSIIPCPSDDTSLVR-NASGLSTAE 55
Q8TG06      -----MQLSVLIASVLAAGAAVDAA--SYTPQNVSCPC-DNANFIRNAADGLSPA 47
           :      .      : ..      . * . * : * : : . : * * . * * *

Q11121      TEWLKRRDVYTREALRSFLDRATSNFSDSS----LVSQLF-S-NASDIPRIAVACSGGGY 106
P39105      TEWLKRRDAYTKEALHSFLNRATSNFSDTS----LLSTLFGS-NSSNMPKIAVACSGGGY 108
Q8TG07      IDWLKRRDVNTREALSFLKRVTSNFSTSNSSASNLIDQLFST-NSSNIPKIGIAASGGGY 112
O59863      ADWLKVRHESTVPALKDFLQGRGFKGFTNDTS---IIDKLLAT--QDTAPKVAIACSGGGY 111
Q08108      SAWLEKRNKVTSVALKDFLTRATANFSDSSE---VLSKLFNDGNSENLPKIAVAVSGGGY 114
Q03674      TDWLKRRDAYTKEALHSFLSRATSNFSDTS----LLSTLFSS-NSSNVPKIGIACSGGGY 110
Q8TG06      KEWLKRRDPITRDALQTLRRRAFANVSTEIT-----SALFND--TENVPKLGIAVAGGGY 100
           * * : * . * * * * * * * . : : * * : * : : * * : * * *

Q11121      RAMLSGAGMLAAMDNRTDGANEHGLGGLLQSTTYLAGLSGGNWLVTGLAWNNWTSVQDIV 166
P39105      RAMLSGAGMLAAMDNRTDGANEHGLGGLLQCATYLAGLSGGNWLVTGLAWNNWTSVQAI 168
Q8TG07      RAMLSGAGMVSAMDNRTDGANEHGLGGLLQAATYLAGLSGGNWLVTGLSWNNWTSVQDI 172
O59863      RAMLSGAGMISAMDNRTDGANEHGLGGLLQSSSTYLAGLSGGNWLVTGLAYNNWTSVQAI 171
Q08108      RSMLTGAGVLAAMDNRTEGAYEHGLGGLLQSTTYLSCASGGNWLVTGLALNNWTSVQDI 174
Q03674      RAMLGGAGMIAAMDNRTDGANEHGLGGLLQSSSTYLSGLSGGNWLVTGLAWNNWTSVQEI 170
Q8TG06      RAMFVGAGAFAMDNRTDGANEHGLGGLLQAATYMAGLSGGNWLVTGLAYNNWTSVQQIL 160
           * : * : * * * . : * * * * * * * * * * * * * * * * * * * * * * * *

Q11121      NNMTEDDSIWDISNSIINPGGFMIVTTIKRWDHISDAVEGKQDAGFNVSRLTDIWRALSY 226
P39105      DNTTESNSIWDISHSILTPDGINIFKTGSRWDDISDDVQDKKQDAGFNISLADVWGRALAY 228
Q8TG07      DSQDNDSAIWDISHSIVSPGGINIFKTGSRWDDISDAVEDKQKAGFNVSRLADVWGRALSY 232
O59863      NNMTEDDSIWDISNSIIVNPGGINIFSSISRWDDISDAVEEKKKAGFNISITDVWGRALSY 231
Q08108      NNMQNDDSIWDLSDSIVTPGGINIFKTAKRWDHISNAVESKQONADYNTSLADIWRALAY 234
Q03674      DHMSEDSISWNITKSIVNPGGSNLTYYTIERWESIVQEVQAKSDAGFNISLSDLWARALSY 230
Q8TG06      EEGDKADAIWNTITNSFLNPNYDKFSKTLARWTAIGSQVQGRDAGFNVTITDLWSRALAY 220
           :      . : * * * * * * * * * * * * * * * * * * * * * * * *

Q11121      NFFPSLYRGGVAYTWSTLRDVEVFQNGEMPFPISVADGRYPGTQIIDLNATVFEFNPFEM 286
P39105      NFWPSLHRGGVGYTWSTLREADVFKNGEMPFPITVADGRYPGTVINLNATLFEFNPFEM 288
Q8TG07      QFFPTLYRGGVAYLWSDLRESDFKNAEMPMPISVADGRYPGTAVIDLNSTVFEYSPFEL 292
O59863      NFFPSLDEGGVGYTWNTLRDVDVFKNGEMPFPISVAVGRYPGTQVVNLNATVFEFNPFEM 291
Q08108      NFFPSLNRGGIGLWSSIRDFPFVQNAEMPFPISVADGRYPGTVINLNATVFEFNPFEM 294
Q03674      NFFPSLPDAGSALTWSSLRDVDVFKNGEMPLPITVADGRYPGTVINLNATLFEFTPFEM 290
Q8TG06      GWFPPLPAGAGLTWSSLRDNIEFMNGEMPMPISVADGRYPGTVINLNATVFEFTPFEM 280
           : : * * * * . * . * . * : * * * * * * * * * * * * * * * * * *

Q11121      GSWDPTLNAFTDVKYLGTKVSNGEPVKNKQCVAGYDNTGFIMGTSSSLFNQFLLQINSTS 346
P39105      GSWDPTLNAFTDVKYLGTNVNNGKPVKNKQCIAGFDNTGFITATSSSLFNQFLLRLNSTD 348
Q8TG07      GSWDPSLSAFTDVQYLGTVSDGKPAEEGKCIAGFDNVGFLMGTSSSLFNQFLLRINDTS 352
O59863      GSWDYTLHTFTDVRYAGTNVNTGTPNVTGKCVAGFDNTGFVMGTSSSLFNQFLLQNTTD 351
Q08108      GSWDPSLNSFANVKYLGTVNNGVPLERKCTAGFDNAGFIMGTSSSLFNQFLLRINSTH 354
Q03674      GSWDPSLNAFTDVKYLGTNVNNGKPVKNKQCVSGYDNAGFVIATSASLNFNEFSLEASTST 350
Q8TG06      GSWDPSLNAFSDIKYLGTVTDGKPE-TERCINGFDDASFIMGTSSSLFNFTMSNDSAV 339
           * * * * * * * * * * * * * * * * * * * * * * * * * * * * * *

Q11121      LPSFIKNLVTGFLDDLSEDEDDIAIYAPNPFKDTSYIQDNFSKSISES DYLYLVDGGEDN 406
P39105      LPSFIANLATDFLEDLSDNSDDIAIYAPNPFKEANFLQKNATSSII ESEYLFVLDGGEDN 408
Q8TG07      IPKFIRNLATHFLKDLSEYDDIAVYAPNPFRRADYVNNNRSKSLSESEYLFVLDGGEDG 412
O59863      LPSFLYNLLHGFLTDDASDDYDDISIWAPNPFYEITNIPSNYSQSISEDDTLYLVDGGEDG 411

```


Subfamily D.III includes two sequences from *Ascomycota* phylum P39457 (*Penicillium notatum*) and O42790 (*Neurospora crassa*). The primary structure of P39457 (Masuda *et al.*, 1991) having 603 amino acids in the main chain (9 amino acids in the signal peptide) was compatible with the primary structure of O42790 (634 amino acids and 19 amino acids known as signal peptide sequence clustering with it as shown in Figure 14). For example, the *pI* value of both native and modified phospholipase B in this group (D.III) was pH 4.0 (Saito *et al.*, 1991; Kawasaki *et al.*, 1975). Moreover, sequence P39457 (*P. notatum*) contained 63 acidic amino residues in the main chain (44 Asp and 19 Glu) and 41 basic residues (17 Arg, 8 His and 16 Lys), and sequence O42790 had 62 acidic amino acid residues in the main chain (38 Asp, 24 Glu) and 49 basic acids (24 Arg, 1His and 24 Lys).

From the eight cysteine residues in both sequences, two cysteine residues were known to make a disulfide bond. *N*-glycosylation sites (Asn-X-Ser/Thr motifs) of phospholipase B from both sequences were distributed along the whole sequences at fourteen residues 41, 81, 116, 150, 223, 267, 306, 335, 427, 440, 446, 477, 498, 526, 532, 567, and 571 in the sequence from *P. notatum* (Figure 14) and were conserved in ten residues at the sequence O42790 derived from *Neurospora crassa* (Okumura *et al.*, 1981; Masuda *et al.*, 1991).

The subfamily D.IV includes five sequences from the fission yeast, *Schizosaccharomyces pombe*. These five sequences contain sequences with accession numbers O13857 (missing signal peptide), Q9P327, and Q9Y7N6 (19 amino acids as signal peptide) and one lysophospholipase with accession number Q9UTH5 (19 amino acids as potential signal peptide) and sequence P78854 with 21 amino acids as signal peptides. Similar to other phospholipases, these sequences contain the conserved Arg-Ser-Asp triad that may form part of the interfacial recognition site (Figure 15). The consensus motif (Gly -X₁- Ser -X₂- Gly /Ser) similar to true lipases were also conserved in this group but in sequences Q9UTH5 and P78854 the serine residue was replaced by an aspartic acid residue (Figure 15). Moreover, at the third position of the putative catalytic site, the aspartic acid residue was conserved in all five sequences. According to Brenda (A Comprehensive Enzyme Information System; <http://www.brenda.uni-koeln.de>), the pH value for optimal activity of these five lipases is 2.5 and no activity was detected at neutral and alkaline pHs. These enzymes were not heat-stable and showed a temperature optimum of 35 °C.

```

O13857      -----MRPG-----MHDTPLSLMQK-----RE 17
Q9P327      --MYVNYIGLEAFVQISLTLAYPPGRVEISEIYDFEESSYKQDIDTSVLYT-----LS 53
Q9Y7N6      ---MKLSSFGLFLALQLLPALGLPSR-----IDEVDVSDPELIGLLKPDNVDKP 46
Q9UTH5      -----MYFQSEFYFLALLLATAVYGO-----VASPELHLSLSR-----R 32
P78854      MLFRGLSLWMLFLASCLSAALPAAE-----DDGSVKVFKR----- 36
          .
          :
O13857      ALAISLSKRDSVGSYAPYNVTCPS-DYMLRPASDG-ISSGEQSFIDKRIPKINTQMRSEFI 75
Q9P327      KRKPALVKRSTDASYAPFNVTCSN-DNLLRPASEG-LNEGEQSYINKRISKVNSELRSFI 111
Q9Y7N6      ANSIPLSKRSTSPSYAPYTVACPS-GSLLRPASDG-LSTGEQEFVDKRVSKVNSALESFI 104
Q9UTH5      NWKKPPFPSTNASYAPVIRSCDSSEIMVNSLPRGELPDLENDFIEKRLSNANEALTTFL 92
P78854      --AKKHSTKQEGPSYAPYYVDCPS-DNIVESLSSNEIPSAESEYLSTRSTITNTAMKDFL 93
          .   ****   *   .   :... . . :   *...*. . *   :   *:
O13857      S--NTGLDVDVNSVINDSDGPRGLGAFSGGGLRAMVHGGGVLNAFDSRNGNGSSLAGILQ 133
Q9P327      S--KTGLNVDLTKVVNSDGPRGLGAFSGGGLRAMVNGGGAFNAFDSRFESDSPLSGLLQ 169
Q9Y7N6      S--KTGLKIDTKSVLNDTDGPRGLGAFSGGFPAMLTGAGAINAFDARNGNTTSLGGILQ 162
Q9UTH5      QSKNTTADLDLSSIVGD-NGPRLGIAVSGGWRSMLFGGGALAALDSR-SNETTLGGLLQ 150
P78854      R--NANLPGLNADTLSGSEGPSIGIALSGGGLRAMILGSGALSAMDARHDNHTVLTLGLLQ 151
          ::   . . . :** :*:*.**** :*: *.*.: **:* . : * **
O13857      SAMYIAGLSGGSWLVGSVAVNNFANITYLRDNVWNLEHSVFAPHGDNVVENLAYDDLDD 193
Q9P327      SAMYISGLSGGSWLVGSVAINNFTNITYLRDNVWNLEHSVFAPHGDNVVENLNLYYNDLRK 229
Q9Y7N6      SSMYITGLSGGSWLVGSVAVNNFANITFLHDDVWNLDHSLFAPY-DDAFENFYIQEWEF 221

```


Subfamily D.V includes two phospholipase B enzymes from *Cryptococcus neoformans* (Q9P8P2 and Q9P8L1). The extra-cellular phospholipase B1 (PLB1) is known as a virulence factor produced by the pathogenic fungus *Cryptococcus neoformans* (Cox *et al.*, 2001). The serine presented in the pentapeptide Gly-Leu-Ser-Gly-(Gly/Ser) of both sequences in group D.V is similar to the serine presented in the “lipase motif” Gly -X₁- Ser -X₂- Gly (Schrag and Cygler, 1997). The catalytic site and the putative catalytic motif of phospholipases, SerGluGluGluXArgAla (Met/Leu), presented in these two sequences are similar to all other enzymes in family D. A group of hydrophobic amino acids at residues 1-20 is known as a signal peptide and thirteen *N*-glycosylation residues in both sequences have been predicted (Figure 16).

```

Q9P8P2    MS IATGTFAFSLFATIAFAVPPETPRIELQAERGLGDKSYAPWQVDCPSNVTWIRNATG 60
Q9P8L1    MS IITTFAFSLSLATTAFAVPPETPRIELQAERGLGDQSYAPWQVDCPS  WIR  TG 60
          *** .:***.***:*.*****.*****.*****

Q9P8P2    LGSGERAYIEAREKLVQPVIEQMMAARGLETTPRTPNIGVALSGGGYRAMLTGLGGIMGM 120
Q9P8L1    LGTGERAYIEAREKLVQPAIEQMMAARGLETTPRTPVIGVALAGGGYRAMLTGLGGIMGM 120
          **.:*****.*****.*****.*****.*****

Q9P8P2    MNESTEASESETGGWLDGVSYWAGLSGGSWATGTFMSNGGQLPTNLLLENLWNIDSNLVFP 180
Q9P8L1    MNESTEASQSETGGWLDGVSYWSGLSGGSWATGSFMSNGGQLPTLLENLWNIDSNLVFP 180
          *****.*****.*****.*****.*****

Q9P8P2    DDDKLSFYTELYTETNAKSDLGFPQITD VWGLAIGSHVLPERYQLSNTPNLTFSSLPVS 240
Q9P8L1    DDGKLSFYTNLYTETNAKSDLGFPQITDIWGLAIGSHVLEPEPYQLSNTP  FSSLPVS 240
          **.:*****.*****.*****.*****.*****

Q9P8P2    VSALGNASLPMPIIIAADRKREAGELVIAENATVWEFTPYEFGSWAFGSQYKSPGAFTP 300
Q9P8L1    VAALGNASLPMPIIIAADRKREAGELVIAENATVWEFTPYEFGSWAFGSQYKSPGAFTP 300
          *.:*****.*****.*****.*****.*****

Q9P8P2    IEYLGTSVDDGSP  CWKGFQDLSFVMGTSATLFNGAFLELNGTDSGLLTNLITAFIAD 360
Q9P8L1    IEYLGTSVDDGSPNGTCWKGFQDLSFVMGTSATLFNGAFLELNGTDSGLLTNLITAFIAD 360
          *****.*****.*****.*****.*****

Q9P8P2    LGEDQADISRIPIPNFSFNYSNGENPIYNLTYITLVDAGETNQNIPLEPLLVPTRDVDAIVA 420
Q9P8L1    LGEDQADISRIPIPNFSFNYSNGENPIYNLTYITLVDAGETNQNIPLEPLLVPTRDVDAIVA 420
          *****.*****.*****.*****.*****

Q9P8P2    FDSSYDTDYIWPNGTALRTTYERAKVLAEHENTRVLMPPEVPSMNGFVNGGYNRSRPTFFGC 480
Q9P8L1    FDSSYDSDYIWPNGTALRTTYERAKILAEHENTRVLMPPEVPSMNGFVNGGYNRSRPTFFGC 480
          *****.*****.*****.*****.*****

Q9P8P2    NDTTTPLIIYVPSYPWSFAANTSTYQLSYENDEANEMLLNGMRSLLTNHSVPTWPTCFAC 540
Q9P8L1    NDTTTPVIIYIPSYPWSFAANTSTYQLSYENNEANEMLLNGMRSLLTNHSVPTWPTCFAC 540
          *****.***.*****.*****.*****

Q9P8P2    ALDRSFMYTSENRSSTTCQKCFDTCWAGDDNTTEPATYEPVINSVPPWLVANNLSIGVA 600
Q9P8L1    ALDRSFMYTSENRSSTTCQKCFDTCWAGDDNTTEPANYEPVINSVPPWLIANNLSIGMA 600
          *****.*****.*****.*****.*****

Q9P8P2    DAPASNESTAGTASSGAANADVSMGMVALAAGLGLML 637
Q9P8L1    DAPGSNESTAGTASSGAAKMGVGMGMVALTAGLGLML 637
          ***.*****.***.*****.*****

```

Figure 16: Multiple sequence alignment of sub-family D.V. The *N*-glycosylation sites (NXS/T) and the predicted *N*-glycosylation sites are also shown in light pink. The conserved active-site is shown in red.

D.1.5 Group E (*ab*-hydrolase)

Branch E which comprises only one sequence (P34163) belongs to the family “*ab*-hydrolase” (<http://www.sanger.ac.uk/Software/Pfam>). Among the fifteen *ab*-hydrolyase sequences in the SWISS-PROT database, only one sequence, P34163, is from fungi (*Saccharomyces cerevisiae*). This sequence has 548 amino acids with a molecular mass of 63 kDa (The sequence is not shown here). This sequence is also called TGL1 (Triglyceride lipase-cholesterol esterase). It has been suggested that TGL1 is a triglyceride-specific lipase on the basis of its homology to lipases from humans and rats, but its enzymatic activity against triacylglycerol has not been demonstrated yet (Abraham *et al.*, 1991).

D.1.6 Group F (*Phospholipase C*)

The three sequences P32383, O13433, and P40977, clustered in group F, are the only biochemically characterized fungal phosphoinositide phospholipase C (PLC) sequences available at SWISS-PROT. The remaining sequences are of human, rat, mouse and bovine origin. The PLC's are large enzymes with 869-1099 amino acids and molecular weights in the range of 100-124 kDa (Figure 17). Group F includes sequences with an EC number of 3.1.4.11. The digit “4” corresponds to Phosphoric diester hydrolases and “11” corresponds to Phosphoinositide phospholipase C (PI-PLCs). The two most highly conserved domains have been designated as X and Y and form the catalytic core of the molecules (Figure 17). Also, a common feature identified at the C-terminus of the Y region is a C2 domain, which is sometimes known as part of an extended Y domain.

P32383 -----MTESAI DDQRFNLTKE LQRHSCRDQG----- 26
O13433 ----MLES LNRRNSIDSNQADNDNDNDNHN S NDELS PSELYYSPSGSPPK S QLLLRKSSSP 56
P40977 MNC EMVTSPE SVNLGDSNR AVSPFCLPDCSENAVAQTR-----SKTLDNALD 48
. * . : : . * .

P32383 -----KITQKDDALDFISYSS FQSSFN TDQKSANNGSTVRRSIR SIFRRA 71
O13433 SSYSPIKSDLPNIYSHLR SNDS EPPQSPKQSS LSSSSSSSSSSSNTK S STTKNIFKKL 116
P40977 LPYVGNRKKSEQDFFKMLSSRDRDAHSTLRKRSNSLSSFLSTKSTSA SENKFHGGLNWLS 108
: : * : : : : : : :

P32383 AELPR-----VHMGPLTYSHGIN----- 89
O13433 LRINKSSDNIDESRSIVSNNGGSPMSDSTTVTSTLSTDTAPKRGKSIQRSQILHHTDSDS 176
P40977 LKLNLLRLQGRMNSARTNTSMNPYSCDSNEN----- 140
. : . * : . .

P32383 -----ELVNKKLRKDCDLSTLCRVLQRGIRMIRMT RRRRKFYEFKLINNGQIIWKDG 142
O13433 LYLENQIELRPEISKSIGNIKIPSI FTNDGMPL LKISHKSKRILFWIDPSCFKFSWRMA 236
P40977 --LSTLSSVQNFNRSQ L L ATIVPESIQNGCSLLRITKKKVRQRKVSLDPI S GYLM LDKN 198
. : : . . : * : : : : : : :

P32383 -----SKYLELD 149
O13433 NSTTTTTSATTSATTSGLPQGITNTALSNSAIISTPAIATS AIHRLSITNRTTHEFVLD 296
P40977 -----TGKAYKKLCVD 209
: : : *

P32383 SVKDIRIGDTASTYQEEVDPKRLRSDSKLWIAI IYKVS NKLKALHVVALNELDFNTFLSC 209
O13433 DIKSIYIQNEGSGYREELNISQKLEKNWIT I IYFNHKKNSLKS LHLITDNDHDFKKLISA 356
P40977 DIKEIRQGRDARNYREQYKISSENER--WFTI IYCADNKLKAMHMI SPTLDAHNQWIMA 267
.: * . * : : . . : : . * . * : : : : . : :

P32383 ICGLVKLRRELMESILPD---NSQFARIHWQITVSE-----KEEDEK KDT 252
O13433 IYNLQQLRSQ LAKEFLIDLNELDENYVKMLLNKELLAGDNGNVDGNEVDIRKSHKHVREF 416
P40977 LEGLKTYR---LNEFI IGLNLVCHQEK MIDYSENLN-----PWEKLEKQSAQ 313
: . * * : : : : : : :

P32383 LSFADVKKLCKDFHIYVSTGQLLEFFQLADINHNG-----LLNYFEFEKFIK 299
O13433 LSFNDILKYSKRLNINVNTNHLQQIFDQVLLSSATEKPVSTPLFEKGLNFEQKFQFVS 476
P40977 LDLDGVHRMCQMLHLNASMEFLEET FQKADADHSG-----KLSFEFQHFVS 360
* . : * : . . : : . . * : * : . . * : : * : :

P32383 ILKNRKEVNMIWSKFTKPPHSHLSFENFFQFLITEQHEQVDRQTAW S-----YFIKY 351
O13433 ILKDRKDLQEIWDSLAQG-KEVLQFDEIKNFI INIQKENFSDDDDNSTINLIFQKYCSND 535
P40977 LLKTRSEIVDIFKEYTSG-SDKMSLEQFRHFLSTSQKARLDSDSIRT-----LYVSF 411
: * * : : * : . . : : : : : * : . . : :

P32383 REPTQLTMGQDGFTKFLK-EQPYLVEVKEEL [REDACTED] 410
O13433 NGWNKESLNEYLLSSYST-PYREITQTQNY [REDACTED] 594
P40977 CSNDDSKMGLIEFTSFLSPHN S PVPVI [REDACTED] 471
. . . : : : . : . * * . * : * : * : * : * : * :

P32383 [REDACTED] 450
O13433 [REDACTED] 654
P40977 [REDACTED] 512
* * * : * : * : * : * : * : * : * : * : * : * : * : * :

P32383 [REDACTED] 508
O13433 [REDACTED] 714
P40977 [REDACTED] 572
* * . * : * : * : * : * : * : * : * : * : * : * : * : * :

P32383 [REDACTED] KTSEATRGLSVNEP---FPSSFSSSYESANEQLRMKDDSTNSSSATN 565
O13433 [REDACTED] KTSFQNL IETENG SFTTSTTTTTTTTTTTTTATSLSEDNENKNKSNSS 774
P40977 [REDACTED] CSATPLHQFSTDLKVGITDSSDTTTESELENS ELTG----- 622
: * : * : * : * : : : : : : : : : : :

P32383 SSSMQRIGRIGLKKHADIINDVSN [REDACTED] 624
O13433 STSSFII RRR-KNKSPKIIN [REDACTED] 833
P40977 -----LRKGKRRMKNIIVQEL [REDACTED] 676
: : . * : : : : : : : * : * : * : * : * : * :

D.1.7 Group G (Phospholipase D)

The fungal lipase P36126 from *Saccharomyces cerevisiae* (Baker's yeast) is on a branch closely related to group F. This is a long protein (1380 amino acids) with a molecular mass of 160 kDa. P36126 has two domains known in all PLD1 and PLD2 enzymes, including PX and PH domains. The analysis of 3-D structure of several PH domains suggests a structure consisting of two perpendicular anti-parallel β sheets, followed by a carboxyl-terminal α -helix along one end of the barrel (Ferguson *et al.*, 1995; Ferguson *et al.*, 1994). The loops connecting the β -strands differ greatly in length, making the PH domain difficult to detect. The PX domain contains several conserved positively charged and hydrophobic residues characteristic of domains known to bind to specific phosphopeptides or phospholipids.

D.1.8 Other lipases

D.1.8.1. Triacylglycerol lipase 2 (P54857)

The lipase P54857 from *Saccharomyces cerevisiae* with an EC number of 3.1.1.3 does not show the classical consensus core of Gly -X₁- Ser -X₂- Gly but rather displays the Ala-His-Ser-Met-Gly core -- the first Gly is replaced by an alanine residue (Dartois *et al.*, 1992). A BLAST search on EXPASY/SIB shows that this sequence is more similar to the lipases from the bacterium *Pseudomonas* species than fungal lipase (Figure 18). Based on the sequence similarity with the lipase sequence from *Pseudomonas*, the catalytic site (not shown in the SWISS-PROT database) has the conserved serine and glutamic acid residue yet there is no histidine residue found in this sequence. Lipase P54857 has a molecular mass of 35 kDa with the sequence length of 326 amino acid and

shows peak activity at pH 8.0 -- similar to active pH values of group B enzymes, which cluster closer to this group.

```

P25275      -MARTMRSRVVAGAVACAMSIAPFAGTTAVMTLATTHAAMAATAPADGYAATRYPIILVH 59
P22088      -MARTMRSRVVAGAVACAMSIAPFAGTTAVMTLATTHAAMAATAPAAAGYAATRYPIILVH 59
P54857      MKNDNKANDIIIDSVKVPDSYKPPK-NPIVFCHGLSGFDKLLILPSVFHLTNLISNSIVH 59
              . . . : : * . * * . . * : . : * : : . . : **

P25275      GLSG-----TDKYAG--VVEYWYGIQEDLQONGATVYVANLSGFQ--SDDGANRGR 106
P22088      GLSG-----TDKYAG--VLEYWYGIQEDLQONGATVYVANLSGFQ--SDDGPNGR 106
P54857      NMAENFMQDDEDKSDNKYTNLLEIEYWIGVKKFLQSKGCTVITTKVPGFGSIEERAMALD 119
              . : . . . . . : ** . : *** * : : * : * : * . . . . . * : : . . .

P25275      EQLLAYVKTVLAATGATKVNLYGHSQGGITSRVVAAPD---LVASVTTIGTPHRGSEF 163
P22088      EQLLAYVKTVLAATGATKVNLYGHSQGGISSRYVAAPD---LVASVTTIGTPHRGSEF 163
P54857      AQLQKEVKKIESKDKRHSNLIAHSMGGLDCRYLICNIKRNRYDILSLTTISTPHRGSEM 179
              ** * : : . : ** . * * * * . * : . . : * : * * . * * * * * :

P25275      ADFVQNVLAYDPTGLSSSVIAAFVNVFGILTSSSHNTNQDALAALQTLTTARAATYNQNY 223
P22088      ADFVQDVLAYDPTGLSSSVIAAFVNVFGILTSSSHNTNQDALAALQTLTTARAATYNQNY 223
P54857      ADYVVDLFEN-----LNALRVSQKILP-----ICFYQLTTAYMKYFNLVT 219
              ** : * : : . : * : * . . : * * * * : *

P25275      PSAGLGAPGSCQTGAPTETVGGNTHLLYSWAGTAIQPTLSVFGITGATDTSTVPLVDLAN 283
P22088      PSAGLGAPGSCQTGAPTETVGGNTHLLYSWAGTAIQPTLSVFGVTGATDTSTLPLVDPAN 283
P54857      PNS-----PKVSYFSYGCSFVPKWYNVFCTPWKIVYERSKGCPCNDGLVTINSSK 268
              * . : . . . . . * . . * * : . . : : : :

P25275      VLDPS-TLALFGTGTVMINRSGQNDGLVSKCSALYGKVLSTSYKWNHLDEINQLLGVRG 342
P22088      VLDLS-TLALFGTGTVMINRSGQNDGLVSKCSALYGKVLSTSYKWNHLDEINQLLGVRG 342
P54857      WGEYRGTCLKMDHLDVINWKNKLODD-----WSKFFRTTTVGEKVDILNFYLKITD 319
              : . * * . : * : . . * * . . . . . * : * : * * * * * :

P25275      AYAEDPVAVIRTHANRLKLAGV 364
P22088      AYAEDPVAVIRTHANRLKLAGV 364
P54857      DLARKGF----- 326

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Figure 18: Multiple sequence alignment of *Saccharomyces cerevisiae* (P54857) with two bacteria sequences from *Pseudomonas* species (P25275 and P22088).

D.1.8.2. CALB (P41365)

Candida antarctica lipase B (CALB) (with a molecular mass of 36 kDa), is among the smallest lipase enzymes known in fungi. Lipase P41365 (*Candida antarctica*), like most other fungal lipases, is from the *Ascomycota* phylum. A BLAST search of this sequence shows striking similarity to lipases from the *Basidiomycota* phylum. This aspect was observable in this case and it was placed near to the sequences Q9P8L1 and Q9P8P2; both of which are sequences from *Basidiomycota* phylum. Lipase P41365 from *Candida antarctica* (3.1.1.3), is similar to all other triacylglycerol lipases at one or more of the three ester bonds of these triacylglycerols substrates (IUBMB, 1992). The amino acid sequence similarity, according to the UPGMA dendrogram, shows no significant homology to other lipase sequences. The catalytic triad, as mentioned by Uppenberg *et al.*, (1994), contains the Ser-Asp-His sequences. Conversely, the consensus sequence found in lipases around the active site serine, Gly -X₁- Ser -X₂- Gly, is not present in lipase P41365. This motif has been changed to Thr-Trp-Ser-Gln-Gly by replacing threonine instead of glycine in the first amino acid, which makes this sequence unique compared to the other lipases. Similar to group A (AB hydrolyze), in this sequence glutamic acid, as a second catalytic site, was replaced by aspartic acid. The histidine residue in this sequence is known as the third active site residue in this catalytic triad.

D.2 Evaluation of Secondary Structure Prediction Tools for fungal Lipases

D.2.1 Secondary structure and structural feature prediction family B

Using multiple sequence alignments often enables accurate prediction of secondary structures. Hence, we decided to predict secondary structures by analyzing the predictions made by different tools and techniques. To evaluate the different secondary structure prediction tools, we analyzed all of the sequences with known X-ray crystal structures using fifteen tools available on SWISS-PROT and NPS@ separately, and the output of each tool was compared with the X-ray crystallography based structure. The tool that predicted the structure that was the most similar to the crystallography based structure was chosen as the best predictor of a given group of lipases. The sequences with unknown secondary structures were separately analyzed with the chosen tool. Three sequences in sub-family B.I (P20261, P32947, and P32946) have known 3-dimensional structures based on the X-ray crystallography method. The analysis of these sequences suggested that the tool PREDATOR (Argos *et al.*, 1996) had the highest score (highest score = the most similar to the X-ray structure) for P20261 and P32947 (Figure 21) and PSI-PRED had the highest score for P32946 (Figure 21). Since PREDATOR (Argos *et al.*, 1996) was the tool that had the most similar prediction to the X-ray crystal structure, we used this tool to predict the secondary structure for the P32949 sequence (Figure 19) in sub-family B.I and compared it with the other sequences of known secondary structure (P20261 and P32947). The secondary structure of P32948 was best predicted using the PSI-PRED tool (Figure 19).

```

P32947 MKLALALSLIASVAAAPTAKLANGDTITGLNAINAEFLGIPFAEPPVGNLFKDPVPYS 60
P32949 MKLALALSLIASVAAAPTALANGDTITGLNAINAEFLGIPFAEPPVGNLFKDPVPYR 60
P20261 MELALALSLIASVAAAPTATLANGDTITGLNAINAEFLGIPFAEPPVGNLFKDPVPYS 60
P32948 MKLALVLSLIVSVAAPATLANGDTITGLNAINAEFLGIPFAEPPVGNLFKDPVPYS 60
P32946 MKLCL-LALGAAVAAPTATLANGDTITGLNAIVNEKFLGIPFAEPPVGNLFKDPVPYS 59
*:* *:* :*****.*****.*****.*****.*****.*****.*****

P32947 GSLNGQKFTSYGSPSCMQQNPEGTFEENLGTALDLVMQSKVFQAVLPQSDCLTINVVRE 120
P32949 GSLNGQSFTAYGSPSCMQQNPEGTYEENLPKVALDLVMQSKVFQAVLPNSDCLTINVVRE 120
P20261 GSLDGQKFTSYGSPSCMQQNPEGTYEENLPKAAALDLVMQSKVFQAVLPNSDCLTINVVRE 120
P32948 ASLNGQKFTSYGSPSCMQMNLPLGNWSSLPKAAINSLMQSKLFQAVLPNGSDCLTINVVRE 120
P32946 ASLNGQQFTSYGSPSCMQMNPMSFEDTLPKNARHLVLOSQKIFOVVLPNGSDCLTINVVRE 119
*:*:*:*:*:*:*:*:*:*:*:*:*:*:*:*:*:*:*:*:*:*:*:*:*:*:*:*:*:*:*:*:*:*:*:*

P32947 PGTKAGANLPVMLWIFGGGFEIGSPTIFPPAQMVTKSVLMGKPIIHVAVNYRVASWGFLA 180
P32949 PGTKAGANLPVMLWIFGGGFEIGSPTIFPPAQMVSKSVLMGKPIIHVAVNYRVLASFGFLA 180
P20261 PGTKAGANLPVMLWIFGGGFEVGGTSTFPPAQMITKSIAMGKPIIHVSVNYRVSWSWGFLA 180
P32948 SGTKPGANLPVMLWIFGGGFEVGGSSLPFAQMITASVLMGKPIIHVSVNYRVSWSWGFLA 180
P32946 PGTRASAGLPVMLWIFGGGFELGSSSLFPDQMVAKSVLMGKPIIHVSMNYRVASWGFLA 179
*:*:*:*:*:*:*:*:*:*:*:*:*:*:*:*:*:*:*:*:*:*:*:*:*:*:*:*:*:*:*:*:*

P32947 GDDIKAFGSSNAGLNDQRLGMQWADNIAFGGDPKVTIFGESAGSMVSLCHLWINDGD 240
P32949 GPDIKAFGSSNAGLNDQRLGMQWADNIAFGGDPKVTIFGESAGSMVSLCHLWINDGD 240
P20261 GDEIKAFGSSNAGLNDQRLGMQWADNIAFGGDPKVTIFGESAGSMVSMCHLWINDGD 240
P32948 GPDIKAFGSSNAGLNDQRLGMQWADNIAFGGDPKVTIFGESAGSMVSLCHLWINDGD 240
P32946 GPDIKAFGSSNAGLNDQRLGMQWADNIAFGGDPKVTIFGESAGSMSTFVHLWINDGD 239
*:*:*:*:*:*:*:*:*:*:*:*:*:*:*:*:*:*:*:*:*:*:*:*:*:*:*:*:*:*:*:*

P32947 NTYKPKPLFRAGIMQSGAMVPSDPVDGTYGNEIYDLFVSSAGCGSASDKLACLGLSSSDT 300
P32949 NTYKPKPLFRAGIMQSGAMVPSDPVDGTYGTQIYDTLVASTGCSASNSDKLACLGLSSSDT 300
P20261 NTYKPKPLFRAGIMQSGAMVPSDAVDGIYGNEIFDLLASNAGCGSASDKLACLGLSSSDT 300
P32948 NTYNGKPLFRAAIMQSGAMVPSDPVDGTYGTEIYNQVVASAGCGSASDKLACLGLSSSDT 300
P32946 NTYNGKPLFRAAIMQSGCMVPSDPVDGTYGTEIYNQVVASAGCGSASDKLACLGLSSSDT 299
*:*:*:*:*:*:*:*:*:*:*:*:*:*:*:*:*:*:*:*:*:*:*:*:*:*:*:*:*:*:*:*

P32947 LLDATNNTPGFLAYSSLRLSYLPRPDGKNITDDMYKLVDRDGKYASVPVVIIGDQNDDEGTF 360
P32949 LLDATNNTPGFLAYSSLRLSYLPRPDGANITDDMYKLVDRDGKYASVPVVIIGDQNDDEGTF 360
P20261 LEDATNNTPGFLAYSSLRLSYLPRPDGVNITDDMYALVREGKYANIPVVIIGDQNDDEGTF 360
P32948 LLDATNNTPGALAYPSLRLSYLPRPDGTFITDDMYKLVDRDGKCANVPVVIIGDQNDDEGTF 360
P32946 LYQATSDTPGVLAYPSLRLSYLPRPDGTFITDDMYALVDRDGKYAHVPVVIIGDQNDDEGTF 359
*:*:*:*:*:*:*:*:*:*:*:*:*:*:*:*:*:*:*:*:*:*:*:*:*:*:*:*:*:*:*

P32947 GLSSLNVTTDQAQAPYFQSFIFHASDAEIDTLMAAYPQDITQGSFEDTGFIFNAITPQFKR 420
P32949 GLSSLNNTTDAQAPYFQSFIFIHATDGLMAAYPQDITQGSFEDTGFILNALTPOFKR 420
P20261 GTSSLNVTTDQAQAPYFQSFVHASDAEIDTLMTAYPGDITQGSFEDTGFILNALTPOFKR 420
P32948 GLSSLNVTTDQAQAPYFQSFIFHASDAEIDTLMAAYSDITQGSFEDTGFIFNAITPQFKR 420
P32946 GLSSLNVTTDQAQAPYFQSFIFHASDAEIDTLMAAYTSDITQGSFEDTGFIFNAITPQFKR 419
*:*:*:*:*:*:*:*:*:*:*:*:*:*:*:*:*:*:*:*:*:*:*:*:*:*:*:*:*:*

P32947 ISAVLGDLEAFIHAQYFVFLNHFQGGTKYSFSLKQLSGLPIMGTFHNDIVWQDYLLGSGSV 480
P32949 INAVLGDLEFTLSIHAQYFVFLNHFQGGTKYSFSLKQLSGLPILGTFHNDIVWQHFLGSGSV 480
P20261 ISAVLGDLEFTLAFIHAQYFVFLNHFQGGTKYSFSLKQLSGLPVLGTFHNDIVWQDYLLGSGSL 480
P32948 ISAVLGDLEFTLAFIHAQYFVFLNHFQGGTKYSFSLKQLSGLPVLGTFHNDIVWQDYLLGSGSV 480
P32946 ISAVLGDLEFTLAFIHAQYFVFLNHFQGGTKYSFSLKQLSGLPVLGTFHNDIVWQDYLLGSGSV 479
*:*:*:*:*:*:*:*:*:*:*:*:*:*:*:*:*:*:*:*:*:*:*:*:*:*:*:*:*

P32947 IYNNAFIRFATLDPNTAGLLVNWPKYTSSSQSGNNLMMINALGLYTKDNFRFAGYDAL 540
P32949 IYNNAFIRFATLDPNTAGLSVQWPKSTSSQAGDNLMINALGLYTKDNFRFAGYDAL 540
P20261 IYNNAFIRFATLDPNTAGLLVWPKYTSSSQSGNNLMMINALGLYTKDNFRFAGYDAL 540
P32948 IYNNAFIRFATLDPNKAGLLVNWPKYTSSSQSGNNLMMINALGLYTKDNFRFAGYDAL 540
P32946 IYNNAFIRFATLDPNKAGLWNWPKYTSSSQSGNNLMQINGLYTKDNFRFAGYDAL 539
*:*:*:*:*:*:*:*:*:*:*:*:*:*:*:*:*:*:*:*:*:*:*:*:*:*:*:*

P32947 WTNPSSEFFV 549
P32949 FADPSHEFFV 549
P20261 FSNPSSEFFV 549
P32948 WTNPSSEFFV 549
P32946 FSNPSSEFFV 548
*:*:*:*:*

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Figure 19-A: ClustalW alignment of protein sequences in sub-family B.I. The “green flashy” line shows the salt bridge, the blue line shows the disulfide bonds and the red box is the flap region for sequences P20261 and P32947 and the predicted flap region for sequence P32949. The secondary structures from the crystal structure for sequences of P20261 and P32947 are available and are shown in bold “flashy green” for alpha helix and bold red for Beta sheet. For P32949 the secondary structure was predicted based on the PREDATOR (Argos *et al.*, 1996) method. The α -helices and β -sheets are shown in light-green and orange respectively.



Figure 19-B: The stereo image of comparison between sequences P20261 and P32947 derived from VAST (NCBI).

A.

```

P22394      MVSKSLFLAAAVNLAGVLAQAPRPSLNGNEVISGVLEGKVDTFKGI PFADPPLNDLRFKH 60
P79066      MVSKSLFLAAAVNLAGVLAQAPTAVLNGNEVISGVLEGKVDTFKGI PFADPPLNDLRFKH 60
P17573      MVSKTFFLAAALNVVGTLAQAPTAVLNGNEVISGVLEGKVDTFKGI PFADPPVGLRFXH 60
*****:*****:*.*****.*****:*****:*****

P22394      P Q P F T G S Y Q G L K A N D F S I A C M Q L D P G N S L T L L D K A L G L A K V I P E E F R G P L Y D M A K G T V S M 120
P79066      P Q P F T G S Y Q G L K A N D F S I A C M Q L D P G N S L T L L D K A L G L A K V I P E E F R G P L Y D M A K G T V S M 120
P17573      P Q P F T G S Y Q G L K A N D F S I A C M Q L D P G N A I S L L D K V V G L G K I I P D N L R G P L Y D M A Q G S V S M 120
*****:*****:*****.*****.*****.*****:*****:*****

P22394      N E D C L Y L N V F R P A G T K P D A K L P V M V W I Y G G A F V Y G S S A A Y P G N S Y V K E S I N M Q P V V F V S 180
P79066      N E D C L Y L N V F R P A G T K P D A K L P V M V W I Y G G A F V Y G S S A A Y P G N S Y V K E S I N M Q P V V F V S 180
P17573      N E D C L Y L N V F R P A G T K P D A K L P V M V W I Y G G A F V G S S A S Y P G N G Y V K E S V E M Q P V V F V S 180
*****:*****:*****:*****.*****:*****:*****

P22394      I N Y R T G P F G L G G D A I T A E G N T N A G L H D Q R K G L E W V S D N I A N F G G D P D K V M I F G E S A G A M 240
P79066      I N Y R T G P F G L G G D A I T A E G N T N A G L H D Q R K G L E W V S D N I A N F G G D P D K V M I F G E S A G A M 240
P17573      I N Y R T G P Y G F L G G D A I T A E G N T N A G L H D Q R K G L E W V S D N I A N F G G D P D K V M I F G E S A G A M 240
*****:*****:*****:*****:*****:*****:*****

P22394      S V A H Q L I A Y G G D N T Y N G K K L F H S A I L Q S G G P L P Y H D S S S V G P D I S Y N R F A Q Y A G C D T S A S 300
P79066      S V A H Q L I A Y G G D N T Y N G K K L F H S A I L Q S G G P L P Y H D S S S V G P D I S Y N R F A Q Y A G C D T S A S 300
P17573      S V A H Q L V A Y G G F N T Y N G K Q L F H S A I L Q S G G P L P Y F D S T S V G P E S A Y S R F A Q Y A G C D A S A G 300
*****:*****:*****:*****:*****:*****:*****

P22394      A N D T L E C L R S K S S S V L H D A Q N S Y D L K D L F G L L P Q F L G F G P R P D G N I I P D A A Y E L F R S G R Y 360
P79066      A N D T L E C L R S K S S S V L H D A Q N S Y D L K D L F G L L P Q F L G F G P R P D G N I I P D A A Y E L F R S G R Y 360
P17573      D N E T L A C L R S K S S D V L H S A Q N S Y D L K D L F G L L P Q F L G F G P R P D G N I I P D A A Y E L Y R S G R Y 360
*:*:*****:***.*****:*****:*****:*****

P22394      A K V P Y I S G N Q E D E G T A F A P V A L N A T T P H V K K W L Q Y I F Y D A S E A S I D R V L S L Y P Q T L S V G 420
P79066      A K V P Y I S G N Q E D E G T A F A P V A L N A T T P H V K K W L Q Y I F Y D A S E A S I D R V L S L Y P Q T L S V G 420
P17573      A K V P Y I T G N Q E D E G T I L A P V A I N A T T P H V K K W L K Y I C S E A S D A S L D R V L S L Y P G S W S E G 420
*****:*****:*****:*****:*****:*****:*****

P22394      S P F R T G I L N A L T P Q F K R V A A I L S D M L F Q S P R R V M L S A T K D V N R W T Y L S T H L H N L V P F L G T 480
P79066      S P F R T G I L N A L T P Q F K R V A A I L S D M L F Q S P R R V M L S A T K D V N R W T Y L S T H L H N L V P F L G T 480
P17573      A P F R T G I L N A L T P Q F K R I A A I F T D L L F Q S P R R V M L N A T K D V N R W T Y L A T Q L H N L V P F L G T 480
:*****:*****:*****:*****:*****:*****:*****

P22394      F H G N E L I F Q F N V N I G P A N S Y L R Y F I S F A N H H D P N V G T N L L Q W D Q Y T D E G K E M L E I H M T D N 540
P79066      F H G N E L I F Q F N V N I G P A N S Y L R Y F I S F A N H H D P N V G T N L L Q W D Q Y T D E G K E M L E I H M T D N 540
P17573      F H G S D L L F Q Y Y V D L G P S S A Y R R Y F I S F A N H H D P N V G T N L K Q W D M Y T D S G K E M L Q I H M I G N 540
***:*.***:*.***:*.*****:*****:*****:*****

P22394      V M R T D D Y R I E G I S N F E T D V N L Y G 563
P79066      V M R T D D Y R I E G I S N F E T D V N L Y G 563
P17573      S M R T D D F R I E G I S N F E S D V T L F G 563
*****:*****:*****:*****

```

Figure 20-A. Multiple sequence alignment of the protein sequences in sub-family B.II. The two red boxes show the flap region for sequences P22394 and P17573 and the predicted flap region for sequence P79066. The blue lines show the disulfide bond and the pink line shows the salt bridge between Glu 119 and Arg-309

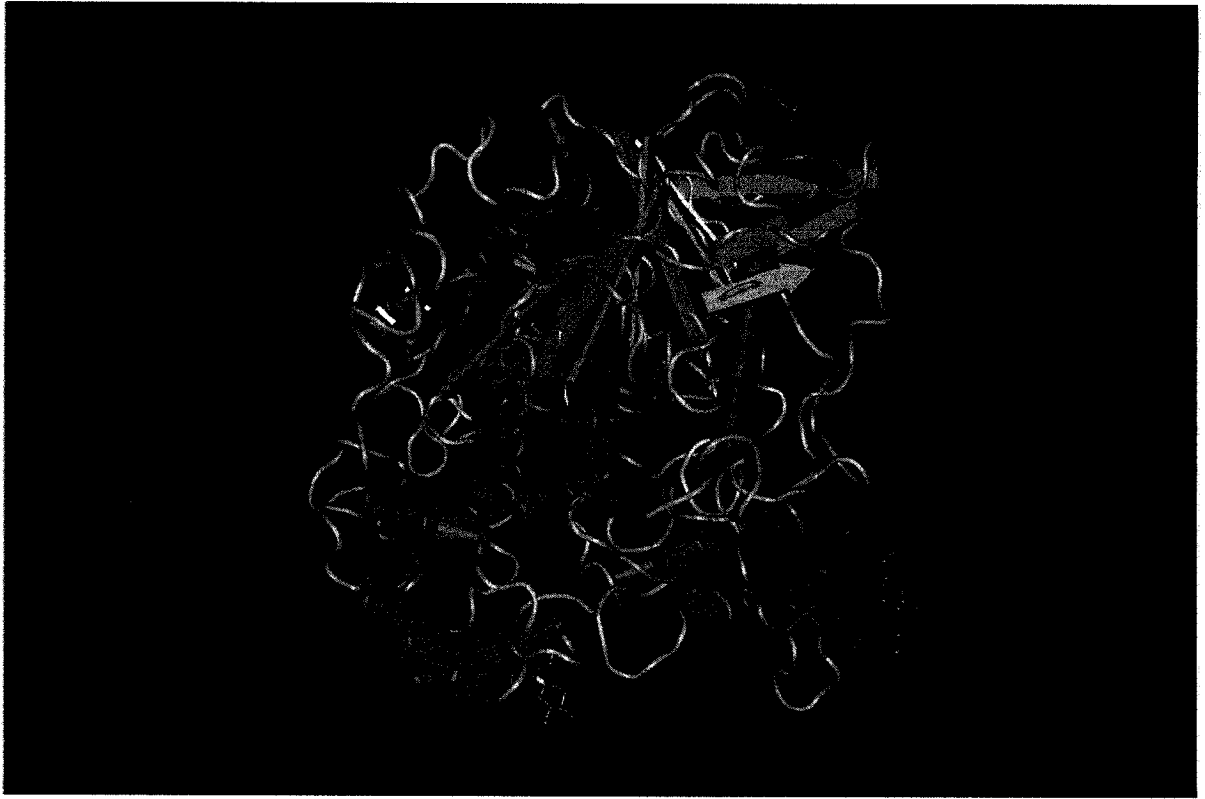


Figure 20-B: 3D image of P22394 (PDB# 1THG) derived from Cn3D 4.1 shows the α -helices and β -sheets in the tertiary structure of this sequence.

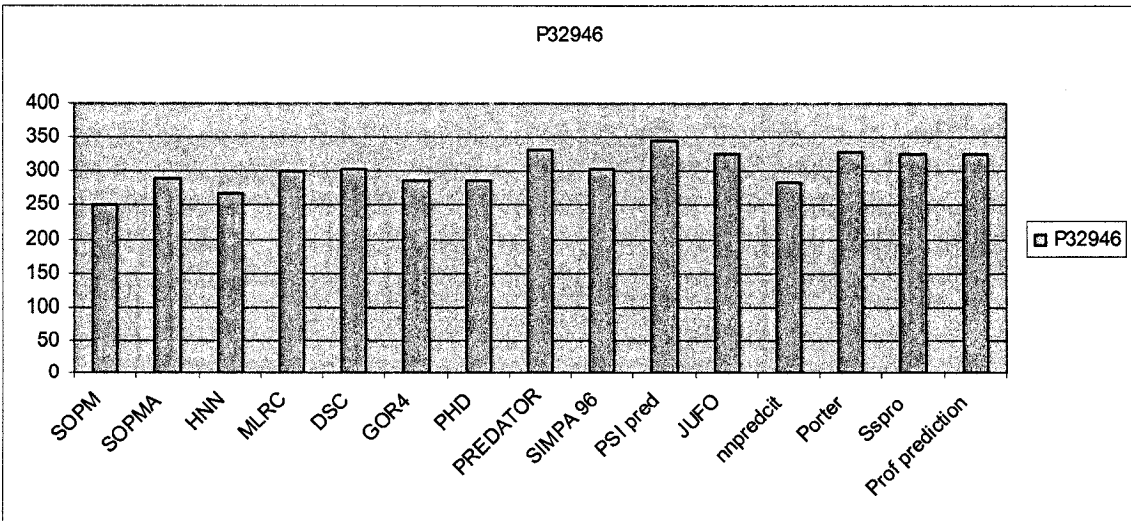
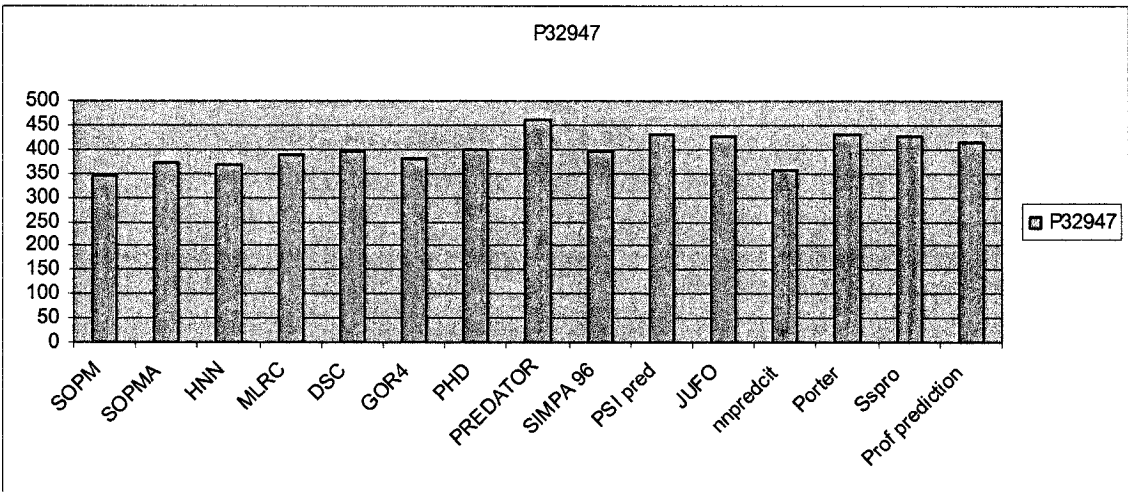
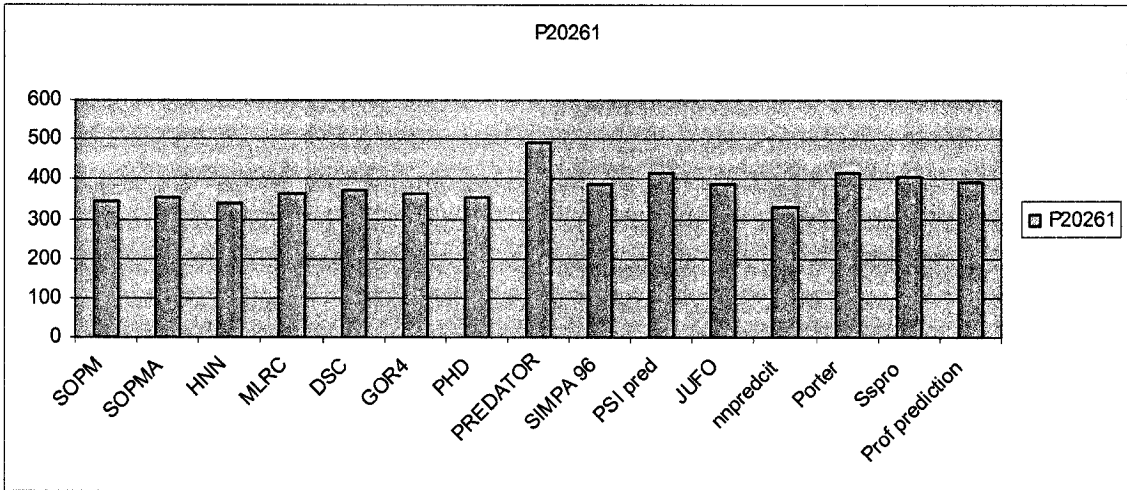


Figure 21: Comparing fifteen SS tools for sequences P20261, P32947 and P32946. Y axis indicates the protein accuracy for individual methods.

For the sub-family B.II, PORTER had the highest score and predicted the alpha helices and beta sheets closer to the structure based on X-ray diffraction studies. Thus, the sequence P79066 was analyzed with the PORTER program to predict its secondary structure (Figure 20). There is no tertiary structure available for any sequences of the sub-family B.III. Using sequence alignment, we predicted not only the secondary structures, but also some important structural features. In group B, which belongs to carboxylase-esterases, the active site includes three residues as the catalytic triad. However, for sequence Q96VC9 in sub-family B.III only two catalytic residues, serine, (at positions 200) and histidine (at positions 400) were predicted by similarity in SWISS-PROT. Moreover, according to multiple sequence alignments (Figure 7) the third residue in the catalytic triad (glutamic acid) was predicted and it perfectly matched with the other sequences in this cluster. Moreover, the catalytic residue, serine, in lipases is shielded from the solvent by one or more loops called the flap; the rearrangement of the flap opens access to the active site for the substrate (Brady *et al.*, 1990; Brozozowski and Thim 1991). The flap region and the salt bridges for sequences P32949 and P32948 in sub-family B.I, and P79066 in sub-family B.II were predicted according to sequence similarities (Figure 19-20).

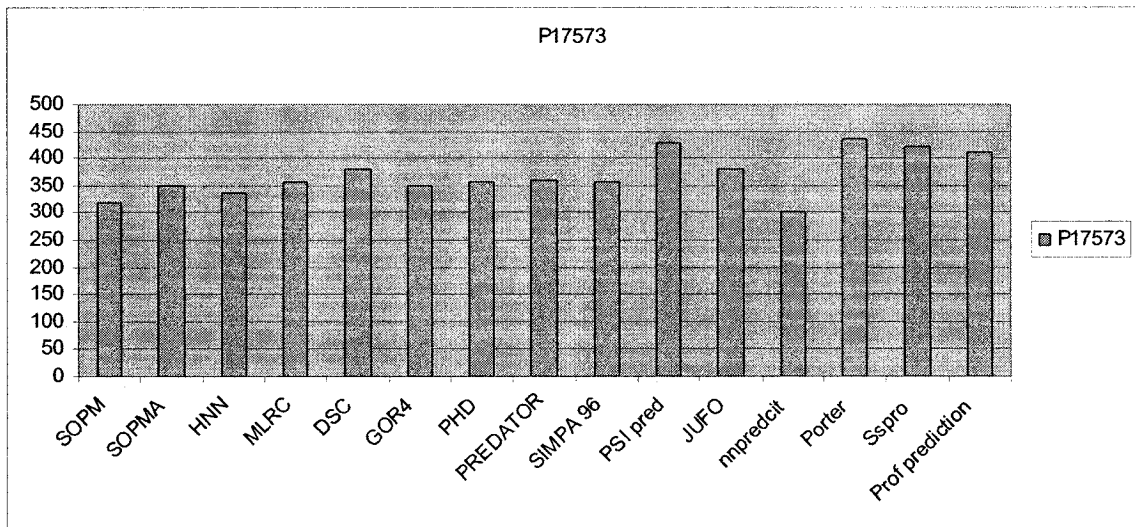
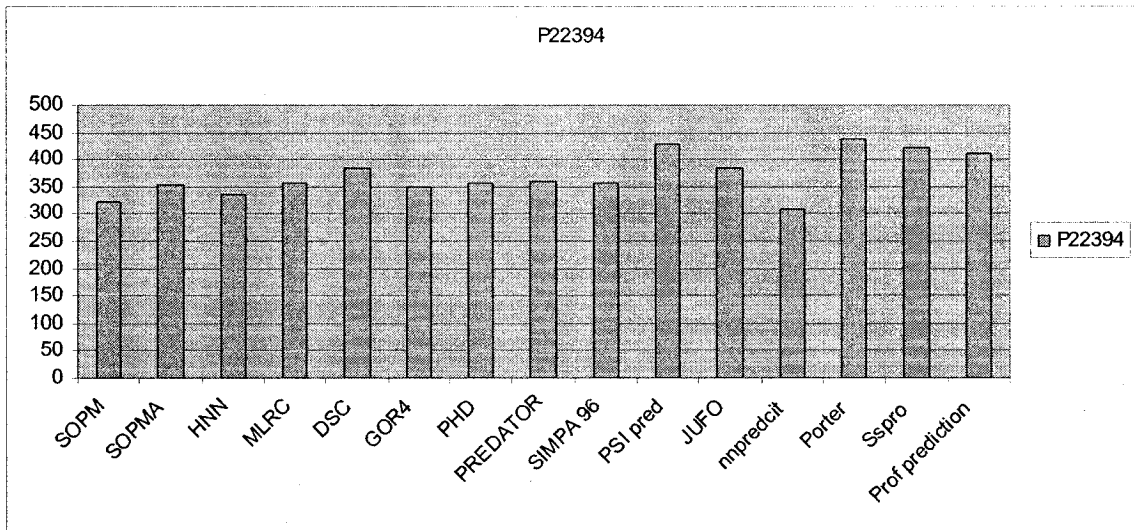


Figure 22: Comparing fifteen secondary structure prediction tools for sequences P22394 and P17573. Y axis indicates the protein accuracy for individual methods.

D.2.2 Secondary structure prediction in Subfamilies C.I and C.II:

For group C, X-ray crystallography based on structures of three sequences (P19515, P61871 and O59952) is available. These sequences were compared separately with the fifteen secondary structure prediction tools (Figure 23). The highest score for structure prediction with sequences of the sub-family C.I and C.II was obtained with SSPRO followed by a slightly lower score with PSI-PRED and PROTER. Thus, SSPRO was chosen to predict the secondary structure for sequences P61872 in sub-family C.I and P61870 and P61869 in sub-family C.II.

D.2.3 Structural feature prediction of group D

There is no known structure available for fungal phospholipase B in group D. Several lines of evidence suggested that human cytosolic phospholipase A2 (cPLA2) may be a member of the α/β hydrolase family. cPLA2 is also similar to fungal phospholipase B. To get a better understanding of the structure of fungal phospholipase B, we compared the human cytosolic phospholipase A2 (cPLA2) structure (which has a known X-ray crystal structure, 1CJY) with all of the fungal sequences in group D (Appendix 2). cPLA2 contain a dyad catalytic motif including serine (Ser-228) as the nucleophilic residue (Sharp *et al.*, 1994) and aspartic acid (Asp-549) located in a deep cleft. This serine is present in a pentapeptide sequence Gly-Leu-Ser-Gly-Ser. A different residue, Arg-200, is required for catalysis, suggesting that cPLA2 may employ a novel mechanism (Dessen *et al.*, 1999). Also the structure reveals a flexible lid that must move to allow substrate access to the active site; thus explaining the interfacial activation of this lipase. A Blast search (Altschul *et al.*, 1997) of the cPLA2 catalytic domain shows low degree of similarity with fungal phospholipase B that includes three glycines (196-

197-198) of the oxy-anion hole. Arg-200 (in sequence Q9Y7N6 was replaced by proline) and the “lipase motif”, (which contains Ser-228), that was conserved in all sequences (Appendix 2). Only at Q9UTH5 and P78854 lysophospholipase sequences was the serine in “lipase motif” replaced by aspartic acid. Eight conserved cysteine residues were predicted according to sequence similarities that probably compose the disulfide bonds (Appendix 2). The eight cysteine residues were not aligned with the eight cysteine residues found in cytosolic phospholipase A2.

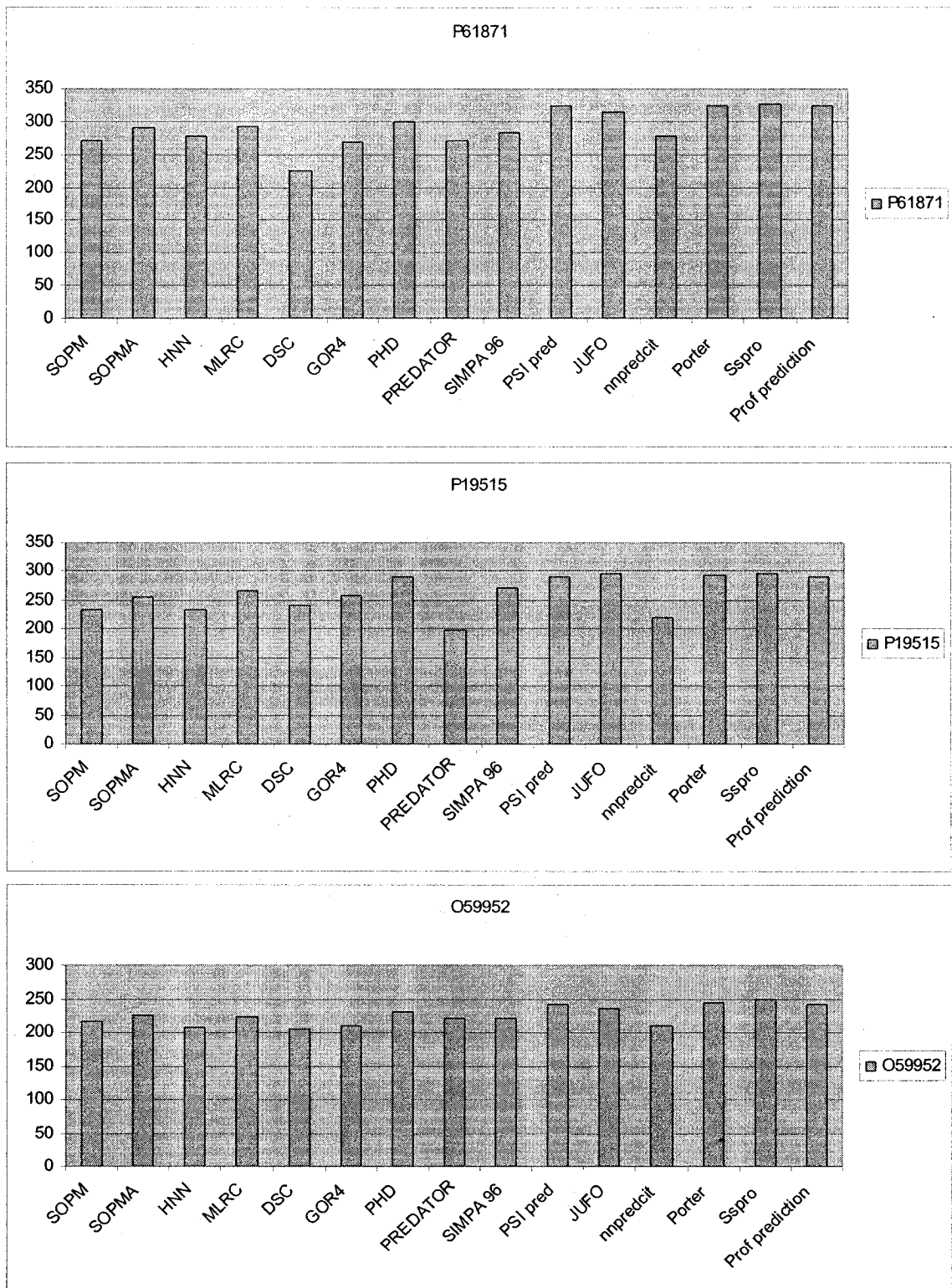


Figure 23: Comparing fifteen secondary structure prediction tools for sequences P61871, P19515 and O59952. Y axis indicates the protein accuracy for individual methods.

E. DISCUSSION

E.1 Comparative Study of Fungal Lipases

Comparative sequence analysis revealed phylogenetic relationships among the members of the lipase gene family. In this study, the 52 sequences of fungal lipases obtained from the SWISS-PROT database were categorized via sequence comparisons, crystal structure, and structure-function relationships. In agreement with the classification system derived from pfam (Protein Family Database), the present set of 52 fungal lipases can be divided into five major groups (A, B, C, D, and F), two additional groups each of which with only one member (E and G), and two sequences, P41365 and P54857, without resemblance to the rest (except in some conserved motifs) (Figure 4). This division is based on the phylogenetic tree (Figure 4) and sequence alignment shown in appendix 1.

E.1.1 Group A (Secretory lipase)

High similarity amongst sequences of group "A" (80% identical amino acid sequences) suggests that they are derived from the same ancestral gene. Amino acid sequences in group "A" contain putative *N*-glycosylation sites and also had four conserved cysteine residues may form disulfide bridges contributing to a similar three-dimensional structure. In addition, all sequences in this group, except lipase 7 (Q9P4E7) contain a putative N-terminal signal sequence. The division of group "A" into two subfamilies is based on sequence similarity. ClustalW (Thompson *et al.*, 1994) alignment shows that sequence divergence in this group happens more at the C-terminal end of the sequence and high similarities can be seen more at the N-terminus and

especially around the catalytic site (Figure 5). The fact that secreted lipases of *Candida albicans* are encoded by a family of genes may indicate that different lipase genes are needed during different stages or types of infection (Hube *et al.*, 2000). The lipase genes in group “A” constitute a large gene-family that may have evolved to adapt to the permanent association of *Candida albicans* with the human or animal host and, therefore, may also have important functions during persistence processes (Hube *et al.*, 2000).

E.1.2 Group B (Carboxylesterases)

Carboxylesterases have been divided into three categories (A, B, and C) on the basis of differential patterns of inhibition by organophosphates (Myers *et al.*, 1988; Krejci *et al.*, 1991; Cygler *et al.*, 1993). In this study group “B” (carboxylesterases) were classified into three subfamilies: BI, BII and BIII, that all belong to the *Ascomycota* taxa. Unlike the other lipases, group “B” lipases have a catalytic triad made up of the amino acids Ser-Glu-His with glutamic acid replacing the usual aspartic acid (usual in serine proteases) (Schrag *et al.*, 1991). The only exception was seen in sequence Q99156, in which the aspartic acid catalytic residue remained unchanged. The catalytic serine is shielded from the solvent by one or more loops called the “flap”. The rearrangement of the flap region allows the substrate access to the active site (Brady *et al.*, 1990; Brozowski *et al.*, 1991). The formation of Gly -X₁- Ser -X₂- Gly in the active site (Figure 7) is conserved in this group. Also, X₁ in this group is glutamic acid and X₂ is alanine. This family includes two disulfide bonds that make the structure more flexible than group “A” (Secretory lipases), which contains four disulfide bonds (Brady *et al.*, 1990).

E.1.3 Group C (AB hydrolase super family/TAG)

Group “C” is known as “true lipases”, expression first used by Arpigny and Jaeger (1999) to describe classification of bacterial lipolytic enzymes. This group (group C) is divided into two subfamilies: “C.I” and “C.II”. “C.I” consists of three sequences all of which are from the *Zygomycota* taxa. These lipases not only hydrolyze ester bonds of triacylglycerols, but also synthesize ester bonds in transesterification. Transesterification by lipases is particularly useful in industry, for example, in the production of cocoa butter substitutes (Matsuo *et al.*, 1980). The lipases produced by these fungi code for proteins of 363-392 amino acids with a total relative molecular mass of 40-42 kDa. The sequence differences between sequence P19515, and the rest of the *Rhizopus* lipases in sub-family “C.I” could be attributed to amino acid insertion of residues 68-76 and 80-85 in the signal peptide (prepeptide). The catalytic center of the “true lipase” is made up of three residues: Ser-Asp-His and it is responsible for the nucleophilic attack on the ester carbonyl carbon atom (Derewenda *et al.*, 1992). The oxyanion hole which helps to stabilize the growing negative charge on the peptide oxygen through hydrogen bonding also is present in these sequences (Derewenda *et al.*, 1992). This oxyanion hole is responsible for stabilizing the tetrahedral intermediate and the helical “lid”. This helical “lid” is responsible for the interfacial activation upon adsorption of the enzyme to an oil-water interface.

Sub-family “C.II” includes three sequences with smaller molecular masses (30-32 kDa). Also sequences in “C.II” members, P61870 and P61869, have unique substrate specificity, in that they are strictly specific to mono and diacylglycerols but not triacylglycerols. Due to the exclusive substrate specificity of these two sequences, they

are classified as E.C. 3.1.1.- (E.C.3.-.-, stands for *hydrolases*, and E.C. 3.1.-. - indicating action on *ester bonds* and E.C.3.1.1. - *Carboxylic ester hydrolases*) but have different functions from triacylglycerols or other lipases with E.C. 3.1.1.3 and (Table 3 and Figure 10).

E.1.4 Group D (Lysophospholipase catalytic domain)

Group “D” encompasses 19 sequences of phospholipase B; and is further divided into five subfamilies with more than 45% sequence similarities. According to the specificity of ester linkage that will be cleaved by phospholipases, these sequences will be classified into different groupings including, A1, A2, B, C, and D (Ansell and Hawthorne, 1964; Wang and Dennis, 1999, Ghannoum, 2000). Among this group of multi-functional phospholipases, phospholipase B (PLB) enzymes are perhaps the most poorly understood-- particularly in their biological functions (Kuwabara and Shimooka 1989; Gassama-Diagne *et al.*, 1989; Satio *et al.*, 1991; Witt *et al.*, 1989). Fungi possess a class of highly homologous PLB enzymes that exhibit little sequence similarity to PLB proteins identified thus far in other eukaryotes. The physiological functions of fungal PLB enzymes are largely unknown. The phospholipase B refers to an enzyme that can remove either *Sn-1* or *Sn-2* fatty acids from a glycerophospholipid. However, fungi can have a single enzyme that has not only the hydrolase (fatty acid release) activities of phospholipase B, but also lysophospholipase (LPL) and transacylase activities (Cox *et al.*, 2001). The finding of a single enzyme having these multiple and seemingly paradoxical functions was seen before in *Candida albicans*, *Penicillium notatum* and *Saccharomyces cerevisiae* (Saito *et al.*, 1991; Lee *et al.*, 1994; Leidich *et al.*, 1998). Phospholipase B and phospholipase A2 belongs to a family called “lysophospholipase

catalytic domain” which has a C2 domain. The C2 domain is a Ca^{2+} -dependent membrane-targeting module found in many cellular proteins involved in signal transduction or membrane trafficking. The only known 3-D structure for “lysophospholipase catalytic domain” family member is “Human cytosolic phospholipase A2” (cPLA2) (accession code on PDB: 1cgy). In this study all phospholipase B sequences were compared to “Human cytosolic phospholipase A2” sequence to find conserved motifs. All phospholipase B sequences were aligned with “Human cytosolic phospholipase A2” and showed not only a conserved serine corresponding to Ser 228 of cPLA2, but also the consensus sequence Gly-(Leu)-Ser-(Gly)-Ser motif. Moreover, the second probable active site residue (Aspartic acid) was conserved in all sequences. Searching for the histidine as the third catalytic residue showed that none of the 19 histidine residues was enzymatically relevant in the lysophospholipase catalytic domain family (Pickard *et al.*, 1996). Also, Arg200, a different residue, is required for catalysis, suggesting that cPLA2 may employ a novel mechanism (Dessen, 2000). An interfacial activation phenomenon has been shown in both cPLA2s and lysophospholipases that preferentially cleave substrates presented at a membrane interface rather than in monomeric form (Nalefski *et al.*, 1994).

E.1.5 Group E (ab-hydrolase)

Triglyceride lipase-cholesterol esterase (P34163) is the only sequence belonging to the “AB hydrolase superfamily” and branched off from the rest of the true lipases and phospholipases (Figure 4).

E.1.6 Group F (Phospholipase C)

Group F, which includes three sequences from phosphoinositide-specific

phospholipase C enzymes (PI-PLCs) that play a critical role in receptor-linked signaling at the plasma membrane of eukaryotic cell (Singer *et al.*, 1997). In yeast, the PLC1 gene has been shown to be important for growth, but the functional reason for this behavior remains unclear (Singer *et al.*, 1997).

Hydrolysis of inositol phospholipids, particularly phosphatidyl-inositol-4, 5-bisphosphate (PIP₂) by PI-PLCs, results in the formation of secondary messenger molecules, *sn*-1, 2-diacylglycerol (DAG) and inositol 1, 4, 5-triphosphate (IP₃). These are important in regulating many cellular functions (Berridge, 1993). IP₃ causes the release of Ca²⁺ ions from internal reserves and DAG activates protein kinase C (PKC) (Singer *et al.*, 1997). Calcium mobilization and PKC activation are necessary for many cellular activities, including secretion, cell growth and proliferation (Nishizuka, 1992; Mitchell, 1992). PI-PLCs have been categorized into three subfamilies, β , γ and δ with two to four isoforms in each. Each sub-family has two conserved motifs ('X-box' and 'Y-box' important for enzyme activity) that show low similarity among subfamilies. However, they can be distinguished by their primary amino acid structure and molecular size (Rhee *et al.*, 1989). The order of these two regions is always the same (NH₂-X-Y-COOH), but the spacing is variable. There is usually a distance of 50-100 residues between these two regions and the distance in this case is about 70 residues (Singer *et al.*, 1997). The two conserved regions have been shown to be important for the catalytic activity (Singer *et al.*, 1997). At the C-terminus of the Y-box, there is a C2 domain possibly involved in Ca-dependent membrane attachment that was described in the phospholipase B family. By profile analysis, it shows that sequences with significant similarity to the X-box domain occur also in prokaryotic and trypanosome PI-specific

phospholipases C. Apart from this region, the prokaryotic enzymes show no similarity to their eukaryotic counterparts. The smallest PI-PLCs are δ isoenzymes, as they contain only sequences common to all PI-PLCs subfamilies and are frequent to all eukaryotes, from yeast and molds to plants and mammals (Singer *et al.*, 1997).

E.1.7 Group G (Phospholipase D)

P36126 from *Saccharomyces cerevisiae* in group “G” belongs to phospholipase D (PLD) and has the largest molecular mass as compared to the other lipase sequences studied here. PLD, in general, hydrolyzes phospholipids at the terminal phosphorus ester bond, leading to formation of phosphatidic acid (PA) and the free hydrophilic alcohol substituent (Figure 12). PLD1 known as phosphatidylcholine (PC), has been observed in response to a variety of agents including hormones, neurotransmitters, growth factors and phorbol esters. PA has been shown to stimulate DNA synthesis, cell proliferation, and phosphatidylinositol 4, 5-bisphosphate-phospholipase C and phosphatidylinositol phosphate kinase (Wang *et al.*, 1994).

E.2 Evaluation of Secondary Structure Prediction Tools for fungal Lipases

E.2.1 Multiple Sequence Alignment and Secondary Structure prediction for families B and C

Protein secondary structure (SS) prediction is an important stage in the prediction of protein structure and function. Accurate SS information improves the sensitivity of threading methods (Jones, 1999) and is at the core of most *ab initio* methods (Bradley *et al.*, 2003 b) for the prediction of protein structure. Multiple alignment provides much more structural information than a single sequence could (Russell and Sternberg, 1995).

Undoubtedly, the accurate alignment of various sequences is crucial for predicting secondary structures accurately. Recent studies have accurately predicted secondary structures from multiple alignments (Musacchio *et al.*, 1994 and Jenny and Benner, 1994 a). We carried out multiple sequence alignments for each sub-family separately. The results were compared with the best secondary structure prediction tools (section D.2.1). Musacchio *et al.* (1994) and Jenny and Benner (1994 b), used multiple sequence alignment to predict the secondary structure of pleckstrine homology (PH) domain protein. A year later, when the tertiary structure of this protein was determined, Russell and Sternberg (1995) used different secondary structure tools to test the accuracy of these predictions for the PH domain. They showed that multiple sequence alignment prediction was remarkably accurate and also that the PHD method (Rost B., 1996) (a secondary structure prediction tool) had the highest accuracy for this specific protein (Russell and Sternberg, 1995).

In this study we used fifteen different SS prediction tools from which we chose the best tool for structure predictions of unknown lipase sequences. To make this choice we applied our fifteen tools to known homologous sequences and decided which tools provided more accurate results. Comparing all the tools with the lipase sequences shows that the classical methods such as SOMP (Geourjon and Delage, 1994), SOMPA (Geourjon and Delage, 1995), HNN (Guermeur, 1997), MLRC (Guermeur *et al.*, 1998), DSC (King and Stenberg, 1996) and GORIV (Garnier *et al.*, 1996) do not perform well in predicting the secondary structure of these sequences. None of these methods incorporates multiple sequence alignment information. They cannot benefit from any associated improvement in accuracy. The PHD method (Rost B., 1996), on the other

hand, uses neural networks and multiple sequence alignment information, and gives improved results over the classical methods. “PROF. Prediction” (Rost, 2001) showed high accuracy for lipases via an extension of concepts implemented in PHD (Rost, 1996) in that it uses a larger data set as well as a third layer of network. The PREDATOR method (Argos *et al.*, 1996), which relies on only a single protein sequence showed high accuracy for sub-family B.I, but low predictability for other subfamilies (Figure 23). The unique feature of this approach involves database-derived statistics on residue type occurrences in different classes of beta-bridges to delineate interacting beta-strands. PSIPRED (Jones, 1999), PORTER (Pollastri *et al.*, 2002) and SSPRO (Baldi *et al.*, 2003) methods, predict protein secondary structure using the position specific scoring matrices generated by PSI-BLAST (Altschul *et al.*, 1997) and two layers of Bidirectional Recurrent Neural Networks (BRNN). This shows high accuracy for lipase sequences in group C (Baldi *et al.*, 1999). For the sub-family B.II, PORTER had the highest score and predicted the alpha helices and beta sheets closer to the structure based on X-ray diffraction studies. Thus, the sequence P79066 was analyzed with the PORTER program to predict its secondary structure (Figure 20).

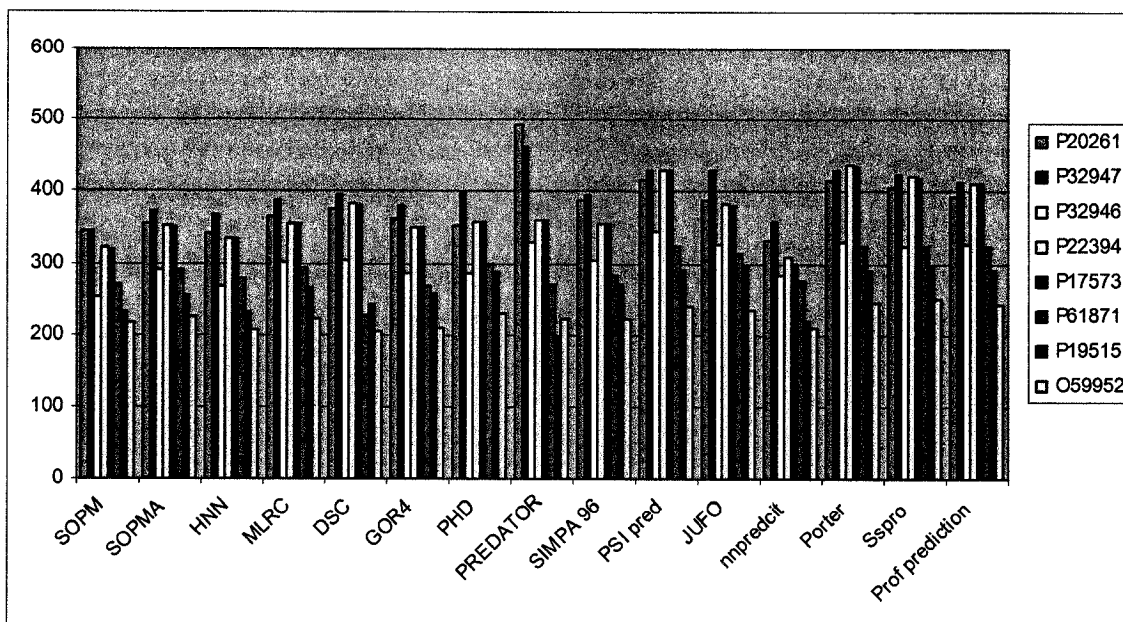


Figure 24: Comparison of fifteen secondary structure prediction tools shows the most accurate prediction for each protein.

The only noticeable errors in prediction of the B.I and B.II subfamilies (Figure 16, 17 and 19) are at the middle of the secondary structural elements, with a few motifs falsely predicted as alpha helices and beta sheets. For sub-family C.I and C.II (Figure 9 and 10), the errors are mostly at the ends of the secondary structure elements, where small errors are to be expected given the variation in accuracy of different secondary structures. Predicting the correct core secondary structure elements in the correct places is the primary point in predicting the three-dimensional fold of proteins (Russell and Sternberg, 1995).

E.3 Structural Feature Prediction

Groups B and C, comprising eight sequences with known 3-dimensional structures, were used as targets to predict the secondary and tertiary structures for unknown sequences. The secondary structures were predicted using variable structural tools as described in the previous section. Structural elements such as cysteine residues,

catalytic motifs and flap regions were predicted according to multiple sequence alignment and compared with sequences with known structural motifs. The flap region, an important feature of the majority of lipases (Schrag *et al.*, 1991), shields the catalytic site from the external environment by loops or helices (variously referred to as “loops”, “flaps”, or “lids”), which lie over the catalytic triad. The lids are displaced to different extents during the process of the interfacial activation, allowing the lipid substrate to enter the active site. Flap regions, described as responsible for interfacial activation in lipases were predicted for P32949, P32948, P79066, P61872, P61870 and P61869 sequences. In sub-family BI, three sequences (including P20261, P32947 and P32946) have known 3-D structure and two (P32949 and p32948) have unknown structure. Within the three known structures, the following motifs are strictly conserved:

- (i) Residues of catalytic triad (Ser-His-Asp)
- (ii) Residues forming salt bridges (Arg-37-Glu95 and Glu172-Arg279)
- (iii) Cysteine residues involved in formation of disulfide bonds (Cys60-Cys97 and Cys268-Cys277)
- (iv) Residues that form the oxy-anion hole upon interaction with the substrate (Gly124-Ala210)

High sequence similarity and high similarity in secondary structure may indicate that the flap region for sequences P32949 and P32948 is between residues 77-108 (Figure 19). Considering this α helix (77-108) region as the flap region for these two sequences, this motif should cover the serine active site. It is probably stabilized by a disulfide bond and a salt bridge similar to that seen for the other three sequences (P20261, P32947 and

P32946) with known structures. The lid structure shown in sequences with known X-ray crystal structures indicates one flap region covering the active site. However, comparisons with sequences in group B.II show that instead of one flap region covering the active site, there are two flap regions coming from two different surface loops, covering the active site in residues 85-94 and 313-326 (Figure 25). Having two flap regions coming from two different surfaces to cover the active site indicates that the active site serine residue is probably buried even deeper in this protein than equivalent serines in the other lipases. This is possibly the reason for the enzyme preference for long chain fatty acids (Schrag *et al.*, 1991).

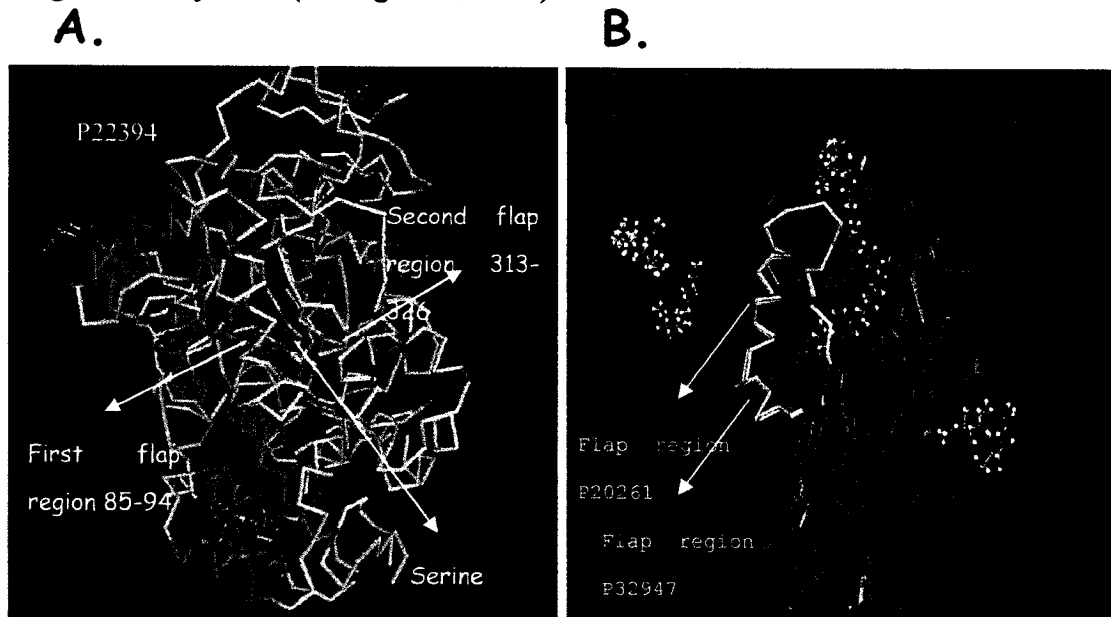


Figure 25: Comparison of Lid structure B. **A.** Two flap regions coming from two different surfaces in sequence P22394 sub-family BIII. **B.** Comparison of two similar flap regions in sequences: P20261 and P32947 (sub-family BI) using VAST derived from NCBI database.

Early reports regarding the substrate specificity of *Geotrichum candidum* lipases (GCLs) point to their preference for long chain fatty acids and for those with cis-9

unsaturated bonds (Alford and Pierce, 1961; Jensen *et al.*, 1965). Also, a study (Bertolini *et al.*, 1994) for comparison against a series of triacylglycerol substrates (including: Butyrin C4, Caproin C6, Caprylin C8, Caprin C10, Laurin C12, Myristin C14, Palmitin C16, Stearin C18:0, Triolein C18:1, Linolein C18:2) showed that these enzymes display a higher affinity for Triolein (C18:1) (Figure 26) that has a longer chain.

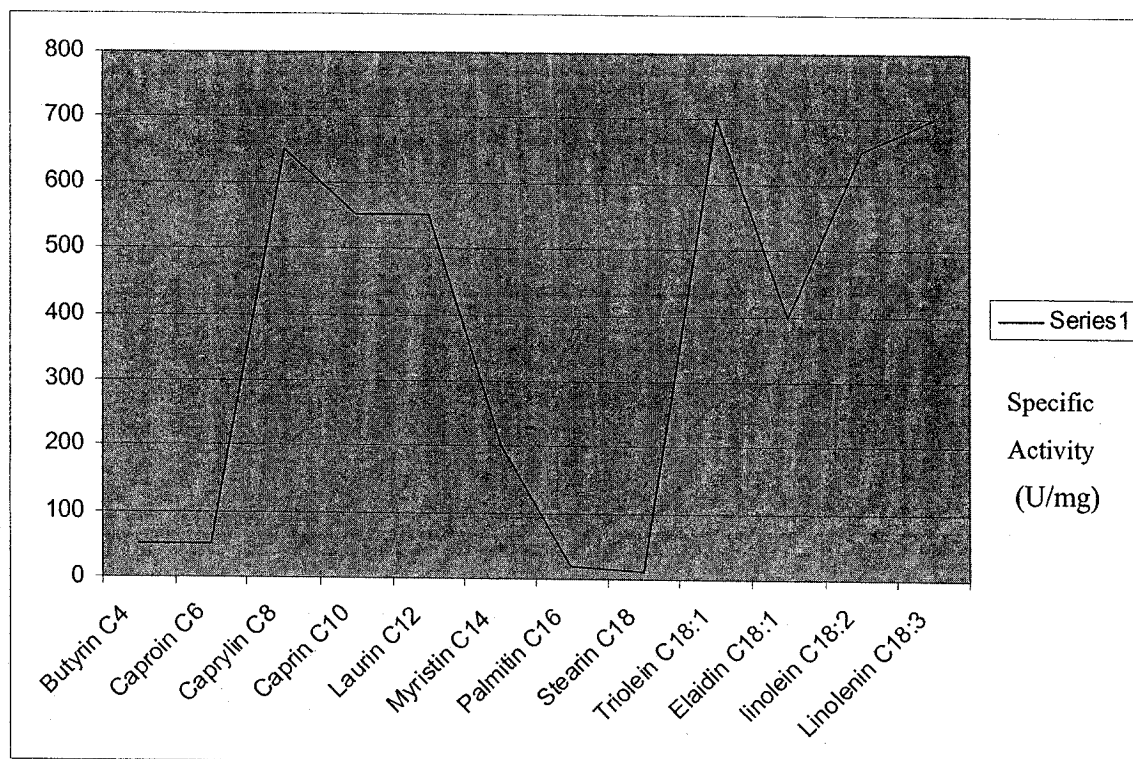


Figure 26: Comparison of different TAG substrates on *Geotrichum candidum* lipases (Adopted from Bertolini *et al.*, 1995)

Moreover, Bertolini (1995) identified the regions of the molecule that were involved in substrate differentiation. The flap region is not involved in discriminating between substrates. It is responsible for interfacial activation in lipases (Brzozowski *et al.*, 1991; van Tilbeurgh *et al.*, 1993; Grochulski *et al.*, 1993).

The flap region of sequence P19515 (*Rhizomucor miehei*) (RmL) was

experimentally proven when a complex of RmL with *n*-hexylphosphonate ethyl ester inhibitor was characterized by X-ray diffraction at 3.0 Å resolution (Brzozowski *et al.*, 1991). It includes only seven residues 179-185 (Figure 9). The same region was predicted for sequence P61871 from the experimentally crystal structure of *Rhizopus niveus* at 2.2 Å (Kohno *et al.*, 1996).

In sub-family CII, the lid structure in O59952 contains two hinge regions: N-terminal (residues 83-84) and C-terminal (residues 91-95). This sequence was aligned with sequences P61870 and P61869 and showed some similarity in their flap regions (Figure 10-A and 10-B). Furthermore, the cysteine residue in all three sequences was conserved and forms the disulfides bonds.

E.4 Structure of CALB (P41365)

As it is shown in Cn3-D view (Uppenberg *et al.*, 1994), the catalytic serine is located in the tight turn between β 4 and the following α 4 with similar conformation with other lipases. Uppenberg *et al.*, 1995 indicate that the crystal structures of four forms of CALB lack lid/lids. Although the overall structure and serine triad conformation are similar in all lipases, their substrates specificities and degrees of stereoselectivity differ widely (Kazlauskas *et al.*, 1991; Santaniello *et al.*, 1992).

This study agrees with these results by noting that P41365 is the smallest lipase among fungi. It is possible that they lack in some motifs, it is also shown by the UPGMA tree (Figure 4) that this specific triglyceride does not cluster with the rest of the other lipase, but rather, shows a close clustering with lipases that lack such motifs.

F. CONCLUSION

Through comparative and evolutionary analysis of protein sequences of biochemically characterized fungal lipases, I developed a comprehensive classification system for lipases of fungal origin. Using fungal lipases with known secondary and tertiary structure, and a variety of structure prediction software tools, I predicted the putative active sites, disulfide bonds and salt bridges of several lipases. The developed framework phylogenetic tree will serve as an important tool for predicting putative function and reaction conditions of newly identified lipase gene sequences. The results of this study will be invaluable for improving the properties of industrially important lipases through protein engineering.

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HLRLERYLRL LNIALCLRPH ANRLFEFYEL SPLGNLLSRE SGFQ GKQGYL
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-----MKTLLIFLAFLSSI FAS-----
-----MKGLVFLGLLPTI YAS-----
-----MRDLILFLSLLHTI FAS-----
-----MRGIAVFLAFISLI FAS-----
-----MKTLVVLCTLLSII FAS-----
-----MLYLILFLIAPI YAG-----
-----MLFLLFLLITPI YAG-----
-----MLFLLFLLVAPI YAG-----
-----MLYLILFLIAPI YAG-----
-----MFVFLALITLT TCL-----
TATLANGDTI TGLNAINEA FLG-----
TAKLANGDTI TGLNAINEA FLG-----
TATLANGDTI TGLNAINEA FLG-----
TATLANGDTI TGLNAIVNEK FLG-----
TATLANGDTI TGLNAINEA FLG-----
RPSLNGNEVI SGVLEGVDT FKG-----
TAVLNGNEVI SGVLEGVDT FKG-----
TAVLNGNEVI SGVLEGVDT FKG-----
-----MVSFISIS-----
-----MVSFISIS-----
-----MVLKQRAN-----
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GYAPGPVSCP --SSQLIRSG SQG-----
LYTPGYVQCP --EGKLTRSS LDG-----
DYAPFNLTCP --SKKTFIRT ASE-----
SYTPANVSCD -EDINLIRQA S-G-----
SYVPANVTCD -DDINLVREA S-G-----
SYVPANVTCP -NDINLLRNA T-G-----
SYVPGTVSCP -DDINLVREA T-S-----
GYAPGVVDCD -ENINLVRKA D-A-----
-YTPQNVSCP -DNANFIRNA ADG-----
GYAPSIIPCP SDDTSLVRNA S-G-----
-YVPTSVSCP --ASRPTVRS AAK-----
-YAPAVVDCP --KTKPTLRK AVD-----
SYAPYVNTCP S-DYMLRPAS DG-----
SYAPFNVTCS N-DNLLRPAS EG-----
SYAPYTVACP S-GSLLRPAS DG-----
SYAPVIRSCD SSEIMVNSLP RGE-----
SYAPYYVDCP S-DNIVESLS SNE-----
SYAPWQVDCP -SNVTWIRNA TTG-----
SYAPWQVDCP -SNVTWIRNA TTG-----
TIVPESIQMG CSLLRITKVK VRQRKVS LDP
--LCRVLQIR MIRMTRRR RKFYEFKLN
KIPSIFTNDG MPLLKISHKS KKRILEWIDP SCFKFSWRMA NSTTTTTSAT
-----
---MSVTSTS LNG-----
----MPLELP SLNASIVGNT VQN-----
VIRSTAKAQG WRVSHFGKHA FKDMIDRHTT KWFLVRN----SYLTY
MYFFFLGRLS ITDYIIIVLV YIES-----
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-----LIG LTPPSKDSFY
-----LVH ITPASEDDFY
-----LFS LKPPSQDDFY
-----PLT VKSPLVDDFY
-----PLS LKSPLVDDFY
-----LIF PTKPSSDPFY
-----LIF PTKPSSDPFY
-----LIL PTKPSNDPFY
-----VLL PTKPSIDPFY
-----QIP LN-PTIDDFY
-----IPF AEPPVGNLRF
-----IPF AEPPVGNLRF
-----IPF AEPPVGNLRF
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-----IPF AEPPVGT LRF
-----IPF AQPPVGNLRF
-----IPF ADPPLNDLRF
-----IPF ADPPLNDLRF
-----IPF ADPPVGD LRF
-----QGVSLCLLV
-----QGVSLCLLV
-----YLGFLIVFF
-----MRLSFFTAL
-----MRLSFFTAL
-----MRSSLV LFF
-----INPNEQSYI
-----INSNEKAYI
-----LSQEKDYI
-----PSDNETEWL
-----LSDNETEWL
-----LSQSEIDWL
-----ISQNESAWL
-----VSDDEADWL
-----LSPA EKEWL
-----LSTAETDWL
-----LSTNETSWL
-----LSNEEKNWL
-----ISSGEQ SFI
-----LNEGEQSYI
-----LSTGEQEFV
-----LPDLENDFI
-----IPSAESEYL
-----LGSGERAYI
-----LGTGERAYI
-----I SGYLMLDKNT GKAYKLCVD
-----N NGQIIWRDGS ----KYLELD
TSATTSGLPQ GITNTTALS N SAIISTPAIA TSAIHRLSIT NRTTHEFVLD
-----TFNGISEDGI
-----GA VEQFLNIRYA
VSDLSSTTPL DVFLIDWKF K VRFSGNKNNI LDNENEINWI IHDPNLEIND
-----I ISSVLKLI
-----MK LLSLTGVAGV

SPPVGFATAK PGDILKIRN--TPSAP----SSLYLPI
NPPAGFESAK NGDILKLRN--SPNRL----ASFYFPI
KAPAGFKA AK PGDILKTRK--SENKP----SSLYAPV
TAPDGYESAK LGEILKLRK--TPSKL----SSMFFEI
NPPRGYESAK LGEILKLRK--TPGKI----SSLFIPV
NPPKGFENAA VGDILQ SRA--TPKSI----TGGFTPL
NPPKGFEEAA VGDILQ SRE--TPKSI----TGRFAPL
NAPAGFEKAA VGDILQ S RK--TPKPI----TG VFPV
NAPEGFKNAT VGDILQ FRK--TPKSI----TGGFVPL
NPPKDLETSQ LGDVLKWRK--MPFPV----TSMFVNL
KDPVPYSGSL DGQKFTSYGP SCMQQNP----EGTYEEN
KDPVPYSGSL NGQKFTSYGP SCMQQNP----EGTYEEN
KDPVPYRGS L NGQSFTAYGP SCMQQNP----EGTYEEN
KPPVPYSAS L NGQQFTSYGP SCMQMNP----MG SFEDT
KPPVPYSAS L NGQKFTSYGP SCMQMNP----LGNWDSS
KHPQPFTGSY QGLKANDFSP ACMQLDP----GNSLTLL
KHPQPFTGSY QGLKANDFSP ACMQLDP----GNSLTLL
KHPQPFTGSY QGLKANDFSS ACMQLDP----GNAISLL
SSMLGSSAV P VSGKSGSSN TAVSASD----N AALPPLISSR
SSMLGSSAV P VSGKSGSSN TAVSASD----N AALPPLISSR
TAFLV--EAV PIKRQS--N STVDS-----LPPLIPSR
SAVASLGYAL PGKLQS-----
SAVASLGYAL PGKLQS-----
---VSAWTAL ASPIR-----
NARYPIAKQA LSKFLHN--A NLQNF D----VDSFLAH--SNPTIGLA
DRRYANAKSE LSRFLHN--A KMVDFD----VDGFLN--SNPTIGLA
HKRQETT NKN LIDFLSKR--A NLSDFD----AKSFINDNA PNHNITIGLS
KKRDVYTREA LRSFLDRATS NFSDSS----LVSQLF--SN ASDIPRIAVA
KKRDAYTKEA LHSFLNRATS NFSDTS----LLSTLFG--SN SSMMPKIAVA
KKRDVNTREA LESFLKRVTS NFTSNSSASN LIDQLFS--TN SSNIPKIGIA
EKRNKVTSVA LKDFLTRATA NFSDSS--E VLSKLFNDGN SENLPKIAVA

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|------------|--------------|-------------|------------|------------|
| KVRHESTVPA | LKDFLQRGFK | GFTNDS--- | IIDKLLA--T | QDTAPKVAIA |
| KKRDPIRDA | LQFLRRFA | NVS-TE---- | ITSALFN--D | TENVPKLGIA |
| KKRDAYTKEA | LHSFLSRATS | NFSDS----- | LLSTLFSS-N | SSNVPKIGIA |
| EVRRGKTLA | LKDFFGHVKV | GDYDVGA--- | YLDKHSN--N | SSSLPNIGIA |
| SIRRKNTIQP | MRDLLKRANI | TGFDSET--- | FMNEAAN--N | ISQLPNVAIA |
| DKRIPKINTQ | MRSFIS--NT | GLDVVDN--- | -----SVIN | DSGDPRLGLA |
| NKRISKVNSE | LRSFIS--KT | GLNVLDL--- | -----KVVN | SSGDPRLGLA |
| DKRVSKVNSA | LESFIS--KT | GLKIDTK--- | -----SVLN | DTDGPRGLIA |
| EKRLSNANEA | LTTFLQSKNT | TADLDLS--- | -----SIVG | D-NGPRLGIA |
| STRSTITNTA | MKDPLR--NA | NLPGLNA--- | -----DTLS | GSEGPSIGIA |
| EAREKLVQPV | IEQMAARGL | ETPPRT---- | ----- | ----PNIGVA |
| EAREKLVQPA | IEQMAARGL | ETPPRT---- | ----- | ----PVIGVA |
| DIKEIRQGRD | ARNYREQYKI | S--SENEKRW | FTIIYCAD-- | NKLKAMHMIS |
| SVKDIRIGDT | ASTYQEEVDP | KRLRSDSKLW | IAIIYKVS-- | NKLKALHVVA |
| DIKSIYIQNE | GSYREELNI | S--QKLEKNW | ITIIYFNHKK | NSLKSLHLIT |
| EIFKGIKYAN | IPYRWAYAER | IDDYDNG--- | ----- | -----VFDCT |
| DIPGKFEKPV | LKNWNGAEI | DATKVGPF--- | ----- | -----VCPQP |
| ELEEFGIEND | ANNILDKNGK | SKTHQKKSNI | SSKLLLLTLE | NSERKLIIC |
| PQPMINLFEW | LINFSTSSDD | NTIEEKL--- | -----S | APTTHEMCAI |
| MKNDNKANDI | IIDSVKVPDS | YKPKPNP--- | ----- | -----IVFC |
| LATCVAATPL | VKRLPSGSDP | AFSQPKS--- | ----- | -----VL |
| | | | | |
| VVKNWQLLI | RS----- | ---EDSFGNP | ----- | ----- |
| DVKNWQLLV | KS----- | ---EDSFGNP | ----- | ----- |
| DVQNSWQLLV | RS----- | ---EDSFGNP | ----- | ----- |
| DIKNSWQLLV | RS----- | ---EDSFGNA | ----- | ----- |
| EVKNSWQLLV | RS----- | ---EDSFGNA | ----- | ----- |
| KIQNSWQLLV | RS----- | ---EDSFGNP | ----- | ----- |
| KIQNSWQLLV | RS----- | ---EDSFGNP | ----- | ----- |
| KIQNSWQLLV | RS----- | ---EDSFGNP | ----- | ----- |
| NVQNSWQFLV | RS----- | ---EDSFGNP | ----- | ----- |
| PISNAWQISV | RS----- | ---QDTLNS | ----- | ----- |
| LPKAALDLVM | QS-----K | VFEAVSPSSE | ----- | ----- |
| LGKTALDLVM | QS-----K | VFQAVLPQSE | ----- | ----- |
| LPKVALDLVM | QS-----K | VFQAVLPNSE | ----- | ----- |
| LPKNARHLVL | QS-----K | IFQVVLPNDE | ----- | ----- |
| LPKAAINSLM | QS-----K | LFQAVLPNGE | ----- | ----- |
| DKALGLAKVI | PEEFRGPLYD | MAKGTVSMNE | ----- | ----- |
| DKALGLAKVI | PEEFRGPLYD | MAKGTVSMNE | ----- | ----- |
| DKVVGLGKII | PDNLRGPLYD | MAQGSVSMNE | ----- | ----- |
| CAPPSNKGSK | SDLQAEPYNM | QKNTEWYESH | ----- | ----- |
| CAPPSNKGSK | SDLQAEPYNM | QKNTEWYESH | ----- | ----- |
| TSAPSSSPST | TDPEAP--AM | SRN----- | ----- | ----- |
| ----- | ----- | ----- | ----- | ----- |
| ----- | ----- | ----- | ----- | ----- |
| ----- | ----- | ----- | ----- | ----- |
| FSGGGYRAML | TGAGEISSLD | SRT-KTNTPV | ----- | ----- |
| FSGGGYRAML | AGAGELLALD | SR--ATNPSV | ----- | ----- |
| FSGGGYRAML | AGAGQILGLD | GRYEDANKHG | ----- | ----- |
| CSGGGYRAML | SGAGMLAAMD | NRTDGANEHG | ----- | ----- |
| CSGGGYRAML | SGAGMLAAMD | NRTDGANEHG | ----- | ----- |
| ASGGGYRAML | SGAGMVSAMD | NRTDGANEHG | ----- | ----- |
| VSGGGYRSML | TGAGVLAAMD | NRTEGAYEHG | ----- | ----- |
| CSGGGYRAML | SGAGMISAMD | NRTDGANEHG | ----- | ----- |
| VAGGGYRAMF | VGAGAFAMAAMD | NRTDGANEHG | ----- | ----- |
| CSGGGYRAML | GGAGMIAAMD | NRTDGANEHG | ----- | ----- |
| VSGGGWRALM | NGAGAVKAFD | SRTDNATATG | H----- | ----- |
| ISGGGYRALM | NGAGFVAAAD | NRIQNTTGAG | G----- | ----- |
| FSGGGLRAMV | HGGGVLNAFD | SRNGNGSSLA | ----- | ----- |
| FSGGGLRAMV | NGGGAFNAFD | SRESDSPLS | ----- | ----- |
| ISGGGFPAML | TGAGAINAFD | ARNGNTTSLG | ----- | ----- |
| VSGGGWRSML | FGGALAALD | SR-SNETTLG | ----- | ----- |
| LSGGGLRAMI | LGSGALSAMD | ARHDNHTVLT | ----- | ----- |
| LSGGGYRAML | TGLGGIMGMM | NESTEASESE | ----- | ----- |
| LAGGGYRAML | TGLGGIMGMM | NESTEASQSE | ----- | ----- |
| PTLDAHNOI | MALEGLKTYR | LN---EFIIG | LNLVCH---- | ----- |
| LNELDFNTFL | SCICGLVKLR | R---ELMES | ILLPDN---- | ----- |
| DNDHDFKKLI | SAIYNLQQLR | SQLAKEFLID | LNELDE---- | ----- |
| QEGMACPQVL | PFDYN----- | -IEKPKEMP | ----- | ----- |
| RTPFNFFSVP | DDLWEKVNVD | TYQDGLLCDN | ----- | ----- |
| KSESSLKQWM | SSIIKMSTST | PWSKPNRFGS | FAPVRTNSFC | KFLVDGRDYF |

| | | | | |
|-------------|-------------|------------|------------|-------------|
| FDISVEDHLV | RTEDNYILTL | HRIPIISKNR | ----- | ----- |
| HGLSGFDKLI | LIP----- | ----- | ----- | ----- |
| DAGLTCQGAS | PS----- | ----- | ----- | ----- |
| -----NAFV | ATLIQPLN-- | ----- | -ANPSKLVSY | QSWEDASHID |
| -----NAFV | TTLIEPYN-- | ----- | -ADPSKVVSY | QTWEDASNIN |
| -----NAFV | TTIIQPKN-- | ----- | -ADPSKVVSY | QNWEDASNIN |
| -----TAIV | TTVIEPYN-- | ----- | -ADPSKVLVS | QTFEDSANIE |
| -----AAIV | TTVIEPFN-- | ----- | -ADPSKVVSY | QSWEDAANIE |
| -----NVIV | TTVIEPVN-- | ----- | -ADPSKIASY | QVFEDAARAD |
| -----NAIV | TTVIEPVN-- | ----- | -ADPSKIASY | QVFEDAARAD |
| -----NVIV | TTVMEPFN-- | ----- | -ADPSKLASY | QVFEDSAKAD |
| -----NVIV | TTVIEPVN-- | ----- | -ADPSKIASY | QVSENAARAD |
| -----LAIV | ATILEPPN-- | ----- | -GDKNKLISH | QAFENSPLLS |
| -----DCLT | INVVRPPG-- | ----- | -TKAGANLPV | MLWIFGGGFE |
| -----DCLT | INVVRPPG-- | ----- | -TKAGANLPV | MLWIFGGGFE |
| -----DCLT | INVVRPPG-- | ----- | -TKAGANLPV | MLWIFGGGFE |
| -----DCLT | INVIRPPG-- | ----- | -TRASAGLPV | MLWIFGGGFE |
| -----DCLT | INVVRPSG-- | ----- | -TKPGANLPV | MVWIFGGGFE |
| -----DCLY | LNVRFPAG-- | ----- | -TKPDAKLPV | MVWIYGGAFV |
| -----DCLY | LNVRFPAG-- | ----- | -TKPDAKLPV | MVWIYGGAFV |
| -----DCLY | LNVRFPAG-- | ----- | -TKPDAKLPV | MVWIYGGAFV |
| -----GGNL | TSIGKRDD-- | ----- | NLVGGMTLDL | PSDAP---- |
| -----GGNL | TSIGKRDD-- | ----- | NLVGGMTLDL | PSDAP---- |
| -----GPL | PS-----DV | ----- | ETKYGMALNA | TSYPD---- |
| ----- | ----- | ----- | ----- | ----- |
| -----LAGIL | QASSYIAG-- | ----- | LSGGSWLVGS | LASNNLNSVD |
| -----LSGIL | QSSSYIVG-- | ----- | LSGGSWLVGS | LASNDLIPVD |
| -----LGGLL | DSSTYVVG-- | ----- | LSGGNWLVS | LALNDWLSVG |
| -----LGGLL | QSTTYLAG-- | ----- | LSGGNWLVT | LAWNNWTSVQ |
| -----LGGLL | QGATYLAG-- | ----- | LSGGNWLST | LAWNNWTSVQ |
| -----LGGLL | QAATYLAG-- | ----- | LSGGNWLTTT | LSWNNWTSVQ |
| -----LGGLL | QSTTYLSG-- | ----- | ASGGNWLVT | LALNNWTSVQ |
| -----LGGLL | QSSTYLAG-- | ----- | LSGGNWLVT | LAYNNWTSVQ |
| -----LGGLL | QAATYMAG-- | ----- | LSGGNWLGT | LAYNNFTSVQ |
| -----LGGLL | QSSTYLSG-- | ----- | LSGGNWLGT | LAWNNWTSVQ |
| -----LGGLL | QSATYISG-- | ----- | LSGGSWLLGS | IYINNFTVD |
| -----IGGLL | QSSTYLAG-- | ----- | LSGGGWLVGS | LFSNNFESSIE |
| -----GIL | QSAMYIAG-- | ----- | LSGGSWLVGS | VAVNNFANIT |
| -----GLL | QSAMYISG-- | ----- | LSGGSWLVGS | VAINNFTNIT |
| -----GIL | QSSMYLTC-- | ----- | LSGGSWLVGS | VAVNNFANIT |
| -----GLL | QSAHYITG-- | ----- | ADGGSWLLSS | LAVNEFRTIQ |
| -----GLL | QASDYLVG-- | ----- | TDGSAWTVGG | IALNNFSTIN |
| -----TGGWL | DGVSYWAG-- | ----- | LSGGSWATGT | FMSNGGQLPT |
| -----TGGWL | DGVSYWAG-- | ----- | LSGGSWATGS | FMSNGGQLPT |
| -----QGEKMI | DYSENLNP-- | -----W | EKLEKEQSAQ | LDLGDVHRMC |
| -----SQFARI | HWQITVS-- | ----- | EKEEDEKDDT | LSFADVKKLC |
| -----NYVKML | LNKELLAGDN | GNVDGNEVDI | RKSHKHVREF | LSFNDILKYS |
| -----FDEFE | CSNLMITR-- | ----- | -PQGATNLPV | FVWIHGGGNI |
| ----- | LIVTRPKG-- | ----- | -VSANARLPT | VVWIHGGGNI |
| WLSLSEALLMA | KDVIIHDWW | LSPELYLRRP | VKGNQGFRID | RMLKSCAEKG |
| -----FN | NKVVYLHHG-- | -----LL | MCSDVWCCNI | ERHKNLFPVL |
| ----- | -SVFHLTN-- | ----- | ----- | ----- |
| -----SVS | KPILLVPG-- | ----- | ----- | ----- |
| CSPSY----- | ----- | ----- | ----- | --GMQFKSPA |
| CSPSY----- | ----- | ----- | ----- | --GAQFGSPL |
| CSPSY----- | ----- | ----- | ----- | --GSQLGAPL |
| CSPSY----- | ----- | ----- | ----- | --GMQYGAPW |
| CSPSY----- | ----- | ----- | ----- | --GMQFGAPL |
| CAPSY----- | ----- | ----- | ----- | --ALQFGSDL |
| CAPSY----- | ----- | ----- | ----- | --ALQFGSDL |
| CAPSY----- | ----- | ----- | ----- | --ALQFGSDV |
| CAPSY----- | ----- | ----- | ----- | --ALQFGSDV |
| CSPSY----- | ----- | ----- | ----- | --SMQVPS-F |
| VG----- | ----- | ----- | ----- | --GTSTFPPA |
| IG----- | ----- | ----- | ----- | --SPTIFPPA |
| IG----- | ----- | ----- | ----- | --SPTIFPPA |
| LG----- | ----- | ----- | ----- | --GSSLFPGD |

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|------------|-------------|------------|------------|------------|-------------|
| VG----- | ----- | ----- | ----- | ----- | ---GSSLFPPA |
| YG----- | ----- | ----- | ----- | ----- | ---SSAAYPGN |
| YG----- | ----- | ----- | ----- | ----- | ---SSAAYPGN |
| FG----- | ----- | ----- | ----- | ----- | ---SSASYPGN |
| ----- | ----- | ----- | ----- | ----- | ---PIS |
| ----- | ----- | ----- | ----- | ----- | ---PIS |
| ----- | ----- | ----- | ----- | ----- | ---SVV |
| ----- | ----- | ----- | ----- | ----- | ----- |
| ----- | ----- | ----- | ----- | ----- | ----- |
| ----- | ----- | ----- | ----- | ----- | ----- |
| DMSLQG---- | -LWELTHSFL | S---Y---- | ----- | ----- | ---YGIEHPIK |
| QLLREDK--- | -LWDIQNSLV | A---Y---- | ----- | ----- | ---YGVN-IVR |
| DIVNGKST-- | -IWQLQDSIL | N---P---- | ----- | ----- | ---SGMR-IDK |
| DIVNNMTEDD | SIWDISNSII | N---P---- | ----- | ----- | ---GGFM-IVT |
| AIVDNTTESN | SIWDISHSIL | T---P---- | ----- | ----- | ---DGIN-IFK |
| DIVDSQDNDS | AIWDISHSIV | S---P---- | ----- | ----- | ---GGIN-IFK |
| DILNNMQNDD | SIWDLSDSIV | T---P---- | ----- | ----- | ---GGIN-IFK |
| AIINNMTDDN | SIWDISNSIV | N---P---- | ----- | ----- | ---GGIN-IFS |
| QILEEGDKAD | AIWNITNSFL | N---P---- | ----- | ----- | ---YDKD-FSK |
| EIVDHMSESD | SIWNITKSIV | N---P---- | ----- | ----- | ---GGSN-LTY |
| KLQTHEAG-- | SVWQFGNSII | EGPDA---- | ----- | ----- | ---GGIQ-LLD |
| TLLSENK--- | -VWDFENSIF | KGPKE---- | ----- | ----- | ---AGLS-TVN |
| ----- | YLRDNVWNLE | HSVFA---- | ----- | ----- | ---PHGDNVVE |
| ----- | YLRDNVWNLE | HSVFA---- | ----- | ----- | ---PHGDNVIE |
| ----- | FLHDDVWNLD | HSLFA---- | ----- | ----- | ---PY-DDAFE |
| ----- | NISKSIWYTR | LGIFP---- | ----- | ----- | ---IEETHFGD |
| ----- | DFSK-LWAFN | HPLMY---- | ----- | ----- | ---PKSAIVFN |
| NLLEN----- | -LWNIDSNLV | FP----- | ----- | ----- | ---DDD |
| TLLEN----- | -LWNIDSNLV | FP----- | ----- | ----- | ---DDG |
| QMLHLNASME | FLEETFQKAD | ADHSG---- | ----- | ---KL | SFEFQHFVS |
| DKFHIVVSTG | QLLEFFQLAD | INHNG---- | ----- | ---LL | NYFEFEKFIK |
| KRLNINVNTN | HLQQIFDQVL | LLSSATTEKP | VSTPLFEKGL | NFEQFKQFVS | --- |
| AGNG----- | ----- | ----- | ----- | ----- | ---YCSDHNPV |
| EGSIYN---- | ----- | ----- | ----- | ----- | ---LIYEPQ |
| IKIFIVYIRN | VGNIVGTDSL | WTKHSMNLH | PNIHIIRSPN | QWLQNTYFWA | --- |
| HDLGYD---- | ----- | ----- | ----- | ----- | ---VWMGNNRG |
| ----- | ----- | ----- | ----- | ----- | ----- |
| ----- | ----- | ----- | ----- | ----- | ----- |
| ----- | ----- | ----- | ----- | ----- | ----- |
| TTVTTQIDMT | LIVPLLQNGY | YVIIPDYEGP | KSTFTVGRQS | GK----- | --- |
| STITTQIDMT | LIVPPLRSGY | YVVTPTYEGP | KATFAVGRQS | GQ----- | --- |
| STILTQLDMT | FIVPPLKSGY | YVVLPTYEGP | KSTFGVGRQS | GK----- | --- |
| STVATQIDMA | LMVPMLKQGY | YVVSPTYEGP | KSTFTVGRQS | GK----- | --- |
| SSVQTQVDMI | FIVPLLDKGC | FVVLPTYEGP | KSTFGVGRQS | GK----- | --- |
| TFVTQAEMY | LMAPLLDQGY | YVVSPTYEGP | KSTFTIGKQS | GQ----- | --- |
| TFVTQAEMY | LMAPLLDQGY | YVVSPTYEGP | KSTFTIGKQS | GQ----- | --- |
| TTIATQVETY | LLAPLLDQGY | YVVSPTYEGP | KSTFTVGRQS | GQ----- | --- |
| STLATQAETY | LLAPLLDKGY | YVVSPTYEGP | KSTFTVGRQS | GQ----- | --- |
| ETFQIQADLI | FISGLLSQGW | YVVVPTYEGP | NSVFPVGRQS | AY----- | --- |
| QMITKSIAMG | KPIIHVSVNY | RVSSWGFLAG | DEIKAEGSAN | AG----- | --- |
| QMVTKSVLMG | KPIIHVAVNY | RVASWGFLAG | DDIKAEGSGN | AG----- | --- |
| QMVSKSVLMG | KPIIHVAVNY | RLASWGFLAG | PDIKAEGSSN | AG----- | --- |
| QMVAKSVLMG | KPIIHVSMNY | RVASWGFLAG | PDIQNEGSGN | AG----- | --- |
| QMITASVLMG | KPIIHVSMNY | RVASWGFLAG | PDIKAEGSGN | AG----- | --- |
| SYVKESINMG | QPVVVFSINY | RTGPFGLGG | DAITAEGNTN | AG----- | --- |
| SYVKESINMG | QPVVVFSINY | RTGPFGLGG | DAITAEGNTN | AG----- | --- |
| GYVKESVEMG | QPVVVFSINY | RTGPFGLGG | DAITAEGNTN | AG----- | --- |
| LSSSTNSASD | GGKVVAATTA | QIQEFTKYAG | IAATAYCRSV | VP----- | --- |
| LSSSTNSASD | GGKVVAATTA | QIQEFTKYAG | IAATAYCRSV | VP----- | --- |
| QAMS----ID | GG-IRAATSQ | EINELTYTT | LSANSYCRTV | IP----- | --- |
| ----- | ----RDVSTS | ELDQFEFWVQ | YAAASYEAD | YT----- | --- |
| ----- | ----RDVSTS | ELDQFEFWVQ | YAAASYEAD | YT----- | --- |
| ----- | ----REVSQD | LFNQFNLFAQ | YSAAAYCGKN | ND----- | --- |
| QVEEWNVGN | QVASKRANF | NVSLTDIYGR | LLSYPLLTNT | ED----- | --- |
| NTAMWGNINL | QVQTKQLAGF | TVSITDVYGR | ALSHQLLTNF | DN----- | --- |
| TIAYYGLAQ | AVQAKEDAGF | QTSVTDTWGR | ALSYQFFED | DSG----- | --- |
| TIKRWDHISD | AVEGKQDAGF | NVSLTDIWGR | ALSYNFFPSL | YR----- | --- |
| TGSRWDDISD | DVQDKKQDAGF | NISLADVWGR | ALAYNFWPSL | HR----- | --- |
| TGSRWDHISD | AVEDKQKAGF | NVSLADVWGR | ALSYQFFPTL | YR----- | --- |
| TAKRWDHISN | AVESKQNADY | NTSLADIWGR | ALAYNFFPSL | NR----- | --- |
| SISRWDDISD | AVEEKKKAGF | NTSITDVWGR | ALSYNFFPSL | DE----- | --- |

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|-------------|-------------|-------------|-------------|------------|
| TLARWTAIGS | QVQGRDAGF | NVTITDLWSR | ALAYGWFPPTL | PN----- |
| TIERWESIVQ | EVQAKSDAGF | NISLSDLWAR | ALSYNFFPSL | PD----- |
| SAGYYKDLAD | AVDGKKKAGF | DTTLTDIWGR | ALSYQMFNAS | NG----- |
| RIQYWESEVAK | EVAKKKDAGF | ETSITDYWGR | ALSYQLIGAD | MG----- |
| NLAYYDLDLDD | EIDQKKDAGF | DTSLTDLWGR | ALSRKLVDAT | QG----- |
| NLNYYNDLRK | EIDQKKHAGF | DCSLTDLWGR | ALSRKLVDAE | RG----- |
| NFYIYQEWFE | QVLQKKNAGF | NVSITDLWGR | ALALKLVNPL | TG----- |
| LKNYYTNVVD | EVNQAAAAGF | NVSLTDYWGR | AIARHFVQQL | RG----- |
| -AHFYSSIMN | EVAEKANAGF | NISLSDYWGR | VISRTLGDIT | YG----- |
| KLSFYTELYT | ETNAKSDLGF | PIQITDVWGL | AIGSHVLPER | YQL----- |
| KLSFYTNLYT | ETNAKSDLGF | PVQITDIWGL | AIGSHVLEPE | YQL----- |
| LLKTRSEIVD | IFKEYTSGSD | -KMSLEQFRH | FLSTSQKARL | DSD----SIR |
| ILKNRKEVNM | IWSKFTKPPH | SHLSFENFFQ | FLITEQHEQV | DRQ----TAW |
| ILKDRKDLQE | IWDSLAAQKE | -VLQFDEIKN | FIINIQKENF | SDDDDNSTIN |
| PFVKHSIVAG | RPVLHVMIIEY | RLSAFGYLAV | PDTNGNWVGN | WG----- |
| FLVAESVRVG | KPIVHVCIIEY | RLGLAGFLTK | NGKGNWG--- | ----- |
| HHEKVVVIDE | TFAFIGGTDL | CYGRYDTFEH | VLRDDAESLL | DQNFPGKDYS |
| NKYSTAHLNK | PPKSNKFWDF | SIDEFAFFDI | PNSIEFILDI | TK----- |
| --LISNSIVH | NMAENFMQDD | EDKSDNKYTN | LLEIEYWIGV | KK----- |
| --TGTTGPQS | FDSNWIPLST | QLGYTPCWIS | PPPFMLNDTQ | VN----- |
| -----AT | LNSIRAALQT | GAFSGI-KKT | AKVALWGYSG | ----GSLATG |
| -----AT | LDSVRAILKS | GSFSGI-NED | AKVALWGYSG | ----GSLATG |
| -----AT | LDSIKAVLKT | KDFSGI-NDD | AKVVLWGYSG | ----GSFASG |
| -----AT | LDSIRAILKS | NKFTGI-KSD | AKVAMWGYSG | ----GSLASG |
| -----AT | LDSIKAVLKT | KDFSGI-NDD | AQVAMWGYSG | ----GTIAAG |
| -----AV | LNSIRAALKS | GKITNI-KDD | AKVVMWGYSG | ----GSLASG |
| -----AV | LNSIRATLKS | SKITNI-KED | AKVVMWGYSG | ----GSLASG |
| -----AV | LNSIRAALKS | GKITNL-AEN | AKVVMWGYSG | ----GSLASG |
| -----AV | LNSIRASLKS | GKITNI-AED | AKVLMWGYSG | ----GSLASG |
| -----SV | LDSIRGTIKF | FNSTG--NST | VKTALLGYSY | ----GAVASL |
| -----LK | DQRLGMQWVA | DNIAAFGGDP | TKVTIFGESA | ----GSMSVM |
| -----LK | DQRLGMQWVA | DNIAGFGGDP | SKVTIFGESA | ----GSMSVL |
| -----LK | DQRLGMQWVA | DNIAGFGGDP | SKVTIFGESA | ----GSMSVL |
| -----LH | DQRLAMQWVA | DNIAGFGGDP | SKVTIYGESA | ----GSMSTF |
| -----LH | DQRLGLQWVA | DNIAGFGGDP | SKVTIFGESA | ----GSMSVM |
| -----LH | DQRKLEWVS | DNIANFGGDP | DKVMIFGESA | ----GAMSPA |
| -----LH | DQRKLEWVS | DNIANFGGDP | DKVMIFGESA | ----GAMSPA |
| -----LH | DQRKLEWVS | DNIANFGGDP | DKVMIFGESA | ----GAMSPA |
| -----GNK | WDCVQCQK-- | ----WVPDGK | IITFTT---- | ----- |
| -----GNK | WDCVQCQK-- | ----WVPDGK | IITFTT---- | ----- |
| -----GAT | WDCIHCD--- | ----ATEDLK | IKTWS---- | ----- |
| -----AQV | GDKLSCSKGN | CPEVEATGAT | VSYDFSD--- | ----- |
| -----AQV | GDKLSCSKGN | CPEVEATGAT | VSYDFSD--- | ----- |
| -----APA | GTNITCTGNA | CPEVEKADAT | FLYSFED--- | ----- |
| -----EGD | AYLWSDVT-S | ASNFAQSHQMP | FPILISD--- | ----GRAPD |
| -----QGA | SFLWSDVT-E | TTSFQNNEMP | YPILAAL--- | ----GREPN |
| -----TGGA | NITWSSIR-N | LSSFQDHSMP | YPIVVAN--- | ----GRTPG |
| -----GGV | AYTWSTLR-D | VEVFQNGEMP | FPISVAD--- | ----GRYPG |
| -----GGV | GYTWSTLR-E | ADVFKNGEMP | FPITVAD--- | ----GRYPG |
| -----GGV | AYLWSDLR-E | SDVFKNAEMP | MPISVAD--- | ----GRYPG |
| -----GGI | GLTWSSIR-D | FPVFQNAEMP | FPISVAD--- | ----GRYPG |
| -----GGV | GYTWNTLR-D | VDVFKNGEMP | FPISVAV--- | ----GRYPG |
| -----AGA | GLTWSSLR-D | NEIFMNGEMP | MPISVAD--- | ----GRYPG |
| -----AGS | ALTWSSLR-D | VDVFKNGEMP | LPITVAD--- | ----GRYPG |
| -----GL | SYTWSSIA-D | TPEFQDGDYP | MPFVVAD--- | ----GRNPG |
| -----GP | AYTFSSIA-Q | TDNFQKAETP | FPILVAD--- | ----GRAPG |
| -----GP | NITFSSIR-N | QTFWQADYP | YPIIISD--- | ----SRLEE |
| -----GP | GITYSSMR-N | QSWFQADYP | YPIIVAD--- | ----SRLEE |
| -----GA | NTFSSVT-N | ETWFQDGEFP | FPIIIAD--- | ----NVIEG |
| -----GP | NLYSSVQ-N | ASWFQTAEYP | YPLIVTQGLT | ----GGLPDG |
| -----FP | NVSLSSIT-S | QEWYRNANFP | YPIITFA--- | ----TQNYG |
| -----SNTF | NLTFSSLPV | VSALGNASLP | MPIIIAADRK | ----RREAG |
| -----SNTF | NLTFSSLPV | VAALGNASLP | MPIIIAADRK | ----RREAG |
| TLYVFCNSND | DSKMGLEIFT | SFLLSPHNSP | VVPVIQDMS- | -----RPL |
| SYFIKYREPT | QLTMGQDGFT | KFLKE-QPYL | VEVKEELYS- | -----KPL |
| LIFQKYCSND | N-GWNKESLN | EYLLSSYSTP | YREITQQTQ | ----YYDYPL |
| -----AR | DQYTALQWIS | KHIVEFGGDP | SQITIGGES- | ----AGSIG |
| -----TW | DQYTGCCQWVN | RHIQDFGGDP | LNVTLTGESA | ----GSVAHV |
| NARIADFHDL | DKPFESMYDR | KVIPRMPWHD | VQMMTLGEP | RDLARHFVQR |
| -----VDK | VICIGFSQGS | AQMFAAFSL | EKLNRKVSHF | IAIAPAMTPK |

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-----F LQSKGCTVIT TKVPGFGSIE ERAMALD--- -----
----- -TEYMNVAIT ALYAGSGNKK LPVLTWSQGG -----

WAISLQSKYA PELKENLIGA AVG--GFATN ITAVAEAVDG TVFSGFIPLA
WAAALQPVYA PELQKNIVGA AVG--GFAAN ITAIAESVDG TIFSGLITLA
WAAVLQPEYA PELKDNLIGA ALG--GFAAN LTGIAESVDG EVFSGFIPLA
WAAALQPKYA PELKKNLIGA ALG--GFVTN ITATAEATDG TLFAGLVPNA
WAAALQPKYA QELKKNLIGA ALG--GFVIN ITATAEATDG TLFAGLIPNA
WAAALQPSYA PELGGNLLGA ALG--GFVTN ITATAQATDG TVFAGIVANA
WAAALQPSYA PELSSSLLGA ALG--GFVTN ITATAQAADG TVFAGIVANA
WAAALQPNYA PELGGNLLGA ALG--GFVTN ITATAEATDG TVFAGIMANA
WAAALQPDYA PELSRNLLGV ALG--GFITN VTATVEATDD TIFAGTAAV
WASIVQPNYA PEL--ELVGA AVG--CTIPN ITAFIEKVDE GPYSGLIVNI
CHILWNDGDN TYKGKPLFRA GIMQSGAMVP SDAVDGIYGN EIFDLLASNA
CHLIWNDGDN TYKGKPLFRA GIMQSGAMVP SDPVDGTYGN EIYDLFVSSA
CHLLWNGGDN TYKGKPLFRA GIMQSGAMVP SDPVDGTYGT QIYDTLVAST
VHLWNDGDN TYNGKPLFRA AIMQSGCMVP SDPVDGTYGT EIYNQVVASA
COLLWNDGDN TYNGKPLFRA AIMQSGAMVP SDPVDGTYGT QIYDQVVASA
HQLIAYGGDN TYNGKPLFHS AILQSGGPLP YHDSVVSGPD ISYNRFAQYA
HQLIAYGGDN TYNGKPLFHS AILQSGGPLP YHDSVVSGPD ISYNRFAQYA
HQLVAYGGDN TYNGKPLFHS AILQSGGPLP YFDSTVSGPE SAYSRFAQYA
-----SLL SDTNGYVLR DKQ----- -KTIYLVFRG TNS-----
-----SLL SDTNGYVLR DKQ----- -KTIYLVFRG TNS-----
-----TLI YDTNAMVARG DSE----- -KTIYIVFRG SSS-----
-----STI TDTAGYIAVD HTN----- -SAVVLAFRG SYS-----
-----STI TDTAGYIAVD HTN----- -SAVVLAFRG SYS-----
-----SGV GDVTGFLALD NTN----- -KLIVLSFRG SRS-----
TTIINLNSTV IELTPYEFGS WDP-----SL NEFVDTRYLG TKLDNGRP-T
TVLMNFNSTV FELTPYEFGS WDP-----SL RSFVDTKYIG TRLDDGAP-V
TYIINENSTI FEISPYELGS WDP-----SL KSFNSIQYLG SSVNNGNPNN
TQIIDLNATV FEFNPFEMGS WDP-----TL NAFTDVKYLG TRVSNGEFVN
TTVINLNATL FEFNPFEMGS WDP-----TL NAFTDVKYLG TNVTNGKPVN
TAVIDLNSTV FEYSPELGS WDP-----SL SAFTDVQYLG TRVSDGKPAE
TKVINLNATV FEFNPFEMGS WDP-----SL NSFANVKYLG TNVSNGVPLE
TQVVLNATV FEFNPFEMGS WDP-----TL HTFTDVRVYAG TNVTNGTPNV
TTVINLNATV FEMTPEFIS WDP-----SL NAFSDIKYLG TQVTDGKPE-
TTVINLNATL FEFTPEFEMGS WDP-----SL NAFTDVKYLG TNVTNGKPVN
ELVIGSNSTV YEFNPWEFGT FDP-----TI FGFVPLEYLG SKFEGGSLPS
DTIISLNATN YEFNPFEFGS WDP-----TV YGFAPTKYLG ANFSNGVIPS
EKAI PANTS I FEFTPEFEGT WDN-----GI KAFIPMEYVG THLKNVGP-P
ETAI PANTS I FEFTAYEFGT WDN-----GI KAFIPMEYVG THLLDGVGP-P
ETVIPLNDTV FEFTPIEFGT WDT-----GV ESFIPMEYTG THLINGIP-L
SNGTATNSSI YEISPYLTS FDN-----NV RSYTPTQYLG TNYSNGTA-V
EDISNVNTTF FEASPNVFGT FDH-----GI NSFIPTEYLG TTLNNGAS-S
ELVIAENATV WEFTPYEFGS WAFGSQYKSP GAFTPIEYLG TSVDDGSP--
ELVIAENATV WEFTPYEFGS WAFGSQYKSP GAFTPIEYLG TSVDDGSP--
NEYLISSSHN TYLLGKQFGG ESS-----I EGYIRSLQG CRCIEIDCWD
NHYFIASSHN TYLLGKQIAE TPS-----V EGYIQVLQG CRCVEIDIWD
NEYFISSSHN TYLTGRQVAG DSS-----V EGYIRTLQG CRCVEIDIWN
LHALMVHESM KPKECIIHN VILS----- SGTMDRMGTG TISENAFKPI
NMLIKDSMNG RKLFRNAVMM SGT-----LET ITPQPPKWA RLEEKVAVT
WNYLLRAKRP SRLTPLLTPP SDLTAEELKS LPMFEILREK STCETQILRS
GLHNRIVDTL AKSSPGFMYL FFG-----RKI VLPSAVIWQR TLHPTLFNLC
---AQLQKEV KKIESKDKRH SLN----- --LIAHSMGG LDCRYLICNI
----LVAQWG LTFPFSIRSK VDR----- LMAFAPDYKG TVLAGPLDAL

LNG----- LANEYPDFKK
LNG----- LANEYDPLKT
LNG----- IANEYPDFKK
LSG----- LANEYPEFKE
LNG----- LANEFPDFKK
LGG----- VANEYPEFKS
LGG----- VANEYPEFKS
LGG----- VANEYPEFKQ
LGG----- IANEYPEFKS
FNG----- LANEYRHFRD
CCG----- SAS---DKLA
CCG----- SAS---DKLA
CCS----- SAS---NKLA
CCG----- SAS---DKLA
CCG----- SAS---DKLA

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| | | | | |
|------------|------------|------------|------------|------------|
| GCD----- | ----- | ----- | ----- | TSASANDTLE |
| GCD----- | ----- | ----- | ----- | TSASANDTLE |
| GCD----- | ----- | ----- | ----- | ASAGDNETLA |
| ----- | ----- | ----- | ----- | FRSAITDIVF |
| ----- | ----- | ----- | ----- | FRSAITDIVF |
| ----- | ----- | ----- | ----- | IRNWIADLTF |
| ----- | ----- | ----- | ----- | VRNWVADATF |
| ----- | ----- | ----- | ----- | VRNWVADATF |
| ----- | ----- | ----- | ----- | IENWIGNLNF |
| GK----- | ----- | ----- | ----- | CYNGFDNAGF |
| SKR----- | ----- | ----- | ----- | CVNGFDNAGF |
| TDI----- | ----- | ----- | ----- | CVNFDNAGF |
| KGQ----- | ----- | ----- | ----- | CVAGYDNTGF |
| KGQ----- | ----- | ----- | ----- | CIAGFDNTGF |
| EGK----- | ----- | ----- | ----- | CIAGFDNVGF |
| RGK----- | ----- | ----- | ----- | CTAGFDNAGF |
| TGK----- | ----- | ----- | ----- | CVAGFDNTGF |
| TER----- | ----- | ----- | ----- | CINGFDDASF |
| KDQ----- | ----- | ----- | ----- | CVSGYDNAGF |
| NES----- | ----- | ----- | ----- | CIRGFDSAGF |
| GGK----- | ----- | ----- | ----- | CVEGLDQAGF |
| DHK----- | ----- | ----- | ----- | CIRNYDNAGF |
| DKS----- | ----- | ----- | ----- | CIHNYDNAGF |
| NES----- | ----- | ----- | ----- | CVRNFDNAGF |
| DGK----- | ----- | ----- | ----- | CVTQFDNVGF |
| NGS----- | ----- | ----- | ----- | CVINYDNFVF |
| NGT----- | ----- | ----- | ----- | CWKGFDQLSF |
| NGT----- | ----- | ----- | ----- | CWKGFDQLSF |
| GN----- | ----- | --GPVVCHGH | TFTSMIKFND | VIDAIRKYAF |
| GEN----- | ----- | --GPVVCHG- | FLTSAIPLKT | VIRVIKKYAF |
| GDSNTTTTV | IGTKDDDDKN | EYEPIVNHGR | TFTKPISFAN | VIRAIKKFAF |
| YDG----- | ----- | ----- | ----- | -----IKT |
| G----- | ----- | ----- | ----- | -----K |
| AGNWSLG--L | KETECSEQNA | YKLEQSEH | FIYIENQFFI | TSTVWNGTCV |
| IDIAN----- | ----- | ----- | ----- | -KILFNWKSF |
| KNR----- | ----- | ----- | ----- | NYDILSLTTI |
| AVS----- | ----- | ----- | ----- | APSVWQQTG |

| | | | | |
|-------------|------------|-------------|------------|------------|
| RLYGEVKLSA | RSTMCKGSON | CLAASLVGYP | MSQYFTGQNR | AFEKGWLLQ |
| AFYEELSDFA | VPEFKAGAEN | CLAENIFHYP | LHQYFTGPKR | AFEKGWLLK |
| RLYEEVKPGA | KADLQKGAEN | CLAASLISYP | MYQYFTGERR | VFEKGSLLLE |
| ILYQKVSAAA | TDNLRQGTET | CIGGAILYFA | EDQYFTGDDR | AFPGGYLLK |
| RMVEVVEKRY | EGALQQGTQH | CLGGAILHFA | FDQVFTGDHR | YFEQGYLLE |
| ILQSDTDK-- | KSVFEEFDGH | CLIDGVLNYI | GTSFLTGDHK | IFKTGWLDLK |
| ILQSDTDK-- | KSVFDEFDSH | CLADGVIDYI | NTSFLTGDNK | IFKTGWLDLK |
| ILQNDTDK-- | QSVFDQFDNH | CLADGVINYI | GKHFLSGTNK | IFKSGWNILK |
| ILQNDTNK-- | SSIFNKINNH | CLTDSFIKYV | GARFLTGDNK | VFKSGWNIFK |
| RLIH----- | FGALQPLG-- | CLFPICRKF | FQKMIGG--- | VYDA--QVLT |
| CLRGVSSDTL | EDATN--NTP | GFLAYSSLRL | SYLPRPDGPN | ITDDMYALVR |
| CLRSASSDTL | LDATN--NTP | GFLAYSSLRL | SYLPRPDGKN | ITDDMYKLVR |
| CLRGLSTQAL | LDATN--DTP | GFLSYTSLRL | SYLPRPDGAN | ITDDMYKLVR |
| CLRGLSQDTL | YQATS--DTP | GVLAYPSLRL | SYLPRPDGTF | ITDDMYALVR |
| CLRSISNDKL | FQATS--DTP | GALAYPSLRL | SFLPRPDGTF | ITDDMFKLVR |
| CLRSKSSSVL | HDAQNSYDLK | DLFGLLPQFL | GFGPRPDGNI | IPDAAYELFR |
| CLRSKSSSVL | HDAQNSYDLK | DLFGLLPQFL | GFGPRPDGNI | IPDAAYELFR |
| CLRSKSSDVL | HSAQNSYDLK | DLFGLLPQFL | GFGPRPDGNI | IPDAAYELFR |
| NFSDY-KPVK | GAKVH---- | ----- | ----- | ----- |
| NFSDY-KPVK | GAKVH---- | ----- | ----- | ----- |
| VPVSY-PPVS | GKLVH---- | ----- | ----- | ----- |
| VHTNP-GLCD | GCLAE---- | ----- | ----- | ----- |
| VHTNP-GLCD | GCLAE---- | ----- | ----- | ----- |
| DLKEINDICS | GCRGH---- | ----- | ----- | ----- |
| FMGTSSALEN | EAVLS-ITEA | N-IPSEFLKDI | IDDILVDPIL | KSNIDVSAYN |
| FMGTSSSLFN | IVLQQ-LNNM | P-IPPFKEL | ISKFTLDPVE | KLNIDIAQYN |
| IMGTSSSLFN | QILLQ-LDNY | S-INSIKMI | LEKVLTD-VS | DEEYDIAVYE |
| IMGTSSSLFN | QFLLQ-INST | S-LPSFIKNL | VTGFLDD-LS | EDEDDIAIYA |
| ITATSSSTLFN | QFLLR-LNST | D-LPSFIANL | ATDFLED-LS | DNSDDIAIYA |
| LMGTSSSTLFN | QFLLR-INDT | S-IPKFIRNL | ATHFLKD-LS | EDYDDIAVYA |
| IMGTSSSTLFN | QFLLR-INST | H-LPSFITRL | ARHFLKD-LS | QDFNDIAVYS |
| VMGTSSSLFN | QFLLQ-LNTT | D-LPSFLYNL | LHGFLTD-AS | DDYDDISIWA |
| IMGTSSSLFN | EFTMS-NDSA | V-AYTYLNTL | SSTLVKG-ID | KENNDIAMYA |

VIATSASLFN EFSLE-ASTS T-YKMINSF ANKYVNN-LS QDDDDIAIYA
VIGTSSSLFN QFLLQ-INTT S-LPSFIKDV FNGILFD-LD KSQNDIASYD
VMGTSSTLFN QFLLANISSY DGVDPVLEA VTSVLKE-IG AKRDDVSQII
VMGTSATLFN TFLLEWSQEV TS-NSTLYDI IHKVE-KLS EDQNDIAPY-
VMGTSATLFN SFLLDWNENV KK-NDTYDI LHAILE-DLS KHQDDIAPY-
LMGTSSNVFS GILPATNASL TASNNTFNNA VLSFLE-MLA EDQLDVGLY-
LVGTSSTRYN EALIDVSLRQ SR----MSRR LGFTLR-HMR INGSSVSFY-
MMGASSTYFN KIMRNFNDSS TK----NGRI IQQYLKGNFS ENGQQIISI-
VMGTSATLFN GAFLELNGTD S---GLLTNL ITAFLAD-LG EDQADISRI-
VMGTSATLFN GAFLELNGTD S---GLLTNL ITAFLAD-LG EDQADISRI-
VVSPYPLFIS LEIHCCPDQQ RQMVSYMKA FGDTLVMKPV TANESVLPSP
ITSPYPLIIS LEINCNDNQ KLASLIMREV LAEQLYFVGT RTDK--LPSP
IVSPWPLIIS LEIHCSPECQ IKVNVILKDI LGENMI IAPI DIDSVILPSP
LVGDINTCSA DELLEAQIKA GLDLGFYLD DFFPPDWRNV RFKVSRLLS
EVADLASLSD KELLDAQIKL NVAVCMTDD GDFEPGWKQ HLTDPWLDKL
LNKIGDALVD RIVKANQEKK PWKAFILIPL MPGFDSPVDT AEASSLRIM
NILPRQKIAS YAKLYSTTSV KSIVHWFQIL RSQKQMFEE SDNMLNSLTR
STPHRGSEMA DYVVD-----
SALTTALRNA GGLTQIVPT N-----

D-----EV FNKT----IE DNLLLKLDKT YLPQVPVLIY
E-----DI FNKS----IQ DNLLIGLNKT YLPQVPVLIY
D-----KT IGKT----LE DNLLIALSKE HMPQIPFVY
E-----EV VNKT----JS ENNLMQMDKD YLPDIPFVY
E-----EV FNRT----IS GNSLLYMQDE YLPDIPFVY
N-----PK IGKV----VE DNGLVYQ-KQ LVPKIPVFIY
S-----PT IAKI----VE DNGLVYQ-KQ LVPKIPFVY
N-----PT ISKI----VE DNGLVYQ-KQ LVPKIPILIY
N-----LV VSKI----VK DNGLVYQ-KQ LIPTIPVFIY
D-----ET IKET----IE INNLLST--R AVPQIPVFLF
EGKYANIPVI IGDQNDGTF FGTS----SL NVTDAQARE YFKQSFVHAS
DGKYASVPVI IGDQNDGTF FGLS----SL NVTNAQARA YFKQSFHAS
DGKYASVPVI IGDQNDGFL FGLS----SL NTTTEADAEA YLRKSFHAT
DGKYAHVPVI IGDQNDGTL FGLS----SL NVTDAQARA YFKQSFHAS
DGKCANVPVI IGDQNDGTV FALS----SL NVTDAQARQ YFKESFIHAS
SGRYAKVPYI SGNQEDEGTA FAPV----AL NATTPHVKK WLQYIFYDAS
SGRYAKVPYI SGNQEDEGTA FAPV----AL NATTPHVKK WLQYIFYDAS
SGRYAKVPYI TGNQEDEGTI LAPV----AI NATTPHVKK WLKYICSEAS
-----AGFLSS YEQV---VND YFPVQEQLT AHPTYKVIVT
-----AGFLSS YEQV---VND YFPVQEQLT AHPTYKVIVT
-----KGFLDS YGEV---QNE LVATVLDQFK QYPSYKVAVT
-----LGFWSS WKLV---RDD IKELKEVVA QNPNYELVVV
-----LGFWSS WKLV---RDD IKELKEVVA QNPNYELVVV
-----DGFTSS WRSV---ADT LRQKVEDAVR EHPDYRVVFT
PNPFKFS---SGS-NTA ISQS---KNL YLVDGGEDGQ NIPISPLLH-
PNPFHKS---NNS-DTK IAQS---RTL YLADGGEDGQ NVPLPLLIH-
PNPFPGA---DSAGIKS ITTN---DTL YLCDGGEDLQ NVPFYPLIQN
PNPFKDTSY--IQDNFSKS ISES---DYL YLVDGGEDNQ NIPLVPLVQD
PNPFKEANF--LQKNATSS IIES---EYL FLVDGGEDNQ NIPLVPLLQK
PNPFRDADY--VNNRNSKS LSES---EYL FLVDGGEDGQ NVPLVPLIQQ
PNPFKDTKF--LSDSYTTS IVDS---DSL FLVDGGEDDE NVPVPLLIQK
PNPFYEITN--IPSNYSQS ISED---DTL YLVDGGEDGQ NIPLTPLLQT
PNPFKGSKY--VDSNYTTS IVDS---DSL FLVDGGEDLQ GIPFVPLLQK
ANPFKDTF--VDRNYTSS IVDA---DDL FLVDGGEDGQ NLPLVPLIKK
PNPFYKYN---EHSSP YAAQ---KLL DVVDGGEDGQ NVPLHPLIQP
PNPFLDWN---NRTNP NADT---LEL DLVDGGEDLQ NIPLNPLTQP
PNPYQNF---TTNTTVKNP FERF---DTI DLVDGGEDDE NIPWPLLHP
PNPYQNY---TTSNTSVVNA FEPY---DTI DLVDGGEDRE NIPWPLLHP
PNPYQGYG---NASNTTNP LEPY---PII ELIDGGSSE GIPFVPLLHP
PNPYTDATDI AGNATAVSED IVDT---PYL DLFDDGGYDQ NIPIWPLLQK
PNPFQGV---ESANSDAANN LGSS---SSL NLVDFTLTGE KIPWPLLQK
PNTFSNYN---SGENP IYNL---TYI TLVDAGETNQ NIPLEPLLVP
PNSFSNYN---SGENP IYNL---TYI TLVDAGETNQ NIPLEPLLVP
EDLLNKILK VKCSATPLHQ FSTD---ILK VGITDSSTDT TESSELEN--
RELKHKILK SKKTSEATRG LSVN---EPF PSSFSSYES ANEQELRMKD
AELKHKFIK VKKTSFQNL IETENGSTTT STTTTTTTTT TTTTATSLSE
DIVVDG-----TN FKNKIN--PA VRVTPENDFD HKVKFLYNIS
IIS-----DC KDEG--MLYF LPVNAQDDEE LLAKVAKSPV
QFQYQISIRG EHSTFSKLLK LNIDP-AQYI OFFSLRKWST FAPNERLITE
PYQIAN-----FP TRTN---IKI PILLIYGID SLVDIVMCK
-----LF ENLNALRVSQ KILPICFYQL

-----L YSATDEIVQP QVSN SPLDSS YLFNGKNVQA

HGTIDEIPI KDAN----- --AQYQI WCD RGIQSLEFAE
HGTVDEIPI KDPH----- --AQYQL WCD WGI ESLEFAE
HGTIDKIPI KDSI----- --KIYKN WCD WGIGSFEFSE
HGALDSIVPI SNVH----- --VTYKN WCD WGINSFEFSE
HGSLDGIVPI PDVH----- --GVYKN WCD WGIDSFEFAE
HGSIDQIVPI VDTK----- --KTYQN WCD AGISSLEFAE
HGSIDQIVPI VNVK----- --KTYQN WCE GGISSLEFAE
HGAI DQIVPI VNVK----- --KTYQN WCD AGIASLEFSE
HGSM DQISPI LNP K----- --KTYQN WCD AGISSIEFAE
HSKF NEMSPF LEIL----- --KLEKL WCS QLGVNLEIAE
DAEID TLM TA YPGD----- --ITQGS PFD TGILNALT PQ
DAEID TLM AA YPQD----- --ITQGS PFD TGIFNAIT PQ
DADIT ALKAA YPSD----- --VTQGS PFD TGILNALT PQ
DAEID TLM AA YTS D----- --ITQGS PFD TGIFNAIT PQ
DAEID TLM AA YPSD----- --ITQGS PFD TGIFNAIT PQ
EASID RVLSL YPQT----- --LSVGS PFR TGILNALT PQ
EASID RVLSL YPQT----- --LSVGS PFR TGILNALT PQ
DASLD RVLSL YPGS----- --WSEGAPFR TGILNALT PQ
GHS LGGAQAL LAGM----- --DLYQREPR LSPKNLSIFT
GHS LGGAQAL LAGM----- --DLYQREPR LSPKNLSIFT
GHS LGGATAL LCAL----- --DLYQREEG LSSSNLFYLT
GHS LGAAVAT LAAT----- --DLR----G KGYP SAKLYA
GHS LGAAVAT LAAT----- --DLR----G KGYP SAKLYA
GHS LGGALAT VAGA----- --DLR----G NGY-DIDVFS
-RNVS AIFAF DNS----- --NDVLN-WP DGTS LVKTYE
-RKVS AIFAF DQS----- --ADKNN-WP DGSALIKT FE
KRGVDVIFAF DNS----- --ADTNSSWP NGTSIQET YK
ERNVDVIFAL DNS----- --ADTDYWP DGASLVST YE
ERELDVIFAL DNS----- --ADTDDYWP DGASLVNTYQ
ERDL D VIFAL DNS----- --ADTEENWP DGASLMHTYR
ERD VDI IFAV DNS----- --ADMRLAWP DGSSLVHTY E
EREIDVIFAL DNS----- --ADTDQSWP DGFSLTQTYA
ERDLDI IFAI DVD----- --TETSDNYP AGGPPMMKTYE
ERDL D VV FAL DIS----- --DNTDESWP SGVCMNTY E
ERHVDVIFAV DSS----- --ADTDYFWP NGTSLVATY E
VRAVDVIFAV DSS----- --ADVTN-WP NGTALRATY E
QRFVDVIFAV DATY----- --DDSN-GWP DGSSIVTTY E
QRFDVVFVFAI DSTY----- --NDPY-GWP LGSSIVATY E
QRD VDVIFAI DGGY----- --QSATSGWP DGSSLVSTY E
ERKLDVVFVAF DSSG----- --DTSN-FWP NGSSLVATY E
GRD VDVIVAV DNG----- --DDSEWLWP NGNSLVQTY E
TRD VDAIVAF DSS----- --YTDYIWP NGTALRTY E
TRD VDAIVAF DSS----- --YSDYIWP NGTALRTY E
----SELTGL RKGK----- --RRMKNIIV QELQQLAPYA
DSTNSSSATN SSSM----- --QRIKRIGL KKHADIINDV
DNENKSNSS STSS----- --FIIRRRKN KSPKIINELS
TEDTWEDYHY KMML----- --FKGDETFI RGNQQLELLF
GKEISELYGI KEGG----- --DIKSACLD LKTDFNFNYF
QLYVHAKILI ADDRRCIIGS ANINERSQLG NRDSEVAILI RDTDLIKTKM
NLPFNSVFDV KVDN----- --YEHLDLIWG KDADTLVIAK
TTAYMKYFNL VTPN----- --SPKVSYFS YGCSFVPKWY
QAVCGPLFVI DHAG----- --SLTSQFSYV VGRSALRSTT

DLSAGHLAET FTG----- ----APAAL SWIDARFSGK PAVNGCQRTI
DLSTGHLAET FTG----- ----APAAL AWIDARFDGK TPIQGCSTTT
DKSNGHTTET VVG----- ----APAAL TWIDARFAGK PAVEGCSFTT
DLLNGHITET IVG----- ----APAAI TWLEARFDGE PVVKGCKKTS
DSLNGHLTEI VVG----- ----APAAI TWLDARFDGQ PVVEGCKKTT
DASNGHLTEA IMG----- ----APAAL TWIIDRFDGK QTVSGCQHIO
DGTNGHLTET VVG----- ----APAAL TWIIDRFNGK QTVSGCQHDK
DATNGHITET IVG----- ----APVAL TWIINRFNGK QTVSGCQHVK
DLTNGHFTES IVG----- ----APAAL TWIIDRFSNK PPVDCQHVV
DMSYNHMEVA FSG----- ----MPAAI TWIEKRWN-N STLGGCKHVH
FKRISAVLGD LGFTLARRYF LNHYTGGTKY SFLSKQLSG- LPVLGTFHNS
FKRISAVLGD LAFIHARRYF LNHFQGGTKY SFLSKQLSG- LPIMGTFHAN
LKRINAVLGD LFTLSRRYF LNHYTGGPKY SFLSKQLSG- LPILGTFHAN
FKRISALLGD LAFTLARRYF LNYQGGTKY SFLSKQLSG- LPVLGTFHGN
FKRIAVLGD LAFTLPRRYF LNHFQGGTKY SFLSKQLSG- LPVIGTHHAN
FKRVAAILSD MLFQSPRRVM LSATKDVNRW TYLSTHLHNL VPFLGTFHGN

| | | | | |
|------------|-------------|--------------|-------------|-------------|
| FKRVAAIISD | MLFQSPRRVM | LSATKDVNRW | TYLSTHLHNL | VPFLGTFHGN |
| FKRIAAIIFD | LLFQSPRRVM | LNATKDVNRW | TYLATQLHNL | VPFLGTFHGS |
| V----- | ----GGPRV- | -----GNPT | FAYYVESTG- | IPFQRTVHKR |
| V----- | ----GGPRV- | -----GNPT | FAYYVESTG- | IPFQRTVHKR |
| Q----- | ----GQPRV- | -----GDPA | FANYVVSTG- | IPYRRTVNER |
| Y----- | ----ASPRV- | -----GNAA | LAKYITAQ-- | GNNFRFTHTN |
| Y----- | ----ASPRV- | -----GNAA | LAKYITAQ-- | GNNFRFTHTN |
| Y----- | ----GAPRV- | -----GNRA | FAEFLTQVQ | GTLYRITHTN |
| R--QFSS--Q | GNGIAFPYV- | -----PDQY | TFRNLNLTSK | PTFFGCDAKN |
| R--QFSS--Q | GDGIAFPYV- | -----PDQN | TFRNTNLTSK | PTFFGCDAQN |
| R--QFSK--Q | GKGTFFPFA- | -----PDYK | TFLDKNMGDK | PVFFGCNSSD |
| R--QFSS--Q | GLNMSFPYV- | -----PDKR | TFVNLGLADK | PSFFGCDAQN |
| R--QFGS--Q | GKLNLSFPYV- | -----PDVN | TFVNLGLNKK | PTFFGCDAKN |
| R--QFGF--Q | GQGVTFPSV- | -----PGTD | TFVNLGLNKK | PTFFGCDAKN |
| R--QFVK--Q | GQGMSFPYV- | -----PDTN | TFVNLGLNKK | PTFFGCDAKN |
| R--QFGL--Q | GKGIAPYV- | -----PDVN | TFVNLGLNTR | PTFFGCDAKN |
| R--QFSK--Q | GKGMAFPYV- | -----PDMT | TFVNLGLGKK | PSFYGCDAKN |
| R--QYSK--Q | GKGMAFPYV- | -----PDVN | TFVNLGLTNK | PTFFGCDAKN |
| R--SLNSSGI | ANGTAFPAV- | -----PDQN | TFVNLGLSTR | PSFFGCDAKN |
| R--TFGS--I | SNGTLFPSI- | -----PDDW | TFVNLGLNTR | PSFFGCDAKN |
| RIITYNANKS | VDVRGFPYI- | -----PDED | TIISLGLNTH | PTFFGCDAKN |
| RVVTFNANKS | VDVRGFPYI- | -----PDEN | TIISLGLNTR | PTFFGCDAKN |
| RVLATNSSG- | --VRGFPYI- | -----PDTN | TFVNLGLNTH | PTFFGCDAKN |
| RVTQRASDAV | YDVEDFVHV- | -----PTPE | TFVNLGLNTR | PTFFGCDAKN |
| RVVAAQAAGN | TNVKGFYV- | -----PSQQ | SFVSLHFNDR | PVFFGCDAKN |
| R-AKVLAEHE | NTRVLMPEV- | -----PSMN | GFVNGGYSR | PTFFGCDAKN |
| R-AKILAEHE | NTRVLMPEV- | -----PSMN | GFVNGGYSR | PTFFGCDAKN |
| R-----SLK | FRNFSLPES- | -----KTYS | HIFSFSER-- | TIKKHGKAMV |
| SNISGIHGK | FRNFSLPES- | -----KTIA | HCFSLNERKV | EYMIKDKHLK |
| NLGIYTQGIK | FRNFSLPES- | -----KTFN | HCFSLGEKSI | NRMKDDDKK |
| EQENIPVWRQ | LFDQIHPND- | -----PSRL | CHHAVDLYYM | WDNWEMPEDK |
| NHLLFKKMEE | ARNNGSTSR- | -----VY | RLAVDEPNPH | NPDQRAHHA |
| NGDDYAGKF | PWELRQRLMR | EHLGCDVDLV | EFVEKKFERF | EKFAAKNYEK |
| VLRFIEFFNP | GNVSVKTNQL | LP--SASLVE | ELPSTTWKTT | HPTHGLSYRT |
| N----- | ----- | ----- | VFCTPWKIVY | ERSKGCNDG |
| G----- | ----- | -----QA | RSADYGITDC | NPLPANDLTP |
| R----- | ----- | ---SSNVLYP | GISIT--IRI | YFEGISKTI |
| R----- | ----- | ---LTNLLYP | NTSDS--THS | YFLGIYQAVF |
| R----- | ----- | ---ASNFLYP | NISES--AAS | YFKGIYQITL |
| R----- | ----- | ---ITNFSYP | NISDS--TSS | IFEGILNSVT |
| R----- | ----- | ---ITNFSYP | NISDS--TRN | FFKGIILDSL |
| R----- | ----- | ---FSNLEYP | NIPSS--IAN | YFKAAMDVVL |
| R----- | ----- | ---LSNFQYP | NISSS--ILK | YFKVALDTMM |
| R----- | ----- | ---TSNFEYP | NIPPS--ILN | YFKAALNILI |
| R----- | ----- | ---TTNYEYP | NVSSS--ILD | YFKAAMDVVA |
| R----- | ----- | ---LSNFEYP | GIAPF--LSQ | YFRSSLQMV |
| D----- | ----- | ---IVFQDYL | LGSGSLIYNN | AFIAFATDLD |
| D----- | ----- | ---IVWQDYL | LGSGSVIYNN | AFIAFATDLD |
| D----- | ----- | ---IVWQHFL | LGSGSVIYNN | AFIAFATDLD |
| D----- | ----- | ---IIWQDYL | VGSGSVIYNN | AFIAFANDLD |
| D----- | ----- | ---IVWQDFL | VSHSSAVYNN | AFIAFANDLD |
| E----- | ----- | ---LIFQFNV | NIGPANSYLR | YFISFANHHD |
| E----- | ----- | ---LIFQFNV | NIGPANSYLR | YFISFANHHD |
| D----- | ----- | ---LLFYQYV | DLGPSSAYRR | YFISFANHHD |
| ----- | ----- | ---DIVPHVPPQ | SFGFLHPGVE | SWIKSGTS-N |
| ----- | ----- | ---DIVPHVPPQ | SFGFLHPGVE | SWIKSGTS-N |
| ----- | ----- | ---DIVPHLPPA | AFGFLHAGEE | YWITDNSPET |
| ----- | ----- | ---DPVPKLPLL | SMGYVHVSPE | YWITSPNN-- |
| ----- | ----- | ---DPVPKLPLL | SMGYVHVSPE | YWITSPNN-- |
| ----- | ----- | ---DIVPRLPPR | EFGYSHSSPE | YWIKSGTL-- |
| LTSLT----- | ----KDIYD | VPLVIYLANR | PFTYWSNTST | FKLTYYDDNER |
| LTSLT----- | ----ENIYD | VPVVIYLANR | PFTYFSNIST | FKLYSDTER |
| LEDLVAWHEN | ---DKINVD | VPLVYVTSNT | RMSYNSNFST | FKLSYSDQEK |
| LTDLN----- | -----YI | PPLVVIYIPNA | RHSYNSNTST | FKLSYTDDE |
| LTDLE----- | -----YI | PPLVVIYIPNS | RHSFNGNQST | FKMSYSDSER |
| MTDLE----- | -----YI | PPLVVIYIPNS | RHSYNGNTST | FKLSYSEKER |
| LTDLQ----- | -----YI | PPLVVIYIPNA | EYSFNNSQSA | FKLSYSESQR |
| LTDLE----- | -----SI | PPLVVIYIPNT | RESFNNSNTST | FKMSYSTSER |
| LTDLE----- | -----YI | PPLVVIYIPNS | YHSFNSVST | FKLNYSYSER |
| LTDLE----- | -----YI | PPLVVIYIPNT | KHSFNGNQST | LKMNYVTER |

QTGPS----- -PLVVYIPNA PYSYHSNIST FQLSTDDAER
 FTLNAN----- -QKV PPLIVVYPNA PYTALSNVST FDPSTYMSQR
 TTAGN----- --HTVDNNT PPLLVYFPNY PWVYYSNIST FTMSMNDTSL
 TTAGN----- --HDVDNNT PPLLVYFPNY PWTYYSNIST FTMSMDDKMA
 TTAGN----- --HTVNDDT PPLVVYFPNY PWTMYANVTT YTVQLEDTSL
 TTRGD----- --VPVDHNT PPLVVYMPNT PWTMKSNLVD HRYRIANSEI
 TTAGN----- --HTVTRDT PPLVIYLPNV PYNFYFTNIST DRTYYTEDMI
 ----- TPLIIVVPSY PWSFAANTST YQLSYENDEA
 ----- TPVIIYIPSY PWSFAANTST YQLSYENNEA
 PRLSK----- -HNLRYLCRVY PGPLRVGSTN FNPQVYWRLG
 ISLDK----- -HNRRLMRVY PHVLRKSSN FNPIPFWKAG
 ISLDK----- -HNRRLMRVY PSGTRKSSN FNPLPYWSHG
 H----- --AVARQYQD TLTKFVYQD PWPVDKLHYV
 D----- ---VLYMFNS TKFNEHGDKL SRLFQSHFLR
 LHLSKEGDS GNNWSDREMI DSAMIELGYR EIFGCKFSPQ WKSCHGNSVD
 HSADR----- SPLSVQADEA DEVHNADNSR FLRRVFSTSA
 ----- ---LVTINSS KWGEYRGLK DMDHLDVINW
 E----- -QKVA AAALLAPAAA AIVAGPKQNC

GVNLSGSVNA D-KSISNKFF AYIRKYI---
 GTPLGPGING DNITINSGLL GLVSSII---
 RSKLGSVTS DDVSVN-GLR SLYHT---
 GSELGPGVTS DNITLD-GLT GFLGNFIDLK
 ASQLGPGVTS DNVTL-GLT GFMGGLSKFK
 HLGGLPQVQK DQVSPE-GIK KLGSIEMRWL
 SNGLGSDIQK DKITPD-DLR KF--LLGW-
 QKGLGPDQK DQVNP-GLK KISILV---
 QQGLGPNQK DQLEIK-SNL
 NNNRYFNNTT R-----
 PNTAGLLVKW PEYSSSSQSG NNLMMINALG
 PNTAGLLVNW PKYTSSSSQSG NNLMMINALG
 PNTAGLSVQW PKSTSSSQAG DNLMQISALG
 PNKAGLWTNW PTYTSSSQSG NNLMQINGLG
 PNKAGLLVNW PKYTSSSQSG NNLQINAGL
 PNVGTNLLQW DQYDE---G KEMLEIHMTD
 PNVGTNLLQW DQYDE---G KEMLEIHMTD
 PNVGTNLKQW DMYTDS---G KEMLEIHMTD
 VQICTSELET KDCS----- --NSIVP---
 VQICTSELET KDCS----- --NSIVP---
 VQVCTSDLET SDCS----- --NSIVP---
 ATVSTSDIKV IDGDVDFDGN TGTGLPL---
 ATVSTSDIKV IDGDVDFDGN TGTGLPL---
 VPVTRNDIVK IEG---IDAT GGNNQPN---
 QGMISNGFEI ATRSSGSLDD EWAACVG---
 QGMISNGYDV ASRLNGKLDN EWAACVG---
 FGAIKNGFET VTRNNLTDD EWAACVG---
 LKMIKNGFEA ATRGNLTDD SFGCCVA---
 LGMKNGFEA ATMGNFTDD DFLGCVG---
 LGVIRNGFEA ATMNNLTADS NFAGCIG---
 RSMIQNGFEI ATRNNFTDD EFMGCVG---
 FKMIQNGFEA VTMKNLTDE NFMGCIS---
 VMIRNAFEA TTRNNLTEDA DYVTCVG---
 LGMIRNGFEA ATMGNFTDD NFLGCIG---
 DNIIILNGYEV ATMANSTLDD NWTACVA---
 NDIIGNGWNS ATQNGTLDS EWPTCVA---
 SGILENAALS ATQNN---SD SFAVCLA---
 NGILENAFMS TTQNN---NE SFAVCLA---
 SGMIEAAVA ATQNN---SD SFAVCA---
 QALIQNGFVA TTQDN---ST DFASCLA---
 QLLTNGLIS STVDN---DT YFGQCF---
 NEMLLNGMRS LTLNHS--VP TWPTCFA---
 NEMLLNGMRS LTLNHS--VP TWPTCFA---
 VQMVATNWQT YDTGLQINDA LFIADPP--- --TGYYLLK PP-----
 VQMVATNWQT NDIGQQLNLA MFQILDHQP GSFKSGYVVK PKKLLPVV--
 VQMVATNWQT YDLGQQLNEA LFENKIF--- --QGYVVK PSVLRKPT--
 HDNQFEILDK SQFGDFRNV ALKFLLG---
 LAYGLEPWH RNFVYRNGG YQQLPLSELN
 DGSTQCCINE KEVGREDEV YEKFFNSVDY GKSSRKRTPL PKHNFASLGL
 IDEDNENEHQ DDTEDQIHKE QORRLSA---
 KNKLQDDWSK FFRTTTVGEK VDILNFY---
 EPDLMPYARP FAVGKRTCSG IVTP-----

Appendix 2: Multiple sequence alignment of lysophospholipase from fungi with comparison to protein sequences of cytosolic phospholipase A2 from human. The conserved **oxyanion hole (GGG)** is shown in green. The dyad active sites (Ser- Asp) are shown in red. Eight conserved cysteine residues in lysophospholipases were high-lighted in green and were not conserved in cytosolic phospholipase A2. Conserved residues between lysophospholipase and cytosolic phospholipase A2 are shown in black highlights.

```

Q9UTH5 -----MYFQSFYFLALLLATAVYGVQVAS---- 23
P78854 -----MLFRGLSLWMLFLASCLSLALALP---- 23
O13857 -----MRPG----- 4
Q9P327 -----MYVNYIGLFAFVQISLTLAYPPGRVEI 27
Q9Y7N6 -----MKLSSFGLFLALQLLPALGLPSR---- 23
Q9UWF6 -----MILHLLILLIINYCVATSPTN---- 22
O93795 -----MLVWQSILLFLVGCVLKSPSTN---- 22
Q9UVX1 -----MKVNLKLIIGSILISQAQAIWPFDSGSSSSSDSSPS 37
Q11121 -----MNLKEW-LLFSDAVF--FAQGTLAWSPSN---- 26
P39105 -----MKLQS--LLVSAAVLTLTENVNAWSPNN---- 27
Q8TG07 -----MQLQD--LVVTVSLLAAFNGGVEAWSPTN---- 27
O59863 -----MWFLNSVNLFLVCSVALHLDVNAWSPTN---- 30
Q08108 -----MIRPLCKIIISYIFAISQFLAANAWSPTD---- 31
Q03674 -----MQLRN-ILQASSLISGLSLAADSSSTTGD---- 28
Q8TG06 -----MQLS--VLIASVLAAGAAVDAAS----- 21
P39457 -----DITFAG-----VQRALPNAPDG---- 17
O42790 -----MHLPSLLIAAPLLANVSAEPIRIPQRDVSVVVSTSQQLAVRALPDSPPSGG--- 50
Q9P8P2 -----MSIATGTFAFSLFATIAFAVPPETPRIELQAERG--- 34
Q9P8L1 -----MSIITAFALSLATTAFVPPETPRIELQAERG--- 34
1CJY MSFIDPYQHIIVEHQYSHKFTVVVLRATKVKGAFGDMLDTPDPYVELFISTTTPDSR--- 57

Q9UTH5 -----PELHSLSRRNWK-----KPP 38
P78854 -----AAEDDGSVKVFK-----RAK 38
O13857 -----MHDTPLSLMQK-----REALAI 21
Q9P327 -----SEIYDFEESSSYKQQDIDTQSVLYT-----LSKRKP 57
Q9Y7N6 -----IDEVDVSDPELIGLLKPDNVKPNANSI 50
Q9UWF6 -----
O93795 -----
Q9UVX1 ETGSSGGTFFFDLFGSGSLLTQSSSAQASSTKSTSDSASSTDSLFSSSNSGSSWYQTFL 97
Q11121 -----
P39105 -----
Q8TG07 -----
O59863 -----
Q08108 -----
Q03674 -----
Q8TG06 -----
P39457 -----
O42790 -----
Q9P8P2 -----
Q9P8L1 -----
1CJY -----KRTRHFNNNDINPVWNETFEFILDPN 82

Q9UTH5 PFPSTN--ASYAPVIRSCDSSEIMVNSLPRGELPDLENDFIEKRLSNANEALTTFLQSK- 95
P78854 KHSTKQEGPSYAPYYVDPS-DNIVESLSSNEIPSAESEYLSTRSTIITNTAMKDFLRNA- 96
O13857 SLSKRDSVGSYAPYNVTPS-DYMLRPASDG-ISSGQSFIDKRIKINTQMRSFISNT- 78
Q9P327 ALVKRSTDASYAPFNVTSN-DNLLRPASEG-LNEGEQSYINKRISKVNSSELRSFISKT- 114
Q9Y7N6 PLSKRSTSPSYAPYTVAPS-GSLLRPASDG-LSTGEQEFVDKRVSKVNSALESFISKT- 107
Q9UWF6 -----GYAPGVSFP--SSQLIRSGSQGINPNEQSYINARIPIAKQALSFLHN-- 69
O93795 -----LYTPGYVQFP--EGKLTSSLDGINSNEKAYIDRRYANAKSELRSFLHN-- 69
Q9UVX1 DGDSGDQKTDYAPFNLTCP--SKKTFIRTASELSQEKDYIHKRQETTNNKLIIDFLSKR- 154
Q11121 -----SYTPANVSD-EDINLIRQAS-GPSDNETEWLKKRDVYTRREALRSFLDR-- 73
P39105 -----SYVPANVTD-DDINLVRAS-GLSDNETEWLKKRDVYTRREALRSFLNR-- 74
Q8TG07 -----SYVPANVTP-NDINLNRAT-GLSQSEIDWLKKRDVNTREALRSFLKR-- 74
O59863 -----GYAPGVVDSD-ENINLVRKAD-AVSDDEADWLKVRHESVTPALKDFLQR-- 77
Q08108 -----SYVPGTVSD-DDINLVRAT-SISQNESAWLEKRNKVTVALKDFLTR-- 78
Q03674 -----GYAPSIIPSPSDDTSLVRNAS-GLSTAETDWLKKRDVYTRREALRSFLSR-- 76

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Q8TG06 -----YTPQNVSCP--DNANFIRNAADGLSPAKEWLLKRRDPITRDALQTFRR-- 68
P39457 -----YVPTVSCP--ASRPTVRSAAKLSTNETSWLEVRGKTLKALDKDFFGH-- 63
O42790 -----YAPAVVDCP--KTKPTLRKAVDLSNEEKWLSIRKNTIQPMRDLKLR-- 96
Q9P8P2 -----LGDKSYAPWQVDCP--SNVTWIRNATTGLGSGERAYIEAREKLVQPVIEQMMMAARG 88
Q9P8L1 -----LGDQSYAPWQVDCP--SNVTWIRNATTGLGTGERAYIEAREKLVQPAIEQMMMAARG 88
1CJY QENVLEITLMDANYVMDLETGTATFTVSSMKVGEKKEVPFIFNQVTEMLVEMSLVSSCP 142

* :: :

Q9UTH5 -----NTTADLDLSS-----IVDNGPRLGIAVSGGGWRSM 126
P78854 -----NLPGLNADT-----LSGSEGPSIGIALSGGGLRAM 126
O13857 -----GLDVDVNSV-----INDSDGPRLGLAFSGGGLRAM 108
Q9P327 -----GLNVLDLKV-----VNSSDGPRLGIAFSGGGLRAM 144
Q9Y7N6 -----GLKIDTKSV-----LNDDTGPRLGIAISGGGFPM 137
Q9UWF6 -----ANLQNFVDVS-----FLAH-----SNPTIGLAFSGGGYRAM 100
O93795 -----AKMVDFDVG-----FLN-----SNPTIGLAFSGGGYRAM 99
Q9UVX1 -----ANLSDFDAK-----FINDNAPNHNTIGLSFSGGGYRAM 189
Q11121 -----ATSNFSDSS-----LVSQLF--SNASDIPRIAVACSGGGYRAM 109
P39105 -----ATSNFSDTS-----LLSTLFG--SNSSNMPKIAVACSGGGYRAM 111
Q8TG07 -----VTSNFTSNSSASN-----LIDQLFS--TNSSNPKIGIAASGGGFRAM 115
O59863 -----GFKGFTNDTS-----IIDKLLA--TQDT--APKVAIACSGGGYRAM 114
Q08108 -----ATANFSDSE-----VLSKLFNDGNSENLPKIAVACSGGGFRAM 117
Q03674 -----ATSNFSDTSLLS-----TLFSSNSSNPKIGIACSGGGYRAM 113
Q8TG06 -----AFANVS--TETS-----ALFN--DTENVPKLGLIAGVAGGGYRAM 103
P39457 -----VKVGDYDVGAULD-----KHSNGSSSLPNIGIAVSGGGWRAL 100
O42790 -----ANITGFDSSETFMN-----EAANNISQLPNVAIAISGGGYRAL 133
Q9P8P2 LETPP-----RTPNIGVALSGGGYRAM 110
Q9P8L1 LETPP-----RTPVIGVALAGGGYRAM 110
1CJY DLRFSMALDQEKTFRQORKEHIRESMKLLGPKNSEGLHSARDVPVVAILGSGGGFRAM 202

::: :*** ::

Q9UTH5 LFGGALAAALDSR--SNETTLG---GLLQSAHYITGADGGSWLLSSLAVNEFRTIQNISK 182
P78854 ILGSCALSAMDARHDNHTVLT---GLLQASDLVGTDCSAWTVGGIALNNFSTINDFSKL 183
O13857 VHGGVNLNADFDRNCGSSLA---GLLQSAHYIAGLSCGSWLVGSAVANNFANIYLRDN 165
Q9P327 VNGGAFNADFDRFESDPLS---GLLQSAHYISGLSCGSWLVGSAVANNFNITNYLRDN 201
Q9Y7N6 LTGACAINAFDARNGTTSLG---GLLQSSMYLTGLSCGSWLVGSAVANNFANITFLHDD 194
Q9UWF6 LTGAGEISSLDSRT--KTNTPV--LAGIQAASSYIAGLSCGSWLVGSLASNNLNSVDDMLSQ 158
O93795 LAGACELLALDSR--ATNPSV--LSEILQSSSYIVGLSCGSWLVGSLASNDLIPVQQLRE 156
Q9UVX1 LAGAQILGLDGRYEDANKHG--LGLLQSSSTEVVGLSCGNLVGSLALNDWLSVGDIVNG 248
Q11121 LSGACMLAAMDNRDTGANEHG--LGLLQOSTYLAGLSCGNLVGTLAWNNWTSVQDIVNN 168
P39105 LSGACMLAAMDNRDTGANEHG--LGLLQOGATYLAGLSCGNWLTSTLAWNNWTSVQAIVDN 170
Q8TG07 LSGACMVSAMDNRDTGANEHG--LGLLQOATYLAGLSCGNWLTSTLAWNNWTSVQDIVDS 174
O59863 LSGACMISAMDNRDTGANEHG--LGLLQOSTYLAGLSCGNLVGTLAYNNWTSVQAIINN 173
Q08108 LTGACVLAAMDNRTEGAYEHG--LGLLQOSTYLAGLSCGNLVGTLALNNWTSVQDILNN 176
Q03674 LGGACMIAAMDNRDTGANEHG--LGLLQOSTYLAGLSCGNWLTGTLAWNNWTSVQEIVDH 172
Q8TG06 FVGAQAFAMDNRDTGANEHG--LGLLQOATYLAGLSCGNWLTGTLAYNNWTSVQIILE 162
P39457 MNGACAVKAFDSRTDNATATGHLGGLLQSATYISGLSCGSWLLGSIYINNFTTVDKLOTH 160
O42790 MNGACFVAAADNRITQNTTGAAGGIGLQSSYLAGLSCGSWLVGSLFSSNFFSITFLSE 193
Q9P8P2 LTGLGGIMGMNNESTEASESE--TGSWLDGVSYWAAGLSCGSWATGTFMSNGGQLPTNLEN 169
Q9P8L1 LTGLGGIMGMNNESTEASQSE--TGSWLDGVSYWSGLSCGSWATGTFMSNGGQLPTNLEN 169
1CJY VGFSVMKALYES-----SILDQATYVAGLSCGSTWYMSTLYSHDPFPEKGPPEI 251

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Q9UTH5 IWY-----TRLGIFFIEETHFGDLKNYTNNVDEVNQAAAGFNVSLTDYWGRI 232
P78854 WAF-----NHPLMPKSAIVFN--AHFYSSIMNEVAE--ANAGFNISLSDYWGRI 231
O13857 VWN-----LEHSVFAPHGDNVVENLAYDDLDEIDQKADAGFDTSLTDLWGRAL 215
Q9P327 VWN-----LEHSVFAPHGDNVVENLAYDDLDEIDQKADAGFDTSLTDLWGRAL 251
Q9Y7N6 VWN-----LDHSLFAPY--DDAFENFYIQEWFQVLOKKNAGFNVSITDLWGRAL 243
Q9UWF6 G-----LWELTHSFLS---YYGIEHPKQVEEWNVGNQVAS--RNANFNVSITDIYGRLL 210
O93795 DK----LWDIQNSLVA---YGVN--IVRNTAMWGNINLQVQTLQAGFTVSITDVYGRAL 208
Q9UVX1 KST---IWQLQDSILN---PSGMR--IDKTIAYYYGLAQAVQAE--EDAGFQTSVTDWGRAL 301
Q11121 MTEDDSIWDISNSIIN---PGGFM--IVTTIKRWDHISDAVEGQDAGFNVSITDLWGRAL 224
P39105 TTESNSIWDISHSILT---PDGIN--IFKTGSRWDDISDDVQDQKAGFNISLADVWGRAL 226
Q8TG07 QDNDSAIWDISHSIVS---PGGIN--IFKTGSRWDDISDAVEEQKAGFNVSILADVWGRAL 230
O59863 MTDDNSIWDISNSIIVN---PGGIN--IFSSISRWDDISDAVEEQKAGFNISITDVWGRAL 229
Q08108 MQNDDSIWDLSDSIVT---PGGIN--IFKTAKRWDHISNAVESQADYNTSLADVWGRAL 232
Q03674 MSESDSIWNITKSIVN---PGGSN--LTYTIERWESIVQEVQAE--SDAGFNISLSDLWARAL 228
Q8TG06 GDKADAIWNITNSFLN---PYDKD--FSKTLARWTAIGSQVQGRDAGFNVTITDLWSRAL 218
P39457 EAG--SVWQFGNSIEGPDAGGIQ--LLDSAGYYKDLADAVDGEKAGFDTTLTDIWGRAL 217
O42790 NK----VWDFENSIKGPKEAGLS--VNRRIQYWSEVAKEVAKKADAGFETSITDYWGRAL 248
Q9P8P2 LWN-----IDSNLVFPDDDKLSFYTELYTETNAESDLGFPPIQITDVWGLAI 215
Q9P8L1 LWN-----IDSNLVFPDDDKLSFYTNLYTETNAESDLGFPVQITDIWGLAI 215

1CJY NEELMKN-----VSHNPLLLLLTPQVKRYVESLWK[KSSGQPVTFDDIFGMLI 299
 * : : * : :
 Q9UTH5 ARHFVQGLRG---GPNLTYSVVQ-NASWFQTAEYYPYPLIVTQGLTGGLPDGSGNGTATNSS 288
 P78854 SRTLGDTTYG---FPNVLSLSSIT-SQEWYRNANF[EPIITFATQN----YGEDISNVNTT 283
 O13857 SRKLV DATQG---GPNITFSSIR-NQWTFQADY[EPIIISDSRLE----EKAIPANTS 267
 Q9P327 SRKLVDAERG---GPGITYSSMR-NQSWFQADY[EPIIIVADSRL----EETAI PANTS 303
 Q9Y7N6 ALKLVNPLTG---GANTTFSSVT-NETWFQDGE[EPII IADNVIE----GETVIPLNDT 295
 Q9UWF6 SYPLLTNTED--EGDAYLWSDVT-SASNFSHQME[EPIIISDGRAP----DTTIINLNT 263
 O93795 SHQLLTNFDN--QGASFLWSDVT-ETTSFQNNEM[EPII IALGREG----NTVLMNFNST 261
 Q9UVX1 SYQFFEDDSTGGANITWSSIR-NLSSFQDHSME[EPIIIVVANGRTP----GTI IINENST 356
 Q11121 SYNFFPSLYR--GGVAYTWSTLR-DVEVFQNGEM[EPIISVADGRYP----GTQIIDLNAT 277
 P39105 AYNFWPSLHR--GGVGYTWSTLR-SADVFKNGEM[EPIITVADGRYP----GTTVINLNAT 279
 Q8TG07 SYQFFPTLYR--GGVAYLWSDLR-ESDVFKNAEM[EPIISVADGRYP----GTAVIDLNST 283
 O59863 SYNFFPSLDE--GGVGYTWSTLR-DVDVFKNGEM[EPIISVAVGRYP----GTQVVNLNAT 282
 Q08108 AYNFFPSLNR--GGIGLTWSSIR-DFPVFQNAEM[EPIISVADGRYP----GTVINLNAT 285
 Q03674 SYNFFPSLPD--AGSALTWSSLR-SADVFKNGEM[EPIITVADGRYP----GTTVINLNAT 281
 Q8TG06 AYGWFFPLPN--AGAGLTWSSLR-DNEIFMNGEM[EPIISVADGRYP----GTTVINLNAT 271
 P39457 SYQMFNASNG---GLSYTWSSIA-DTPEFQDGDY[EPIIFVADGRNP----GELVIGSNST 269
 O42790 SYQLIGADMG---GPAYTFSSIA-QTDFQKAET[EPIILVADGRAP----GDTIISLNT 300
 Q9P8P2 GSHVLPERYQLSNTPNLTFSSLPSVVALGNASL[EPII IAADRRR--EAGELVIAENAT 274
 Q9P8L1 GSHVLPERYQLSNTPNLTFSSLPSVVALGNASL[EPII IAADRRR--EAGELVIAENAT 274
 1CJY GETLIHNRMN-----TTLSSLK---EKVNTAQ[EPII IADRRR--EAGELVIAENAT 345
 * * :

Q9UTH5 IYEISPYLTSFDN----NVRSYPTQYLG[TNYSNGTA-VDGK[VTVQFDNVGFLVGTSS 342
 P78854 FFEASPENVGTFDH----GINSFIPT EYLG[TTLNNGAS-SNGS[VINYDNFGFMGASS 337
 O13857 IFEFTPYEFGTWDN----GIKAFIPMEYV[THLKNQVP-PDHK[VIRNYDNAGFVMG TSA 321
 Q9P327 IFEFTAYEFGTWDN----GIKAFIPMEYV[THLDCGVP-PDKS[VHNYDNAGFVMG TSA 357
 Q9Y7N6 VFEFTPIEFGTWDT----GVESFIPMEYV[THLINGIP-LNES[VVRNFDNAGFLMGTSS 349
 Q9UWF6 VIELTPYEFGSWDP----SLNEFVDTRYL[GTKLDNGRP-TGK-[YNGFDNAGFFMGTS 316
 O93795 VFEELTPYEVGSWDP----SLRSFVDTKYI[TRLDGAP-VSKR[VVNGFDNAGFFMGTS 315
 Q9UVX1 IFEISPYELGSWDP----SLKFSNIQYL[GSSVNNGNPNNTDI[VNNFDNAGFVIMGTS 411
 Q11121 VFEFNPFEMGSWDP----TLNAFTDVKYL[GTKVSN[GEVNVKGO[VAGYDNTGFIMGTS 332
 P39105 LFEFNPFEMGSWDP----TLNAFTDVKYL[GTNVN[CKPVNKGQ[VAGYDNTGFITATSS 334
 Q8TG07 VFEYSPPFELGSWDP----SLSAFTDVQYL[GTKVSD[CKPAEEGK[VAGYDNTGFIMGTS 338
 O59863 VFEFNPFEMGSWDY----TLHTFTDVRVAGT[NVNT[CTPNVTGK[VAGFDNTGFVIMGTS 337
 Q08108 VFEFNPFEMGSWDP----SLNSFANVKYL[GTNVSN[VPLERK[VAGYDNTGFIMGTS 340
 Q03674 LFEFTPFEMGSWDP----SLNAFTDVKYL[GTNVN[CKPVNKGQ[VSGYDNAGFVIATSA 336
 Q8TG06 VFEFTPFELGSWDP----SLNAFSDIKYL[GTQVTD[CKPE-TER[INGFDDASFIMGTS 325
 P39457 VFEFNPFEMGSWDP----TIFGVPLEYL[GSKFEGGSLPSNES[VIRGFDNAGFVIMGTS 324
 O42790 NYFEFNPFETGSWDP----TVYGFAPTKYL[CANFSN[VIPSGGK[VVEGLDQAGFVMGTS 355
 Q9P8P2 VWEFTPYEFGSWAFGSQYKSPGAF[PIEYLG[TSVDDGSP--NGT[VWKGFQLSFVMG TSA 332
 Q9P8L1 VWEFTPYEFGSWAFGSQYKSPGAF[PIEYLG[TSVDDGSP--NGT[VWKGFQLSFVMG TSA 332
 1CJY VWFSPYEIGMAKYG-----TFMAPDLFG[SKFFMGTVVKKYEENPLHFLMVGSAFS 398
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Q9UTH5 TRYNEALID---VSLRQSR---MSRRLGFTLR-HMRINGSSVSFY-PNPYTDATDIAGNA 394
 P78854 TYFNKIMRN---FNDSSTK---NGRIIQYLKGNFSENGQQIISI-PNPFQGVESAN--- 387
 O13857 TLFNTFLE---WSQEVTS-NSTLYDIIHKVFEKLSQNDIAPY-PNPYQNF-TTTN-- 373
 Q9P327 TLFNSFLD---WNENVKK-NDTYDILHAILEDLSKHQDDIAPY-PNPYQNY-TTSN-- 409
 Q9Y7N6 NVFSGILPA---TNASLTASNNTFNNAVLSFLEMLAEDQLDVGLY-PNPYQYGNASN-- 403
 Q9UWF6 ALFNEAVLS---ITEAN-IPSFLLKIDIDLVDPIKSNIDVSAYNPNPFFKS---SG-- 367
 O93795 SLFNIVLQ---LNNMP-IPPFLKELISKFTLDPVEKLNIDIAQYNPNPFFKS---NN-- 366
 Q9UVX1 SLFNQILL---LDNYS-INSI I KMLEKVLTD-VSDEYDIAVYEPNPFPGA---DS-- 461
 Q11121 SLFNQFLLQ---INSTS-LPSFIKNLVTFGLDD-LSEDEDDIAIYAPNPFKDTSYIQD-- 385
 P39105 TLFNQFLLR---LNSTD-LPSFIANLATDFLED-LSDNSDDIAIYAPNPFKEANFLQK-- 387
 Q8TG07 TLFNQFLLR---INDTS-IPKFIRNLATHFLKD-LSEYDIAVYAPNPFRRADYVNN-- 391
 O59863 SLFNQFLLQ---LNTTD-LPSFLYNLLHGFLTD-ASDDYDDISIWAPNPFYEITNIPS-- 390
 Q08108 TLFNQFLLR---INSTH-LPSFITRLARHFLKD-LSQDFNDIAVYSPNPFKDTKFLDS-- 393
 Q03674 SLFNEFSLE---ASTST-YYKMINSFANKYVNN-LSQDDDDIAIYAANPFKDTFVDR-- 389
 Q8TG06 SLFNEFTMS---NDSAV-AYTYLNTLSSTLVKG-IDKENNDIAMYAPNPFKGSKYVDS-- 378
 P39457 SLFNQFLLQ---INTTS-LPSFIKDVFNGLFD-LDKSQNDIASYDPNPFYKYN----- 373
 O42790 TLFNQFLLAN---ISSYDGVDPVLEAVTSVLKE-IGAKRDDVSQIIPNPFLLDWN----- 406
 Q9P8P2 TLFNGAFLEL---NGTDS---GLLTNLI TAFLAD-LGEDQADISRI-PNTFSNYNSGEN-- 383
 Q9P8L1 TLFNGAFLEL---NGTDS---GLLTNLI TAFLAD-LGEDQADISRI-PNSFSNYNSGEN-- 383
 1CJY ILFNRVLGVSGSQRGSTMEELENITTKHIVS-NDSSDSDDESHEPKGTENEDAGSDYQ 457
 : : :

Q9UTH5 TAVSEDIVDTPYLDLFDG----GYDGQNIPIWPLLQPERKLDVVFAFDSSGDTSN-FWPN 449
 P78854 SDAANLGSSSSLNLVDT----FLTGEKIPLWPLLQGRDQVAVVAVDNGDDSEW-LWPN 442

013857 TTVKNPFERFDIDLVDG----GEDDENIPIWPLLHPQRFVDVIFAVDATYDDSN-GWPD 428
 Q9P327 TSVVNAFEPYDIDLVDG----GEDRENIPLWPLLHPQRFVDVIFAIDSTYNDPY-GWPL 464
 Q9Y7N6 TTTTNPLEPYPIIELIDG----GSDSEGIWFPLLHPQRFVDVIFAIDGGYQSATSGWPD 459
 Q9UWF6 -S-NTAISQSKNLYLVDG----GEDGQNIPI SPLLH--RNVSAIFAFDNS-NDVLN-WPD 417
 093795 -S-DTKIAQSRITLYLADG----GEDGQNVPLPLI--RKVSAIFAFDQS-ADKNN-WPD 416
 Q9UVX1 -AGIKSITNTDLYLVDG----GEDLQNVFPYPLIQNKRGVDFVIFAIDNS-ADTNSWPN 515
 Q11121 -NFSKSISES DYLYLVDG----GEDNQNIPLVPLVQDERNVDFVIFALDNS-ADTDYWPD 439
 P39105 -NATSSIESEYLFLVDG----GEDNQNIPLVPLLQKERELDVIFALDNS-ADTDDYWPD 441
 Q8TG07 -NRKSLSESEYLFLVDG----GEDGQNVPLVPLIQQERDLDFVIFALDNS-ADTEENWPD 445
 059863 -NYSQSISEDDTLYLVDG----GEDGQNIPLTPLLQTEREIDVIFALDNS-ADTDQSWPD 444
 Q08108 -DYTTSIVDSDSLFLVDG----GEDDENVPVPLI QKERVDVIFAIDNS-ADMRLAWPD 447
 Q03674 -NYTSSIVDADDLFLVDG----GEDGQNLPLVPLIKKERDLVVVFDLIS-DNTDESWPS 443
 Q8TG06 -NYTTSIVDSDSLFLVDG----GEDLQGI PFVPLLQERDLDFVIFAIDV-DTSSDNYPA 432
 P39457 -EHSSPYAAQKLLDVVDG----GEDGQNVPLHPLIQPERHVDVIFAIDSS-ADTDYFWPN 427
 042790 -NRTNPNADTLELDLVDG----GEDLQNIPLNPLTQPVRVDVIFAIDSS-ADVTN-WPN 459
 Q9P8P2 -----PIYNLYITLVDA----GETNQNIPLVPLVPTRDVDAIFAIDSS-YDTDYIWPN 433
 Q9P8L1 -----PIYNLYITLVDA----GETNQNIPLVPLVPTRDVDAIFAIDSS-YDSDYIWPN 433
 1CJY SDNQASWIHRMIMALVSDSALFNTREGRAGKVHNFMLGGLNLTNSYPLSPLSDFATQDSFD 517

: :

Q9UTH5 GSSLVATYERVTQRASDAVYDVEDFVHVPTPETFVNGLNANPTFFG DGRNTRRGD--- 506
 P78854 GNSLVQTYERVVAAQAAGNTNVKGFYVPSQQSFVSLHFNDRPVFFG DGRNTRTAGN--- 499
 013857 GSSIVTTYERIIITYNANKSVDVRGFFYIPDEDTIISLGLNTHPTFFG DGRNTRTAGN--- 485
 Q9P327 GSSIVATYERVVTFNANKSVDVRGFFYIPDENTIISLGLNTRPTFFG DGKNTTAGN--- 521
 Q9Y7N6 GSSLVSTYERVIATNSG---VRGFFYIPDNTFLALGLNTHPTFFG DGRNTRTAGN--- 513
 Q9UWF6 GTSLVKTYER--QFSS--QGNGIAFFYPVDPQYTFRNLNLSKPTFFG DAKNLTSLT--- 470
 093795 GSALIKTFER--QFSS--QGNGIAFFYPVDPQNTFRNTNLSKPTFFG DAKNLTSLT--- 469
 Q9UVX1 GTSIQETYKR--QFSK--QGKGTFFFAPDYKTFLDKNMGDKPVFFG NNSDLEDLVAVH 571
 Q11121 GASLVSTYER--QFSS--QGLNMSFFYPVDPKRTFVNLGLADKPSFFG DANLTDLN--- 492
 P39105 GASLVNTYQR--QFGS--QGLNLSFFYPVDPVNTFVNLGLNKKPTFFG DARNLTDLE--- 494
 Q8TG07 GASLMHTYR--QFGF--QGQGVTFPSVPGTDTFVNLGLNKKPTFFG DARNMTDLE--- 498
 059863 GFSLTQTYAR--QFGL--QGKIAFFYPVDPVNTFVNLGLNTRPTFFG DARNLTDLE--- 497
 Q08108 GSSLVHTYER--QFVK--QGQMSFFYPVDPNTFVNLGLNKKPTFFG DANLTDLQ--- 500
 Q03674 GVCMTNTYER--QYSK--QGKGMFFYPVDPVNTFVNLGLNKKPTFFG DAKNLTDLQ--- 496
 Q8TG06 GGPMMKTYER--QFSK--QGKGMFFYPVDPMTTFVNLGLGKPSFYG DANLTDLE--- 485
 P39457 GTSLVATYER--SLNSSGIANGTAFPAVDQNTFINLGLSTRPSFFG DSSNQTGPS--- 482
 042790 GTALRATYER--TFGS--ISNGTLFPSIPDDWTFINLGLNRRPSFFG DVKNFTLNAN--- 513
 Q9P8P2 GTALRTTYER--AKVLAEHENTRVLMPFVPSMNGFVNGGYSRPTFFG NDTT----- 484
 Q9P8L1 GTALRTTYER--AKVLAEHENTRVLMPFVPSMNGFVNGGYSRPTFFG NDTT----- 484
 1CJY DDELDAAVADPDEFERIYEPLDVKSKKIHVVDLSGLTFNLPYPLILRQGVDLIISFDFS 577

. : :

Q9UTH5 --VPVDHNTPELLVYMPNTPWTMKNLVDHRYRIANSEIQALIQN-GFVATTQDN---ST 560
 P78854 --HTVTRDTPPELLVIYLPNVPYNYFTNISTDRTYTEDMIQQLLTN-GLISSVTDN---DT 553
 013857 --HTVDNNTPELLVYFPNYPWVYFNISTFTMSMNDTLSSGILEN-AALSATQNN---SD 539
 Q9P327 --HDVDNNTPELLVYFPNYPWYYSNISTFTMSMDDKMANGILEN-AFMSTQNN---NE 575
 Q9Y7N6 --HTVNDTPELLVYFPNYPWTMYANVTYTVQLEDTLSSGMIEN-AAVAATQNN---SD 567
 Q9UWF6 ----KDIYDPELLVIYLANRPFTYWSNTSTFKLTYDDNERQGMISN-GFEIATRSGSLDD 525
 093795 ----ENIYDPELLVIYLANRPFTYWSNTSTFKLKYSDTERQGMISN-GYDVASRLNGLDN 524
 Q9UVX1 ENDKINVTPELLVYVTSNTRMSYNSNFSTFKLSYSDQEKFGAIRN-GFETVTRNNTLTDDE 630
 Q11121 -----YIPELLVYIPNARHSYNSNTSTFKLSYTDDERLKMIKN-GFEAATRGNLTDSD 544
 P39105 -----YIPELLVYIPNARHSYNSNTSTFKLSYSDSERLGMIKN-GFEAATMGNFTDSD 546
 Q8TG07 -----YIPELLVYIPNARHSYNSNTSTFKLSYSEKERLGVIRN-GFEAATMNNLTADS 550
 059863 -----SIPPELLVYMPNTRSFNSNTSTFKMSYSTSERFKMIQN-GFEAVTMKNLTKDE 549
 Q08108 -----YIPELLVYLPNAEYSFNSNQSAFKLSYSESQRSMIQN-GFEIATRNNFTDDP 552
 Q03674 -----YIPELLVYIPNTHKSFNGNQSTLKMNYNVTERRLGMIRN-GFEAATMGNFTDSD 548
 Q8TG06 -----YIPELLVYIPNTHKSFNSYHSFESNVSTFKLNYSERVGMIRN-AFEATRNNLTEDA 537
 P39457 -----PLVYIPNAPYSYHSNISTFQLSTDDAERDNIILN-GYEVATMANSTLDD 531
 042790 -----QKVPPELLVYVPNAPYPTALSNVSTFDPSYMSQRNDIIGN-GWNSATQNGTLDS 566
 Q9P8P2 -----TELIIVVPSYPSFAANTSTYQLSYENDEANEMLLN-GMRSLTLNHS--VP 532
 Q9P8L1 -----TEVIIYIPSYPSYPSFAANTSTYQLSYENNEANEMLLN-GMRSLTLNHS--VP 532
 1CJY --ARPSDSSPELLKELLAEKWKAMNKLFPFKIDPYVFDREGLKE YVFKPNPDMEKD P 635

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Q9UTH5 DFASLAVVQPSLERRNQSTSAACQCFQSYWNGTVDN----- 601
 P78854 YFGQFAVAVVKTLEARNNITASPEQQCFYNYWWSGLYDD----- 594
 013857 SFAVQLAVVAVVQPSLERRNMTSPQCSSEFEQYVWNG----- 576
 Q9P327 SFAVQLAVVAVVQPSLERRKLTPTQCSSEFEQYVWNG----- 612
 Q9Y7N6 SFAVQLAVVAVVQPSLERRNMTSPQCSSEFEQYVWNG----- 604
 Q9UWF6 EWAAVGVAVVAVVQPSLERRKLTPTQCSSEFEQYVWNG-----TIYKGEPLG----- 571
 093795 EWAAVGVAVVAVVQPSLERRKLTPTQCSSEFEQYVWNG-----TIYKGEPLG----- 570

Q9UVX1 NWSTVVGCAIIRPQERLGEEQSDEGKKCFQEYCWTTGGFKDAASVSSVSGISGLA----- 685
 Q11121 SFMGEVACAVMRKQQLNATLPPEEETCFNYCWNG-TIDDPVSGLDNSDFDP----- 598
 P39105 DFLGVGCAIIRPKQQLNATLPPEEESQCFNYCWNG-TIDSRVSGVGNDDYSS----- 600
 Q8TG07 NFAGVIGCAIMRRKQQLNATLPKEEETCFNYCWNG-TIDNTPAKGVTASNDFD----- 604
 O59863 NFMGIVSCAILRQKESLNATLPPEEEDAFKEYCWNG-TVDAT-----TPISS----- 596
 Q08108 EFMGVGCAIIRPKQALNATLPPEEETCFKNYCWNG-TLDTTLPDVEKDVHHSFINVN 611
 Q03674 NFLGVIGCAIIRPKQESLNATLPPEEETCFADYCWNG-TLSTSANPELSGNSTYQSGAIA 607
 Q8TG06 DYVTVVGCAIIRPKQQLNATLPDIEGDKCFNYCWNG-TIDNTPPKLLTPN-NQDPAAIS 595
 P39457 NWTAVACAVISRFERTGTLPDIEGDKCFNYCWNG----- 568
 O42790 EWPTVACAVISRFSLDRLGRQTPAAKTCFERYCWNG----- 603
 Q9P8P2 TWPTVACALTDPSFMYTSENRRSTTCQCFDTCWAG----- 569
 Q9P8L1 TWPTVACALTDPSFMYTSENRRSTTCQCFDTCWAG----- 569
 1CJY TIIHFVLANINFRKYKAPGVPRETEEEKEIADDFIDDPESPFSTFN----- 682

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Q9UTH5 -----TPVDDDSKNPTYNPAVKTSSASGVHA 627
 P78854 -----SAANDD---IVYNPTCRLG--EGI-- 613
 O13857 -----TTVNNPSAVSNYAPT VLSASTS--- 599
 Q9P327 -----TLAT--STASVYDPTVMSAATTSRAP 636
 Q9Y7N6 -----TIAS--TTVTTYAPT VLSAKIYK-- 625
 Q9UWF6 -----ENFSDDGLTNSATEYNSNNVAGFND 596
 O93795 -----DNFSDEGLTTSAAAYNSNNVAGIND 595
 Q9UVX1 -----AKHTSGGTSSTTQQTSTTTGSSAN 710
 Q11121 -----TAASSAYSANTESYSSSS----- 617
 P39105 -----SASLSASAAAASASASASASASAS 624
 Q8TG07 -----NASGSAAADMAEQDASGAAS----- 624
 O59863 -----TTSSSASSTSTSDSGN----- 612
 Q08108 SFNSSIGQEE-----SLYAGSSASQSSSSSSSSSSSEIPS 647
 Q03674 SAISEATDGIPITALLGSSTSGNTTSTSTSSNVTSNSSSNTTLNSNSSS--SSISS 666
 Q8TG06 SAIAAVTDDSPIGALLNTGSG--TKSNSSSKTNSTLVTSSRATSTGTLSNSSSNTVSS 653
 P39457 -----TVNSTRPESYDPAFYLADN---SM 589
 O42790 -----TVNSKDTGVYMEPKIADAHALDS 627
 Q9P8P2 -----DDNTEPATYEPVINSVPPWLIVAN 593
 Q9P8L1 -----DDNTEPANYPVINSVPPWLIAN 593
 1CJY -----FQYPNQAFKRLHDLMHFNTLNNIDV 707

Q9UTH5 NILLS-----FFVLLATLLVTA----- 644
 P78854 -----
 O13857 -GTSS-----VRAKPIVFYLFASLLTVSLLL----- 624
 Q9P327 SGTTSGTASSTSSSVASATPTHKHWDSIFEAKENP----- 673
 Q9Y7N6 -----SRLFTYCS----- 633
 Q9UWF6 GGTSILKKA----- 605
 O93795 GGIALVKRDDLSN----- 608
 Q9UVX1 GGSSSTGSSSSSKKNGGDLVNGGVPSSIFLVFN--SLLGLIIAYL 754
 Q11121 ATGS--KKNAG-----LPATPTSFTSILTLT--AIAGFL---- 649
 P39105 ASGSSTHKKNAGNALVNYNSLNTNTFTIGVLSVIS--AVFGLI---- 664
 Q8TG07 ASSSRKKNAAVS-----VDVNAKTLFAIITAMT--AVFQLI---- 659
 O59863 -----KENSAR-----ILAPRSTLSLLIGGLA--SVFISF---- 640
 Q08108 ATATLEKKAATNSG--SHLSGISVKFSAMIMLTL--LMFTGAV--- 686
 Q03674 STARSSSTANKANAAAISYANTNTLMSLLGAIT--ALFGLI---- 706
 Q8TG06 TAARSSTSTAKKNAGSVLKLEFSKASVMVAIAAAAVASLI---- 695
 P39457 ASVSLPTML-----STVVAAGLAMLILV----- 612
 O42790 GAVAI GKMVNVW-----SSVVVGVAATLLL----- 653
 Q9P8P2 NLSIGVADAPASNESTAGTASSGAANADVSMGMVALAAGLGLML-- 637
 Q9P8L1 NLSIGMADAPGSNESTAGTASSGAAKMGVGMGMVALTAGLGLML-- 637
 1CJY IKEAMVESIEYRRQNPSCRCSVLSNVEARRFFNKEFLSKPKA---- 749

| Appendix 3. | | | | |
|-------------|-----------|--------------------------|---|---|
| Sub-family | AC Number | Organism | Substrate Specificity | Reference |
| AI | Q9P4E5 | Candida albicans (Yeast) | olive oil [(carbon source) TAG, Tween 40], α -naphthyl palmitate | "Secreted lipases of Candida albicans: cloning, characterisation and expression analysis of a new gene family with at least ten members.", <i>Arch. Microbiol.</i> 174 :362-374(2000). |
| AI | Q9P8W5 | Candida albicans (Yeast) | olive oil [(carbon source) TAG, Tween 40], α -naphthyl palmitate | "Secreted lipases of Candida albicans: cloning, characterisation and expression analysis of a new gene family with at least ten members.", <i>Arch. Microbiol.</i> 174 :362-374(2000). |
| AI | Q9P4E8 | Candida albicans (Yeast) | olive oil [(carbon source) TAG, Tween 40], α -naphthyl palmitate | "Secreted lipases of Candida albicans: cloning, characterisation and expression analysis of a new gene family with at least ten members.", <i>Arch. Microbiol.</i> 174 :362-374(2000). |
| AI | O94091 | Candida albicans (Yeast) | olive oil [(carbon source) TAG, Tween 40], α -naphthyl palmitate | "Secreted lipases of Candida albicans: cloning, characterisation and expression analysis of a new gene family with at least ten members.", <i>Arch. Microbiol.</i> 174 :362-374(2000). |
| AI | Q9P8W2 | Candida albicans (Yeast) | olive oil [(carbon source) TAG, Tween 40], α -naphthyl palmitate | "Secreted lipases of Candida albicans: cloning, characterisation and expression analysis of a new gene family with at least ten members.", <i>Arch. Microbiol.</i> 174 :362-374(2000). |
| All | Q9P8W0 | Candida albicans (Yeast) | olive oil [(carbon source) TAG, Tween 40], α -naphthyl palmitate | "Secreted lipases of Candida albicans: cloning, characterisation and expression analysis of a new gene family with at least ten members.", <i>Arch. Microbiol.</i> 174 :362-374(2000). |

| Sub-family | AC Number | Organism | Substrate Specificity | Reference |
|------------|-----------|--------------------------------------|--|---|
| All | Q9P8V9 | Candida albicans (Yeast) | olive oil [(carbon source) TAG, Tween 40], α -naphthyl palmitate | "Secreted lipases of Candida albicans: cloning, characterisation and expression analysis of a new gene family with at least ten members.", <i>Arch. Microbiol.</i> 174 :362-374(2000). |
| All | Q9P8W1 | Candida albicans (Yeast) | olive oil [(carbon source) TAG, Tween 40], α -naphthyl palmitate | "Secreted lipases of Candida albicans: cloning, characterisation and expression analysis of a new gene family with at least ten members.", <i>Arch. Microbiol.</i> 174 :362-374(2000). |
| All | Q9P4E6 | Candida albicans (Yeast) | olive oil [(carbon source) TAG, Tween 40], α -naphthyl palmitate | "Secreted lipases of Candida albicans: cloning, characterisation and expression analysis of a new gene family with at least ten members.", <i>Arch. Microbiol.</i> 174 :362-374(2000). |
| A | Q9P4E7 | Candida albicans (Yeast) | olive oil [(carbon source) TAG, Tween 40], α -naphthyl palmitate | "Secreted lipases of Candida albicans: cloning, characterisation and expression analysis of a new gene family with at least ten members.", <i>Arch. Microbiol.</i> 174 :362-374(2000). |
| BI | P20261 | Candida rugosa (Candida cylindracea) | 4-nitrophenyl caprylate, 4-nitrophenyl laurate, sulcatol, tributyrin, triacetin, 2,4,6-trinitrobenzene sulfonic acid | Influence of the conformational flexibility on the kinetics and dimerisation process of two Candida rugosa lipase isoenzymes. Pemas, M.A.; Lopez, C.; Rua, M.L.; Hermoso, J.; <i>FEBS Lett.</i> 501 , 87-91 (2001) |
| BI | P32947 | Candida rugosa (Candida cylindracea) | 4-nitrophenyl caprylate, 4-nitrophenyl laurate, sulcatol, tributyrin, triacetin, 2,4,6-trinitrobenzene sulfonic acid | Influence of the conformational flexibility on the kinetics and dimerisation process of two Candida rugosa lipase isoenzymes. Pemas, M.A.; Lopez, C.; Rua, M.L.; Hermoso, J.; <i>FEBS Lett.</i> 501 , 87-91 (2001) |

| Sub-family | AC Number | Organism | Substrate Specificity | Reference |
|------------|-----------|---|---|--|
| BI | P32949 | Candida rugosa (Candida cylindracea) | 4-nitrophenyl caprylate, 4-nitrophenyl laurate, sulcatol, tributyrin, triacetin, 2,4,6-trinitrobenzene sulfonic acid | Influence of the conformational flexibility on the kinetics and dimerisation process of two Candida rugosa lipase isoenzymes. Pernas, M.A.; Lopez, C.; Rua, M.L.; Hermoso, J.; <i>FEBS Lett.</i> 501 , 87-91 (2001) |
| BI | P32946 | Candida rugosa (Candida cylindracea) | 4-nitrophenyl caprylate, 4-nitrophenyl laurate, sulcatol, tributyrin, triacetin, 2,4,6-trinitrobenzene sulfonic acid | Influence of the conformational flexibility on the kinetics and dimerisation process of two Candida rugosa lipase isoenzymes. Pernas, M.A.; Lopez, C.; Rua, M.L.; Hermoso, J.; <i>FEBS Lett.</i> 501 , 87-91 (2001) |
| BI | P32948 | Candida rugosa (Candida cylindracea) | 4-nitrophenyl caprylate, 4-nitrophenyl laurate, sulcatol, tributyrin, triacetin, 2,4,6-trinitrobenzene sulfonic acid | Influence of the conformational flexibility on the kinetics and dimerisation process of two Candida rugosa lipase isoenzymes. Pernas, M.A.; Lopez, C.; Rua, M.L.; Hermoso, J.; <i>FEBS Lett.</i> 501 , 87-91 (2001) |
| BII | P22394 | Geotrichum candidum (Oospora lactis) | Several simple triglycerides. Long chain fatty acid with cis-9 unsaturated bonds. CaprylinC8, CaprinC10, LaurinC12, TrioleinC18:1, ElaidinC18:1, LinoleinC18:2(opt), LinoleninC18:3 | Influence of the conformational flexibility on the kinetics and dimerisation process of two Candida rugosa lipase isoenzymes. Pernas, M.A.; Lopez, C.; Rua, M.L.; Hermoso, J.; <i>FEBS Lett.</i> 501 , 87-91 (2001) |
| BII | P79066 | Geotrichum fermentans (Trichosporon fermentans) | Several simple triglycerides. Long chain fatty acid with cis-9 unsaturated bonds. CaprylinC8, CaprinC10, LaurinC12, TrioleinC18:1, ElaidinC18:1, LinoleinC18:2(opt), LinoleninC18:3 | Influence of the conformational flexibility on the kinetics and dimerisation process of two Candida rugosa lipase isoenzymes. Pernas, M.A.; Lopez, C.; Rua, M.L.; Hermoso, J.; <i>FEBS Lett.</i> 501 , 87-91 (2001) |
| BII | P17573 | Geotrichum candidum (Oospora lactis) | With unsaturated long fatty acyl chain: triolein, trivaccinin, trilinolein, trilinolenin With saturated fatty acyl chain: tributyrin c4, trihexanoin C6, trioctanoin C8, tridecanin C10, trilaurin C12, trimyristin C14, tripalmitin C16, tristearin C18. | Influence of the conformational flexibility on the kinetics and dimerisation process of two Candida rugosa lipase isoenzymes. Pernas, M.A.; Lopez, C.; Rua, M.L.; Hermoso, J.; <i>FEBS Lett.</i> 501 , 87-91 (2001) |

| Sub-family | AC Number | Organism | Substrate Specificity | Reference |
|------------|-----------|---|---|---|
| BIII | Q99156 | <i>Yarrowia lipolytica</i> (<i>Candida lipolytica</i>) | Several simple triglycerides. Long chain fatty acid with cis-9 unsaturated bonds. CaprylinC8, CaprinC10, LaurinC12, TrioleinC18:1, ElaidinC18:1, LinoleinC18:2(opt), LinoleninC18:3 | Influence of the conformational flexibility on the kinetics and dimerisation process of two <i>Candida rugosa</i> lipase isoenzymes. Pernas, M.A.; Lopez, C.; Rua, M.L.; Hermoso, J.; <i>FEBS Lett.</i> 501 , 87-91 (2001) |
| BIII | Q96VC9 | <i>Yarrowia lipolytica</i> (<i>Candida lipolytica</i>) | Several simple triglycerides. Long chain fatty acid with cis-9 unsaturated bonds. CaprylinC8, CaprinC10, LaurinC12, TrioleinC18:1, ElaidinC18:1, LinoleinC18:2(opt), LinoleninC18:3 | Influence of the conformational flexibility on the kinetics and dimerisation process of two <i>Candida rugosa</i> lipase isoenzymes. Pernas, M.A.; Lopez, C.; Rua, M.L.; Hermoso, J.; <i>FEBS Lett.</i> 501 , 87-91 (2001) |
| CI | P61871 | <i>Rhizopus niveus</i> | Tributyryne, 1,2-didecanoyl-rac-glycerol | Can lipases hydrolyze a peptide bond? Maruyama, T.; Nakajima, M.; Kondo, H.; Kawasaki, K.; Seki, M.; Goto, M.; <i>Enzyme Microb. Technol.</i> 32 , 655-657 (2003) |
| CI | P61872 | <i>Rhizopus oryzae</i> (<i>Rhizopus delemar</i>) | Tributyryne, 1,2-didecanoyl-rac-glycerol | Can lipases hydrolyze a peptide bond? Maruyama, T.; Nakajima, M.; Kondo, H.; Kawasaki, K.; Seki, M.; Goto, M.; <i>Enzyme Microb. Technol.</i> 32 , 655-657 (2003) |
| CI | P19515 | <i>Rhizomucor miehei</i> | tributyryne, 4-nitrophenyl decanoate, 1,2-O-dilauryl-rac-glycero-3-glutaric acid resorufin ester, tridecanin, trihexanin, trioctanin | Properties of recombinant <i>Rhizomucor miehei</i> lipase with amino acid substitutions of Phe94 in the substrate binding domain, Oh, S.-W.; Gaskin, D.J.H.; Kwon, D.Y.; Vulfson, E.N.; <i>Biotechnol. Lett.</i> 23 , 563-568 (2001) |
| CII | P61870 | <i>Penicillium camembertii</i> | tributyryne, 4-nitrophenyl decanoate, 1,2-O-dilauryl-rac-glycero-3-glutaric acid resorufin ester, tridecanin, trihexanin, trioctanin | Properties of recombinant <i>Rhizomucor miehei</i> lipase with amino acid substitutions of Phe94 in the substrate binding domain, Oh, S.-W.; Gaskin, D.J.H.; Kwon, D.Y.; Vulfson, E.N.; <i>Biotechnol. Lett.</i> 23 , 563-568 (2001) |

| Sub-family | AC Number | Organism | Substrate Specificity | Reference |
|------------|-----------|--|---|---|
| CII | P61869 | <i>Penicillium cyclopium</i> | tributyryn, trioctanoin, trihexanin, methyl linoleate, methyl oleate, methyl ricinoleate, polyethylene sorbitan monooleate, trilaurin, trioleoylglycerol, tripalmitin | Biochemical and structural characterization of triacylglycerol lipase from <i>Penicillium cyclopium</i> , Ibrik, A.; Chahinian, H.; Rugani, N.; Sarda, L.; Comeau, L.C.; <i>Lipids</i> 33 , 377-384 (1998) |
| CII | O59952 | <i>Thermomyces lanuginosus</i> (<i>Humicola lanuginosa</i>) | tridodecanoin, tributyrin, 4-nitrophenyl caprylate, 1,2-dioleoylglycerol, methyl linoleate, methyl oleate, methyl oleate, trioleoylglycerol | Production, purification, characterization, and applications of lipases, Sharma, R.; Chisti, Y.; Banerjee, U.C.; <i>Biotechnol. Adv.</i> 19 , 627-662 (2001) |
| DI | Q9UWF6 | <i>Candida albicans</i> | lysophosphatidylcholine, 1-palmitoyl-sn-glycero-3-phosphocholine | Purification and characterization of lysophospholipase-transacylase of pathogenic fungus <i>Candida albicans</i> , Takahashi, M.; Banno, Y.; Shikano, Y.; Mori, S.; Nozawa, Y. <i>Biochim. Biophys. Acta</i> 1082 , 161-169 (1991) |
| DI | O93795 | <i>Candida albicans</i> | lysophosphatidylcholine, 1-palmitoyl-sn-glycero-3-phosphocholine | Purification and characterization of lysophospholipase-transacylase of pathogenic fungus <i>Candida albicans</i> , Takahashi, M.; Banno, Y.; Shikano, Y.; Mori, S.; Nozawa, Y. <i>Biochim. Biophys. Acta</i> 1082 , 161-169 (1991) |
| DI | Q9UVX1 | <i>Candida albicans</i> | lysophosphatidylcholine, 1-palmitoyl-sn-glycero-3-phosphocholine | Purification and characterization of lysophospholipase-transacylase of pathogenic fungus <i>Candida albicans</i> , Takahashi, M.; Banno, Y.; Shikano, Y.; Mori, S.; Nozawa, Y. <i>Biochim. Biophys. Acta</i> 1082 , 161-169 (1991) |
| DII | Q11121 | <i>Torulasporea delbrueckii</i> (<i>Saccharomyces rosei</i>) | lysophosphatidylcholine, 1-palmitoyl-sn-glycero-3-phosphocholine | Purification and characterization of lysophospholipase-transacylase of pathogenic fungus <i>Candida albicans</i> , Takahashi, M.; Banno, Y.; Shikano, Y.; Mori, S.; Nozawa, Y. <i>Biochim. Biophys. Acta</i> 1082 , 161-169 (1991) |

| Sub-family | AC Number | Organism | Substrate Specificity | Reference |
|------------|-----------|--|--|---|
| DII | P39105 | <i>Saccharomyces cerevisiae</i> | lysophosphatidylcholine, phosphatidylinositol-4,5-bisphosphate, phosphatidylcholine, phosphatidylethanolamine, phosphatidylserine PtdSer>PtdIns>>PtdCho.PtdEtn | Purification and characterization of lysophospholipase-transacylase of pathogenic fungus <i>Candida albicans</i> , Takahashi, M.; Banno, Y.; Shikano, Y.; Mori, S.; Nozawa, Y. <i>Biochim. Biophys. Acta</i> 1082 , 161-169 (1991) |
| DII | Q8TG07 | <i>Candida glabrata</i> (<i>Torulopsis glabrata</i>) | lysophosphatidylcholine, 1-palmitoyl-sn-glycero-3-phosphocholine | Purification and characterization of lysophospholipase-transacylase of pathogenic fungus <i>Candida albicans</i> , Takahashi, M.; Banno, Y.; Shikano, Y.; Mori, S.; Nozawa, Y. <i>Biochim. Biophys. Acta</i> 1082 , 161-169 (1991) |
| DII | O59863 | <i>Kluyveromyces lactis</i> (Yeast) | lysophosphatidylcholine, phosphatidylcholine | Purification and characterization of lysophospholipase-transacylase of pathogenic fungus <i>Candida albicans</i> , Takahashi, M.; Banno, Y.; Shikano, Y.; Mori, S.; Nozawa, Y. <i>Biochim. Biophys. Acta</i> 1082 , 161-169 (1991) |
| DII | Q08108 | <i>Saccharomyces cerevisiae</i> | 1-acyl-sn-glycero-3-phosphocholin, 2-acylglycerophosphocholine, phosphatidylinositol, phosphatidylserine, phosphatidylinositol-4,5-bisphosphate, Missing the other 2 from P39105 and Q03674(ptdCho,PtdEtn) | Purification and some properties of soluble phospholipase B from baker's yeast (<i>Saccharomyces cerevisiae</i>), Ichimasa, M.; Shiobara, M.; <i>Agric. Biol. Chem.</i> 49 , 1083-1089 (1985) |
| DII | Q03674 | <i>Saccharomyces cerevisiae</i> | phosphatidylinositol-4,5-bisphosphate, phosphatidylcholine, phosphatidylethanolamine, phosphatidylserine PtdSer>PtdIns>>PtdCho.PtdEtn | Purification and some properties of soluble phospholipase B from baker's yeast (<i>Saccharomyces cerevisiae</i>), Ichimasa, M.; Shiobara, M.; <i>Agric. Biol. Chem.</i> 49 , 1083-1089 (1985) |
| DII | Q8TG06 | <i>Candida glabrata</i> (Yeast) (<i>Torulopsis glabrata</i>) | phosphatidylinositol-4,5-bisphosphate, phosphatidylcholine, phosphatidylethanolamine, phosphatidylserine PtdSer>PtdIns>>PtdCho.PtdEtn | Purification and some properties of soluble phospholipase B from baker's yeast (<i>Saccharomyces cerevisiae</i>), Ichimasa, M.; Shiobara, M.; <i>Agric. Biol. Chem.</i> 49 , 1083-1089 (1985) |

| Sub-family | AC Number | Organism | Substrate Specificity | Reference |
|------------|-----------|---|--|--|
| DIII | P39457 | <i>Penicillium chrysogenum</i> (<i>Penicillium notatum</i>) | phosphatidylinositol-4,5-bisphosphate, phosphatidylcholine, phosphatidylethanolamine, phosphatidylserine PtdSer>PtdIns>>PtdCho.PtdEtn | Purification and some properties of soluble phospholipase B from baker's yeast (<i>Saccharomyces cerevisiae</i>), Ichimasa, M.; Shiobara, M.; <i>Agric. Biol. Chem.</i> 49 , 1083-1089 (1985) |
| DIII | O42790 | <i>Neurospora crassa</i> | phosphatidylinositol-4,5-bisphosphate, phosphatidylcholine, phosphatidylethanolamine, phosphatidylserine PtdSer>PtdIns>>PtdCho.PtdEtn | Purification and some properties of soluble phospholipase B from baker's yeast (<i>Saccharomyces cerevisiae</i>), Ichimasa, M.; Shiobara, M.; <i>Agric. Biol. Chem.</i> 49 , 1083-1089 (1985) |
| DIV | O13857 | <i>Schizosaccharomyces pombe</i> | lysophosphatidylcholine, lysophosphatidylethanolamine, phosphatidylinositol, phosphatidylserine, lysophosphatidylethanolamine, phosphatidic acid | Purification and some properties of phospholipase B from <i>Schizosaccharomyces pombe</i> , Oishi, H.; Tsuda, S.; Watanabe, Y.; Tamai, Y. <i>Biosci. Biotechnol. Biochem.</i> 60 , 1087-1092 (1996) |
| DIV | Q9P327 | <i>Schizosaccharomyces pombe</i> | lysophosphatidylcholine, lysophosphatidylethanolamine, phosphatidylinositol, phosphatidylserine, lysophosphatidylethanolamine, phosphatidic acid | Purification and some properties of phospholipase B from <i>Schizosaccharomyces pombe</i> , Oishi, H.; Tsuda, S.; Watanabe, Y.; Tamai, Y. <i>Biosci. Biotechnol. Biochem.</i> 60 , 1087-1092 (1996) |
| DIV | Q9Y7N6 | <i>Schizosaccharomyces pombe</i> | lysophosphatidylcholine, lysophosphatidylethanolamine, phosphatidylinositol, phosphatidylserine, lysophosphatidylethanolamine, phosphatidic acid | Purification and some properties of phospholipase B from <i>Schizosaccharomyces pombe</i> , Oishi, H.; Tsuda, S.; Watanabe, Y.; Tamai, Y. <i>Biosci. Biotechnol. Biochem.</i> 60 , 1087-1092 (1996) |
| DIV | Q9UTH5 | <i>Schizosaccharomyces pombe</i> | lysophosphatidylcholine, lysophosphatidylethanolamine, phosphatidylinositol, phosphatidylserine, lysophosphatidylethanolamine, phosphatidic acid | Purification and some properties of phospholipase B from <i>Schizosaccharomyces pombe</i> , Oishi, H.; Tsuda, S.; Watanabe, Y.; Tamai, Y. <i>Biosci. Biotechnol. Biochem.</i> 60 , 1087-1092 (1996) |

| Sub-family | AC Number | Organism | Substrate Specificity | Reference |
|------------|-----------|---|--|--|
| DIV | P78854 | <i>Schizosaccharomyces pombe</i> | lysophosphatidylcholine , lysophosphatidylethanolamine, phosphatidylinositol, phosphatidylserine, lysophosphatidylethanolamine, phosphatidic acid | Purification and some properties of phospholipase B from <i>Schizosaccharomyces pombe</i> , Oishi, H.; Tsuda, S.; Watanabe, Y.; Tamai, Y. <i>Biosci. Biotechnol. Biochem.</i> 60 , 1087-1092 (1996) |
| DV | Q9P8P2 | <i>Cryptococcus neoformans</i> var. <i>grubii</i> | 1,2-dioleoylphosphatidylcholine, 1,2-dioleoylphosphatidylethanolamine, 1,2-dioleoylphosphatidylserine, lysophosphatidylcholine , lysophosphatidylethanolamine, phosphatidylinositol, phosphatidylserine, lysophosphatidylethanolamine, phosphatidic acid | Purification and characterization of secretory phospholipase B, lysophospholipase and lysophospholipase/transacylase from a virulent strain of the pathogenic fungus <i>Cryptococcus neoformans</i> |
| DV | Q9P8L1 | <i>Cryptococcus neoformans</i> | 1,2-dioleoylphosphatidylcholine, 1,2-dioleoylphosphatidylethanolamine, 1,2-dioleoylphosphatidylserine, lysophosphatidylcholine , lysophosphatidylethanolamine, phosphatidylinositol, phosphatidylserine, lysophosphatidylethanolamine, phosphatidic acid | Purification and characterization of secretory phospholipase B, lysophospholipase and lysophospholipase/transacylase from a virulent strain of the pathogenic fungus <i>Cryptococcus neoformans</i> |
| E | P34163 | <i>Saccharomyces cerevisiae</i> | phosphatidylinositol, phosphatidylinositol 4,5-bisphosphate, 1-phosphatidyl-1D-myo-inositol 4,5-bisphosphate | Structure, function, and control of phosphoinositide-specific phospholipase C, Rebecchi, M.J.; Pentylala, S.N.; <i>Physiol. Rev.</i> 80 , 1291-1335 (2000) |
| F | P32383 | <i>Saccharomyces cerevisiae</i> (Baker's yeast) | phosphatidylinositol, phosphatidylinositol 4,5-bisphosphate, 1-phosphatidyl-1D-myo-inositol 4,5-bisphosphate | Structure, function, and control of phosphoinositide-specific phospholipase C, Rebecchi, M.J.; Pentylala, S.N.; <i>Physiol. Rev.</i> 80 , 1291-1335 (2000) |

| Sub-family | AC Number | Organism | Substrate Specificity | Reference |
|--------------|-----------|--|--|---|
| F | O13433 | <i>Candida albicans</i> | phosphatidylinositol, phosphatidylinositol 4,5-bisphosphate, 1-phosphatidyl-1D-myo-inositol 4,5-bisphosphate | Structure, function, and control of phosphoinositide-specific phospholipase C, Rebecchi, M.J.; Pentylala, S.N.; <i>Physiol. Rev.</i> 80 , 1291-1335 (2000) |
| F | P40977 | <i>Schizosaccharomyces pombe</i> | phosphatidylinositol, phosphatidylinositol 4,5-bisphosphate, 1-phosphatidyl-1D-myo-inositol 4,5-bisphosphate | Structure, function, and control of phosphoinositide-specific phospholipase C, Rebecchi, M.J.; Pentylala, S.N.; <i>Physiol. Rev.</i> 80 , 1291-1335 (2000) |
| G | P36126 | <i>Saccharomyces cerevisiae</i> (Baker's yeast) | phosphatidylcholine, phosphatidylethanolamin, phosphatidylserine | Molecular and biochemical properties and physiological roles of plant phospholipase D, Pappan, K.; Wang, X.; <i>Biochim. Biophys. Acta</i> 1439 , 151-166 (1999) |
| Other Lipase | P54857 | <i>Saccharomyces cerevisiae</i> (Baker's yeast) | tributyryn, triglyceride with short chain fatty acids | Molecular and biochemical properties and physiological roles of plant phospholipase D, Pappan, K.; Wang, X.; <i>Biochim. Biophys. Acta</i> 1439 , 151-166 (1999) |
| Other Lipase | P41365 | <i>Candida antarctica</i> (Yeast) (<i>Trichosporon oryzae</i>) | Tributyryn | Production, purification, characterization, and applications of lipases, Sharma, R.; Chisti, Y.; Banerjee, U.C.; <i>Biotechnol. Adv.</i> 19 , 627-662 (2001) |